



US 20080233627A1

(19) **United States**(12) **Patent Application Publication**

Mori et al.

(10) **Pub. No.: US 2008/0233627 A1**(43) **Pub. Date: Sep. 25, 2008**(54) **NICOTIANAMINE SYNTHASE AND GENE ENCODING THE SAME**(75) Inventors: **Satoshi Mori**, Chiba-ken (JP); **Kyoko Higuchi**, Gunma (JP); **Kazuya Suzuki**, Tokyo (JP); **Naoko Nishizawa**, Tokyo (JP); **Hiromi Nakanishi**, Tokyo (JP)

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BOSTON, MA 02205 (US)(73) Assignee: **Japan Science and Technology Corporation**, Saitama (JP)(21) Appl. No.: **11/702,690**(22) Filed: **Feb. 5, 2007****Related U.S. Application Data**

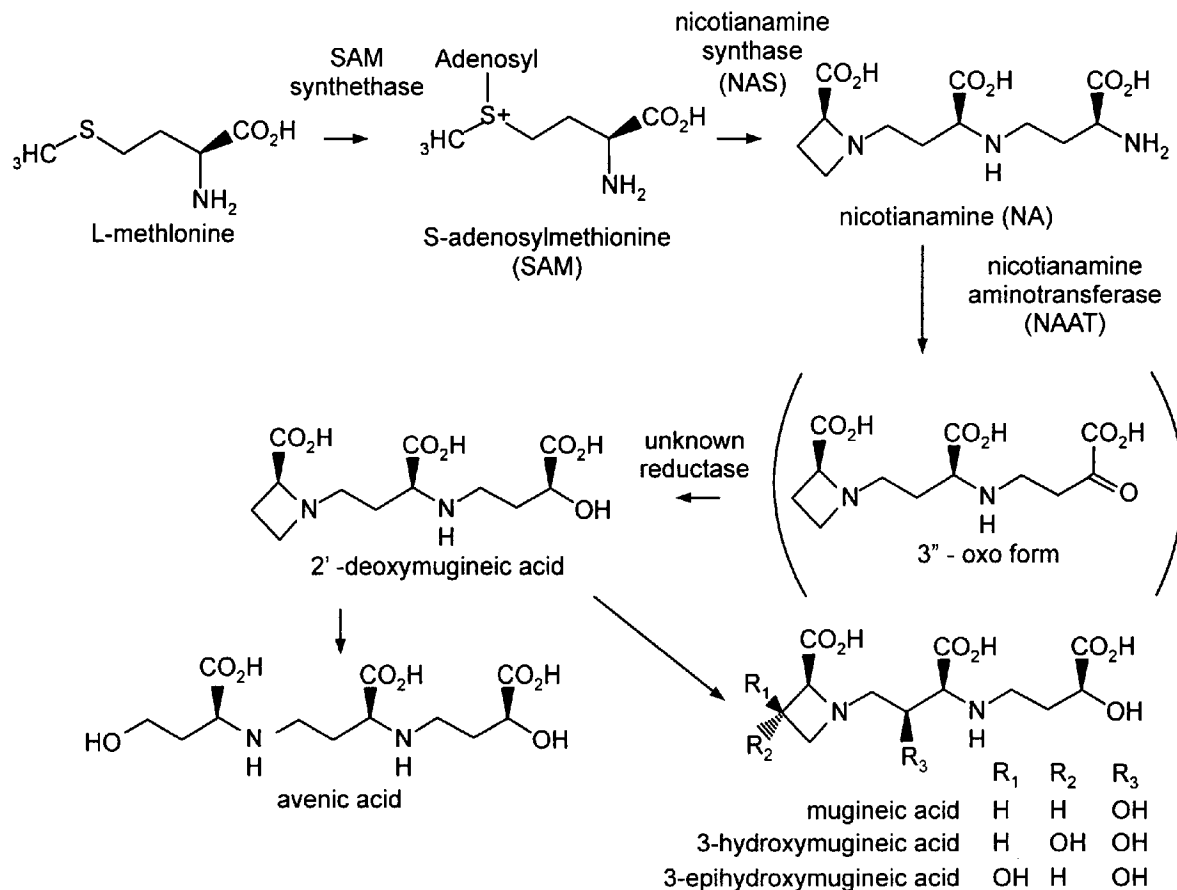
(62) Division of application No. 09/674,337, filed on Jul. 26, 2001, now Pat. No. 7,192,755, filed as application No. PCT/JP99/02305 on Apr. 30, 1999.

(30) **Foreign Application Priority Data**

Apr. 30, 1998 (JP) 10-137685/1988

Publication Classification(51) **Int. Cl.**
C12N 9/88 (2006.01)(52) **U.S. Cl.** **435/232**(57) **ABSTRACT**

A nicotianamine synthase is isolated and purified. Then the gene of this enzyme is cloned and the base sequence and amino acid sequence thereof are determined. This gene is employed in constructing plants, in particular, grass plants highly tolerant to iron-deficiency. A nicotianamine synthase involved in the mugineic acid biosynthesis pathway; the amino acid sequence thereof; a gene encoding the same; a vector containing this gene; cells transformed by the vector; a process for producing nicotianamine by using the same; plants transformed by the gene encoding the nicotianamine synthase; and an antibody against the nicotianamine synthase.



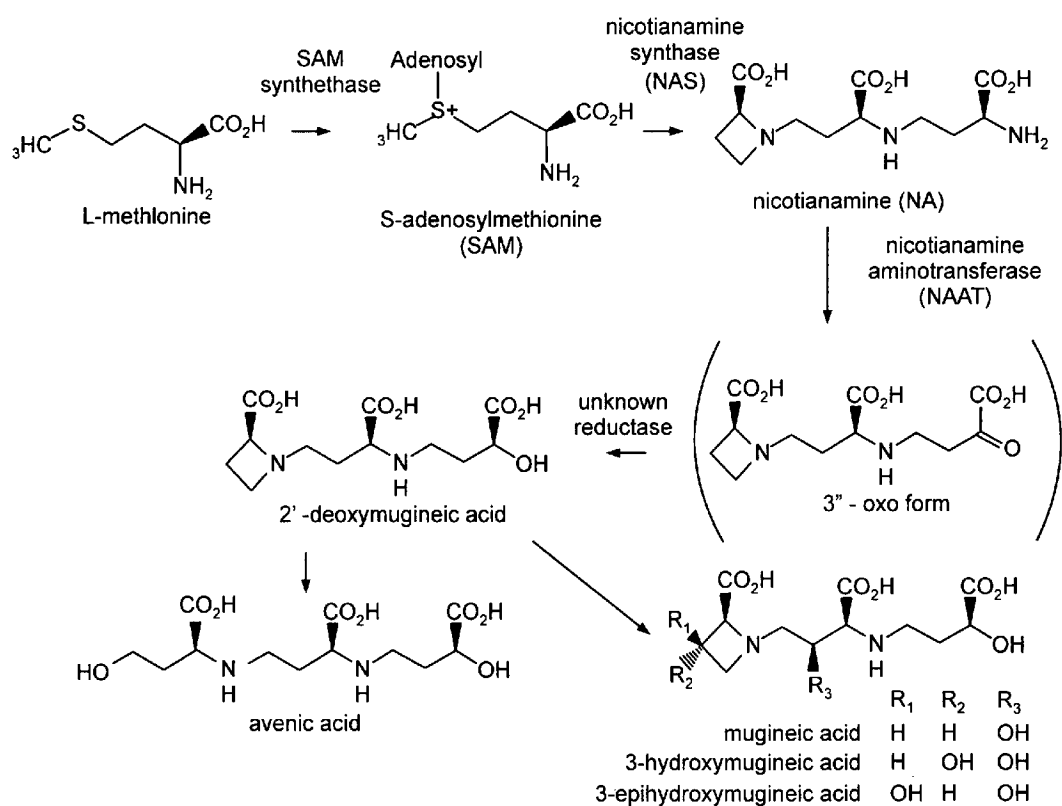


FIG. 1

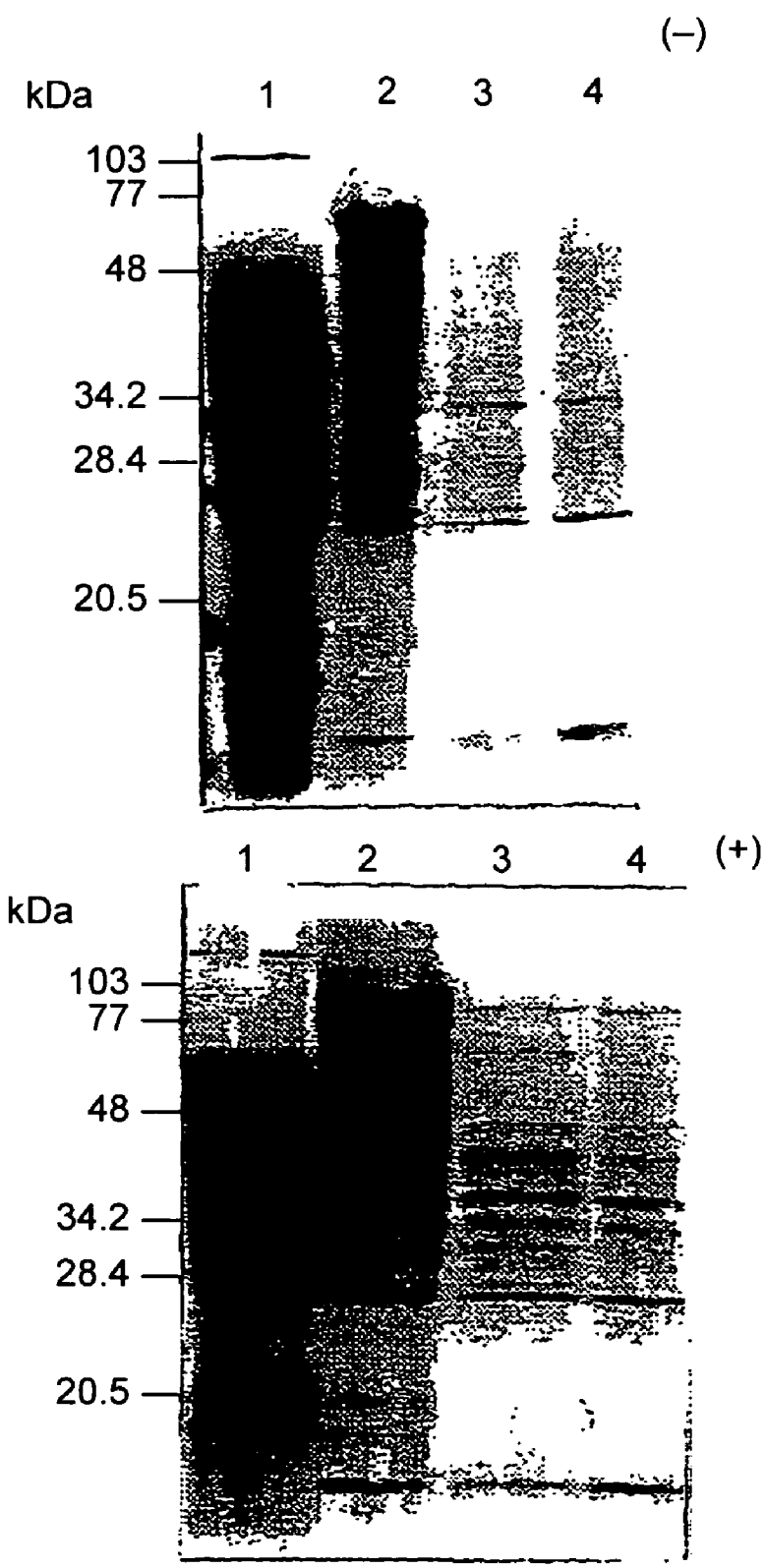


FIG. 2

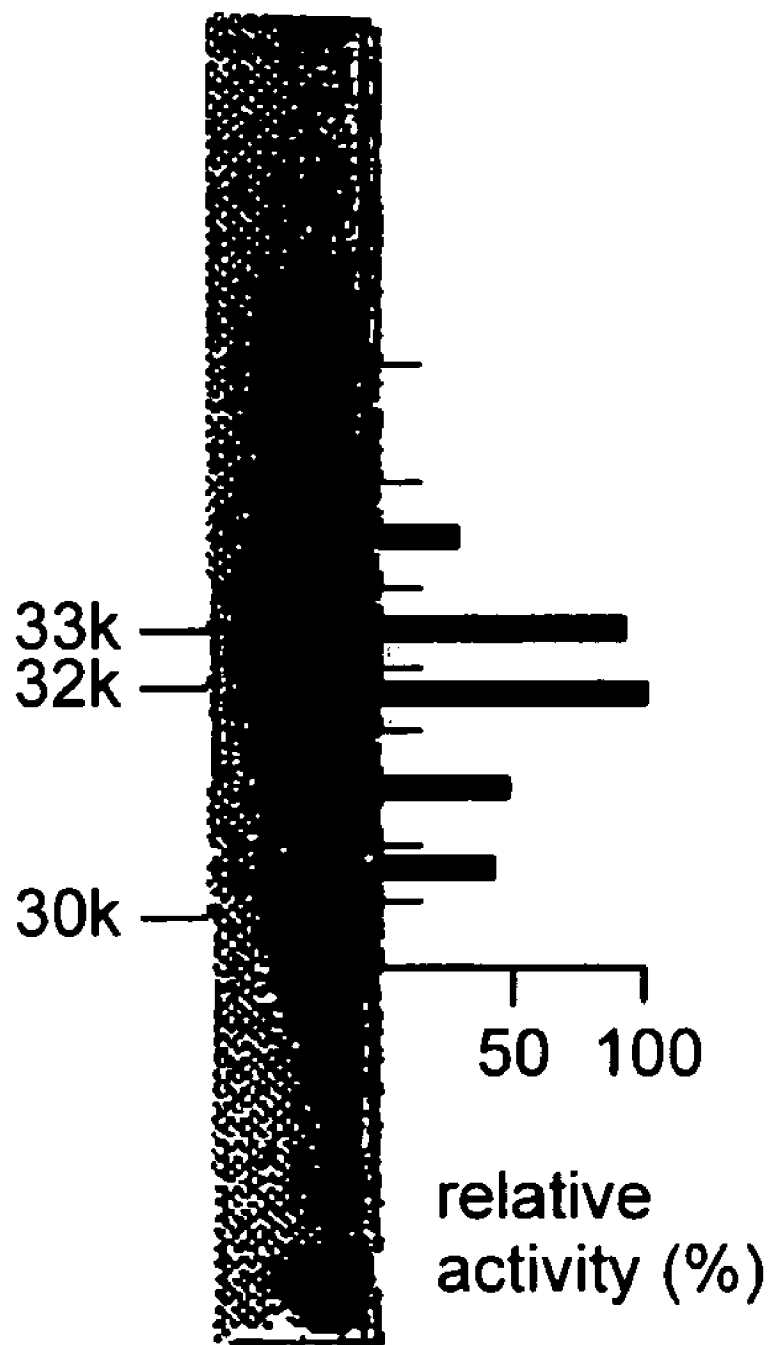


FIG. 3

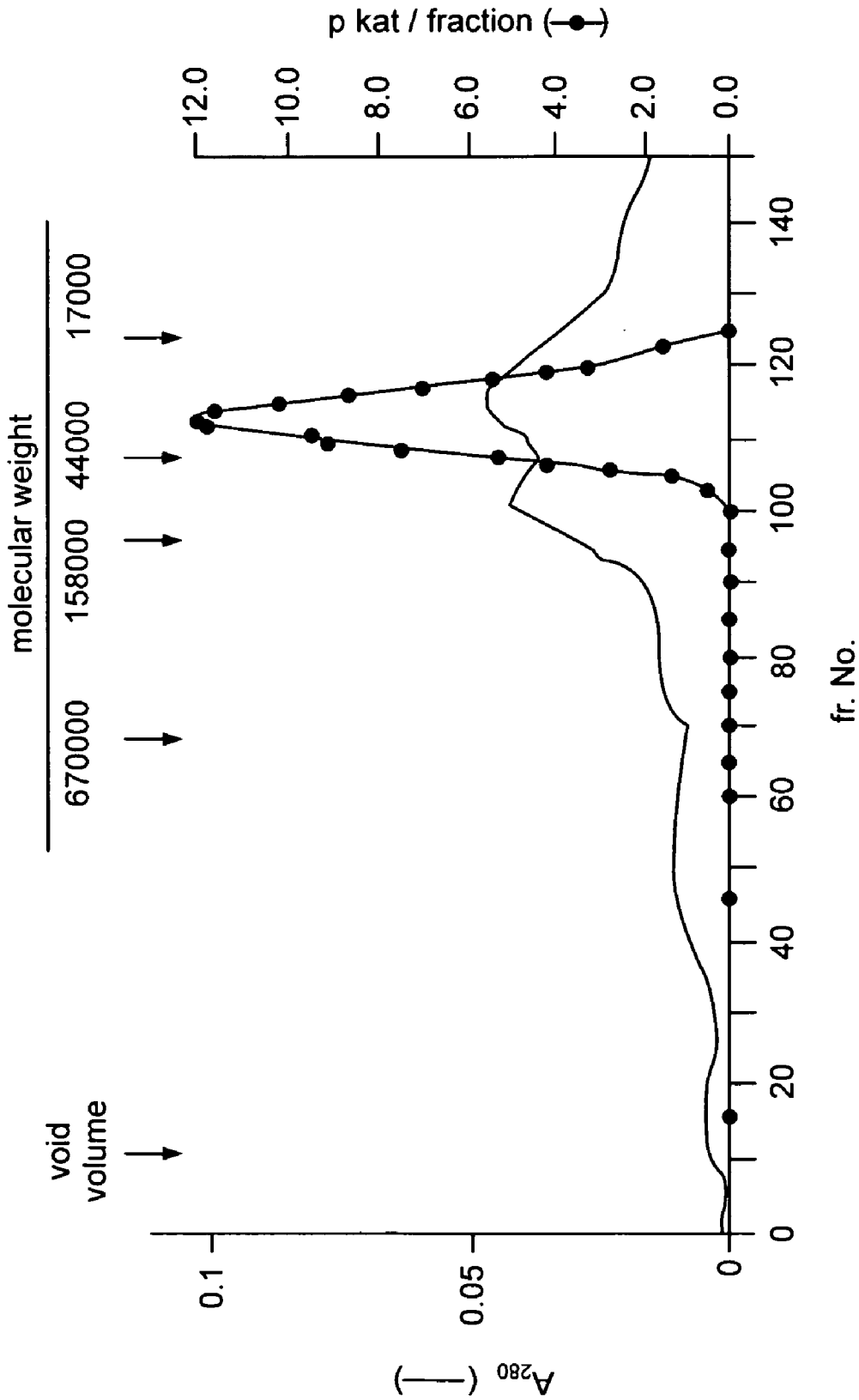


FIG. 4

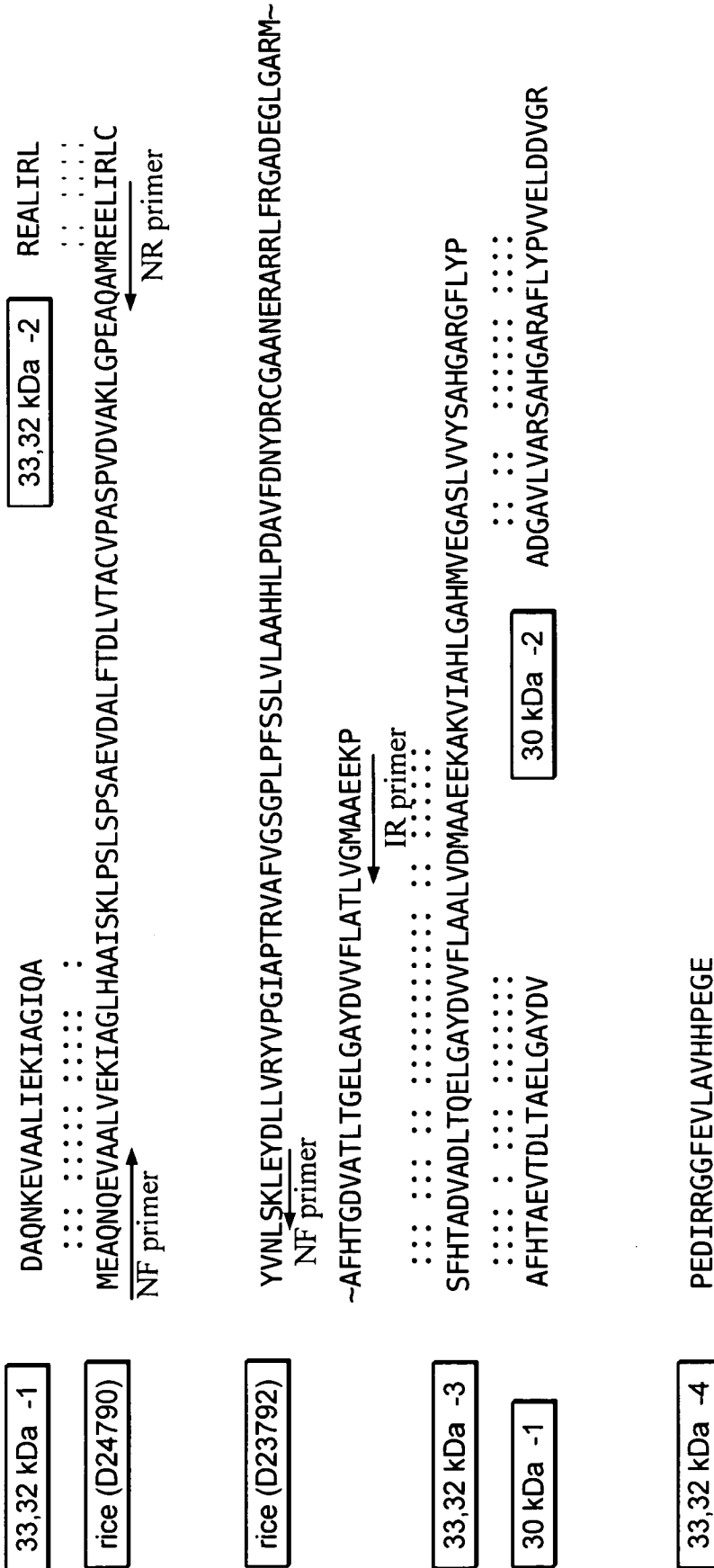


FIG. 5

	GCG TTC AGA GGC TTC CAG AGT TCT TCC GGT CAC CAA GAA GCA TTT GAT CAT AAC	54
	ATG GAT GCC CAG AAC AAG GAG GTC GCT GCT CTG ATC GAG AAG ATC GCC GGT ATC	108
19	<u>M^① D A Q N K E V A A L I E K I A G I</u>	
	CAG GCC GCC ATC GCC GAG CTG CCG TCG CTG AGC CCG TCC CCC GAG GTC GAC AGG	162
37	<u>Q A A I A E L P S L S P S P E V D R</u>	
	CTC TTC ACC GAC CTC GTC ACG GCC TGC GTC CCG CCG AGC CCC GTC GAC GTG ACG	216
55	<u>L F T D L V T A C V P P S P V D V T</u>	
	AAG CTC AGC CCG GAG CAC CAG AGG ATG CGG GAG GCT CTC ATC CGC TTG GTG TCC	270
73	<u>K L S P E H Q R M^② R E A L I R L C S</u>	
	GCC GCC GAG GGG AAG CTC GAG GCG CAC TAC GCC GAC CTG CTC GCC ACC TTC GAC	324
91	<u>A A E G K L E A H Y A D L L A T F D</u>	
	AAC CCG CTC GAC CAC CTC GGC CTC TTC CCG TAC TAC AGC AAC TAC GTC AAC CTC	378
109	<u>N P L D H L G L F P Y Y S N Y V N L</u>	
	AGC AGG CTG GAG TAC GAG CTC CTG GCG CGC CAC GTG CCG GGC ATC GCG CCG GCG	432
127	<u>S R L E Y E L L A R H V P G I A P A</u>	
	CGC GTC GCC TTC GTC GGC TCC GGC CCG CTG CCG TTC AGC TCG CTC GTC CTC GCC	486
145	<u>R V A F V G S G P L P F S S L V L A</u>	
	GCG CAC CAC CTG CCC GAG ACC CAG TTC GAC AAC TAC GAC CTG TGC GGC GCG GCC	540
163	<u>A H H L P E T Q F D N Y D L C G A A</u>	
	AAC GAG CGC GCC AGG AAG CTG TTC GGC GCG ACG GCG GAC GGC GTC GGC GCG CGT	594
181	<u>N E R A R K L F G A T A D G V G A R</u>	
	ATG TCG TTC CAC ACG GCG GAC GTC GCC GAC CTC ACC CAG GAG CTC GGC GCC TAC	648
199	<u>M^③ S F H T A D V A D L T Q E L G A Y</u>	
	GAC GTG GTC TTC CTC GCC GCG CTC GTC GGC ATG GCA GCC GAG GAG AAG GCC AAG	702
217	<u>D V V F L A A L V G M A A E E K A K</u>	
	GTG ATT GCC CAC CTG GGC GCG CAC ATG GTG GAG GGG GCG TCC CTG GTC GTG CGG	756
235	<u>V I A H L G A H M V E G A S L V V R</u>	
	AGC GCA CGG CCC CGC GGC TTT CTT TAC CCC ATT GTC GAC CCG GAG GAC ATC AGG	810
253	<u>S A R P R G F L Y P I V D^④ P E D I R</u>	
	CGG GGT GGG TTC GAG GTG CTG GCC GTG CAC CAC CCG GAA GGT GAG GTG ATC AAC	864
271	<u>R G G F E V L A V H H P E G E V I N</u>	
	TCT GTC ATC GTC GCC CGT AAG GCC GTC GAA GCG CAG CTC AGT GGG CCG CAG AAC	918
289	<u>S V I V A R K A V E A Q L S G P Q N</u>	
	GGA GAC GCG CAC GCA CGG GGC GCG GTG CCG TTG GTC AGC CCG CCA TGC AAC TTC	972
307	<u>G D A H A R G A V P L V S P P C N F</u>	
	TCC ACC AAG ATG GAG GCG AGC GCG CTT GAG AAG AGC GAG GAG CTG ACC GCC AAA	1026
325	<u>S T K M E A S A L E K S E E L T A K</u>	
	GAG CTG GCC TTT TGA TTG AAG AGT GCG CGT GGT CAT TCT GTC GCC TGC GAT CGT	1080
	E L A F *	
	GGT AAC TTT CCT ACT CGT GTG TGT TTT GAT GTT TGT GCC TGT AAG AGT TAT GCT	1134
	TCC GGC CTT GTG CTG TTA ATT TAC ACG CGT TAC ATG TAG TAC TTG TAT TTA TAC	1188
	CTG GAA TAA CCG TAT GTA ACA TAA ATA TTA GTG GGA TTT GAA GTG TAA TGC TAA	1242
	ATA ATA ATA AAA CTT GAT GCA GAC ATT CAA AAA AAA AAA AAA AAA AAA AAA AA	

FIG. 6

FIG. 7A
FIG. 7B

FIG. 7

FIG. 7A

HvNAS4	MDGQSE - - EVDALVQK I TGLHAA I AKLPSLSPSPDVDALFTDLVTACVPPSPVDVTKLAP
HvNAS7	MDAQSK - - EVDALVQK I TGLHAA I AKLPSLSPSPDVDALFTDLVTACVPPSPVDVTKLAP
HvNAS6	MDAQNK - - EVDALVQK I TGLHAA I AKLPSLSPSPDVDALFTDLVTACVPPSPVDVTKLGS
HvNAS2	MAAQNN - QEVDALVEK I TGLHAA I AKLPSLSPSPDVDALFTELVTACVPPSPVDVTKLGP
HvNAS3	MAAQNN K DVAALVEK I TGLHAA I AKLPSLSPSPDVDALFTELVTACVPPSPVDVTKLGP
HvNAS1	MDAQNK - - EVAALI EK I AG IQAA I AELPSLSPSPEVDRFLTDLVTACVPPSPVDVTKLSP
NvNAS5	MEAENG - - EVAALVEK I TGLHAA I SKLPALSPSPQVDALFTELVAACVPSPPVDVTKLGP
	* * * * *
HvNAS4	EAQAMREGLIRLCSAEAGKLEAHYSDMLAAFDNPLDHLGVFPYYSNY I NLSKLEYE LLAR
HvNAS7	EAQAMREGLIRLCSAEAGKLEAHYSDMLAAFDNPLDHLGVFPYYSNY I NLSKLEYE LLAR
HvNAS6	EAQEMREGLIRLCSAEAGKLEAHYSDMLAAFDNPLDHLGMFPYYSNY I NLSKLEYE LLAR
HvNAS2	EAQEMREGLIRLCSAEAGKLEAHYSDMLAAFDKPLDHLGMFPYYSNY I NLSKLEYE LLAR
HvNAS3	EAQEMREGLIRLCSAEAGKLEAHYSDMLAAFDNPLDHLGI FPYYSNY I NLSKLEYE LLAR
HvNAS1	EHQRMREALIRLCSAEAGKLEAHYADLL AT FDNPLDHLGLFPYYSNYVNLRSLEYE LLAR
NvNAS5	EAQEMRQDLIRLCSAEAGLL EAHYSDML TA LDSPLDHLGRFPYFDNYVNLSTREHDE LLAG
	* * * * *
HvNAS4	YVGRHRPAP VAF I GSGPLPFSSYVLAAR HLPD TVFDNYDLCGAANDRAT RLF FRAD KD - V
HvNAS7	YVPG GI APAP VAF I GSGPLPFSSYVLAAR HLPD TVFDNYVPVRAANDRAT RLF FRAD KD - V
HvNAS6	YVPG GI ARPA VAF I GSGPLPFSSYVLAAR HLPD AMFDNYDLCGAANDRAS KLF FRAD KD - V
HvNAS2	YVPGGYRPAR VAF I GSGPLPFSSYVLAAR HLPD TMFDNYDLCGAANDRAS KLF FRAD RD - V
HvNAS3	VVRR - HRPAR VAF I GSGPLPFSSYVLAAR HLPD TMFDNYDLCGAANDRAS KLF FRAD TD - V
HvNAS1	HVPG - I APAR VAF V GSGPLPFSSYVLAAR HLPETQFDNYDLCGAANERAR KLF FGAT ADGV
NvNAS5	HVAA - - - PAR VAF I GSGPLPFSSYVLAAR HLPD TRFDNYDRCS VANGRAMKLVVGAADGV
	* * * * *

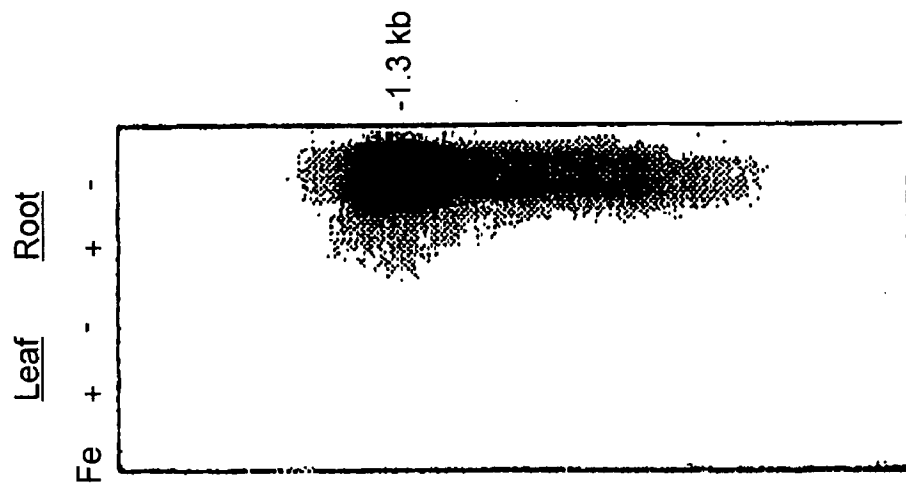


FIG. 9

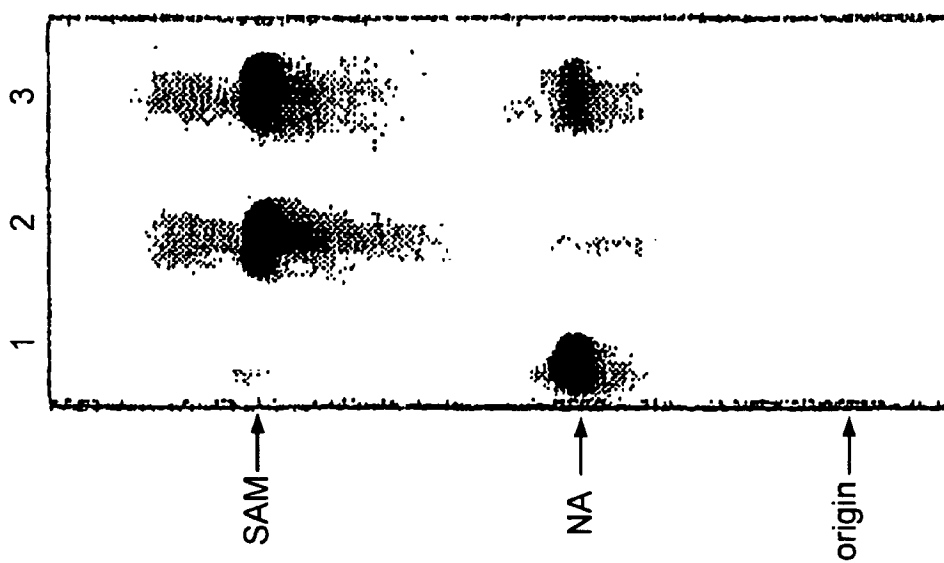


FIG. 8

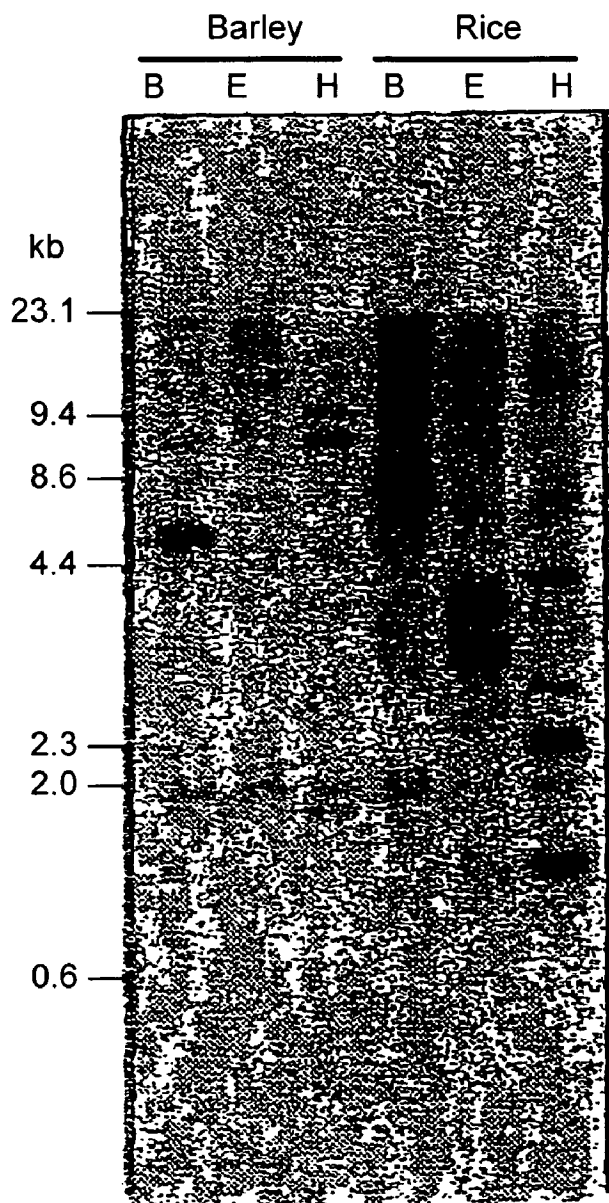


FIG. 10

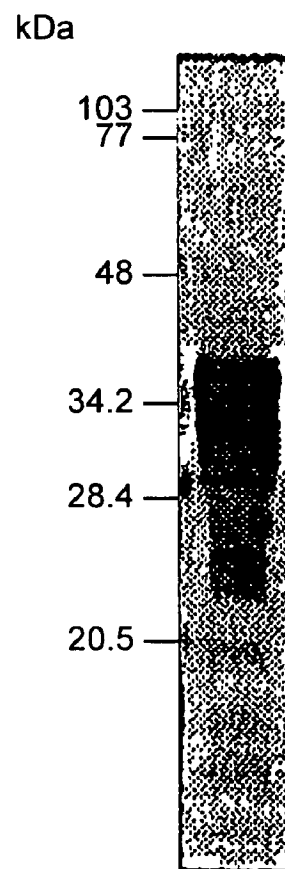


FIG. 11

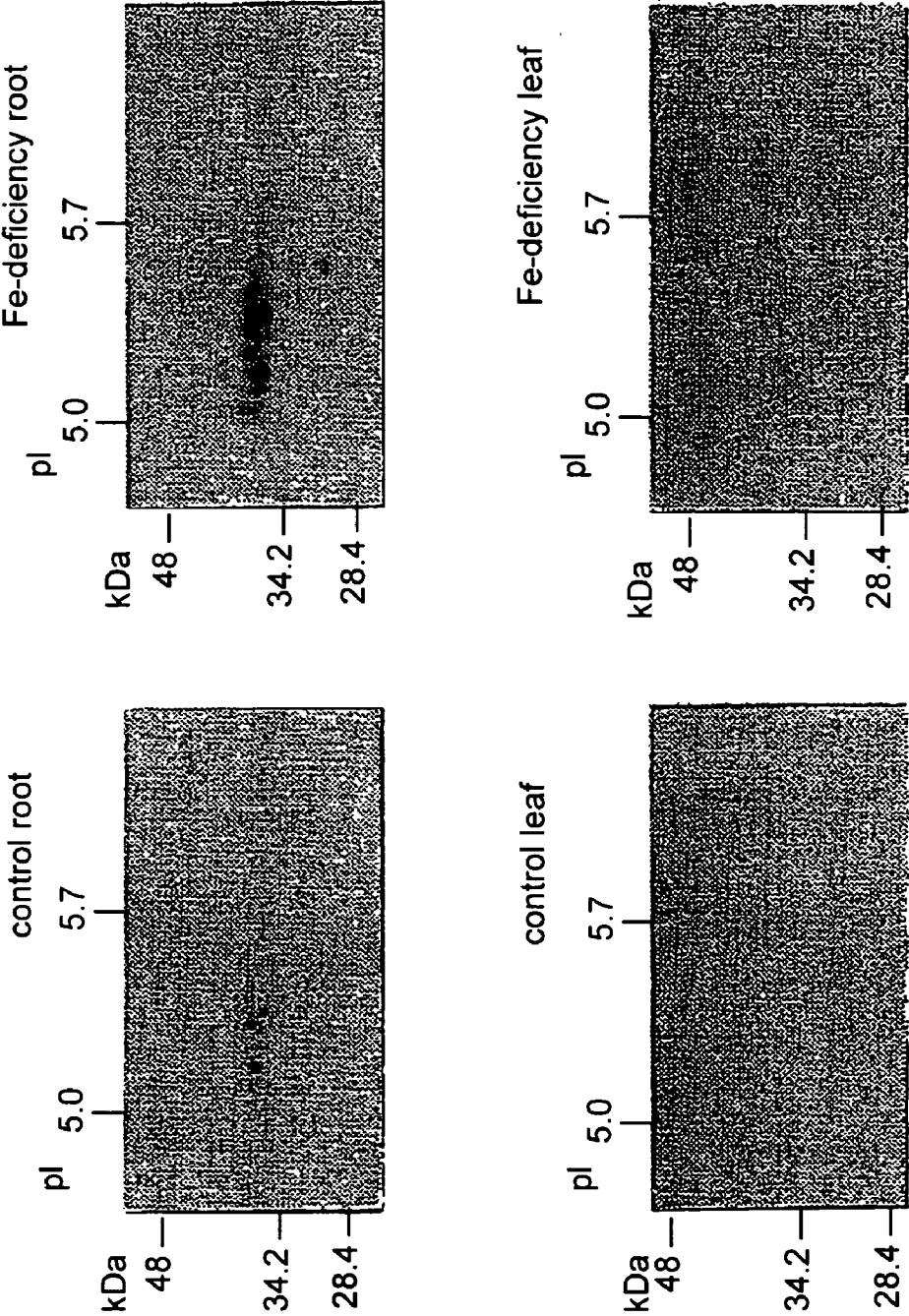


FIG. 12

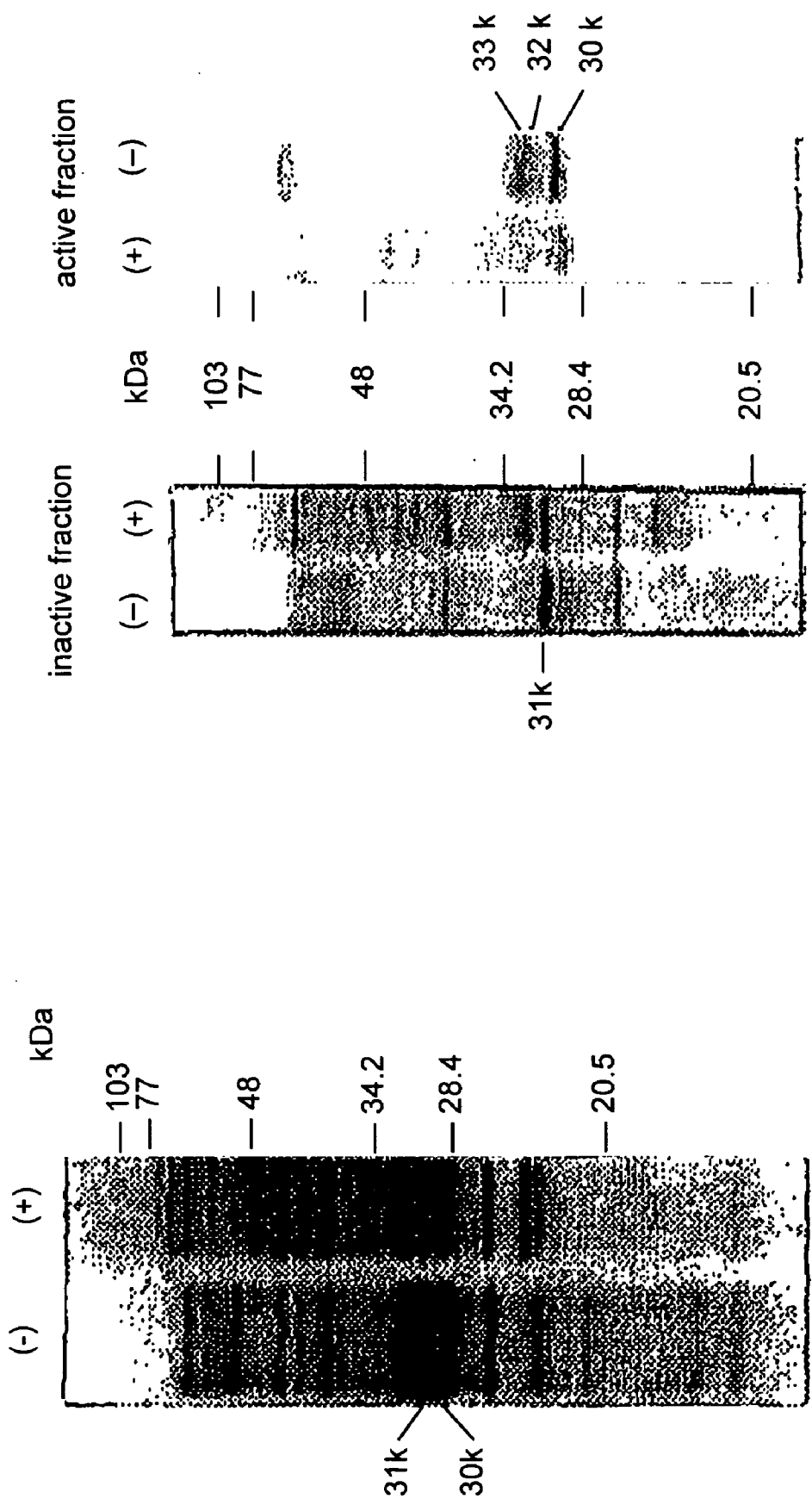


FIG. 14

FIG. 13

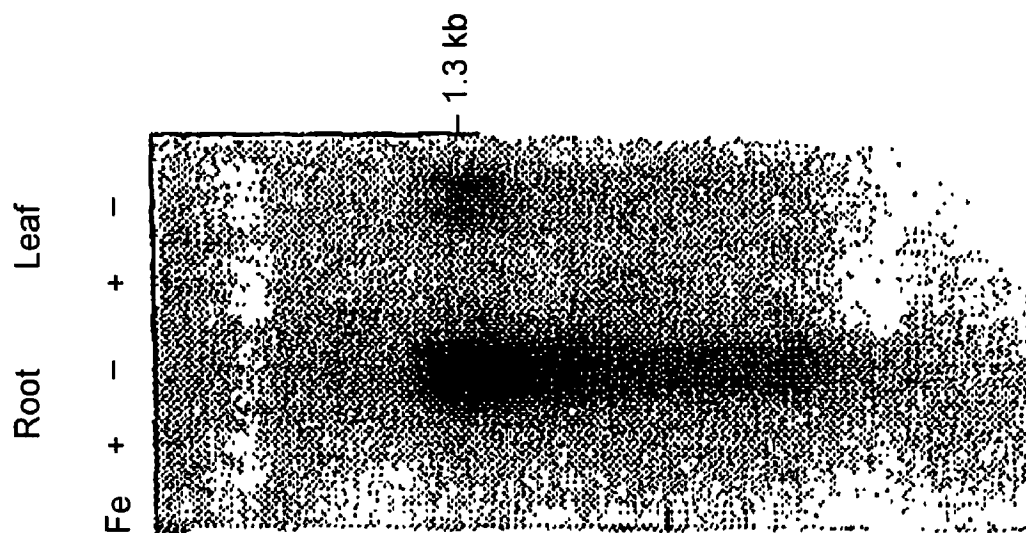


FIG. 16

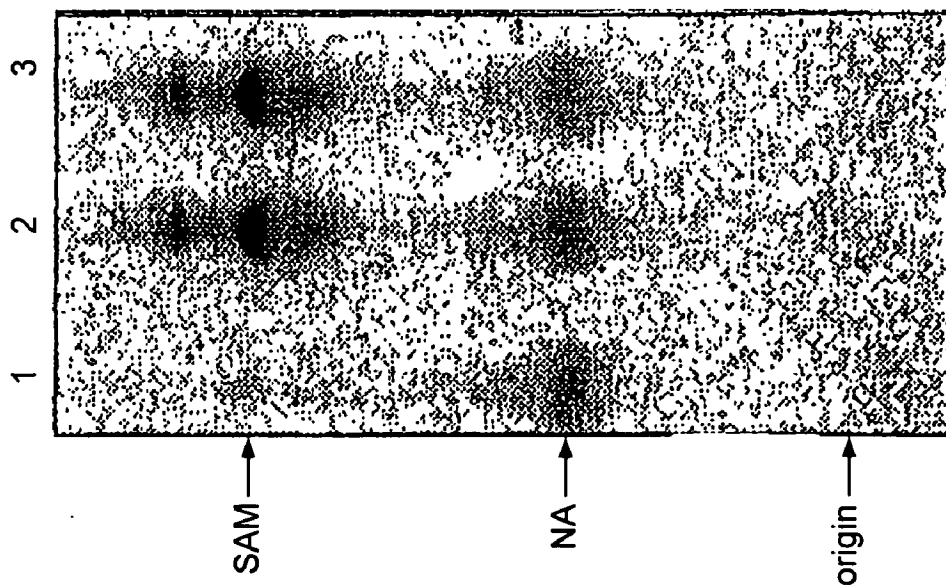


FIG. 15

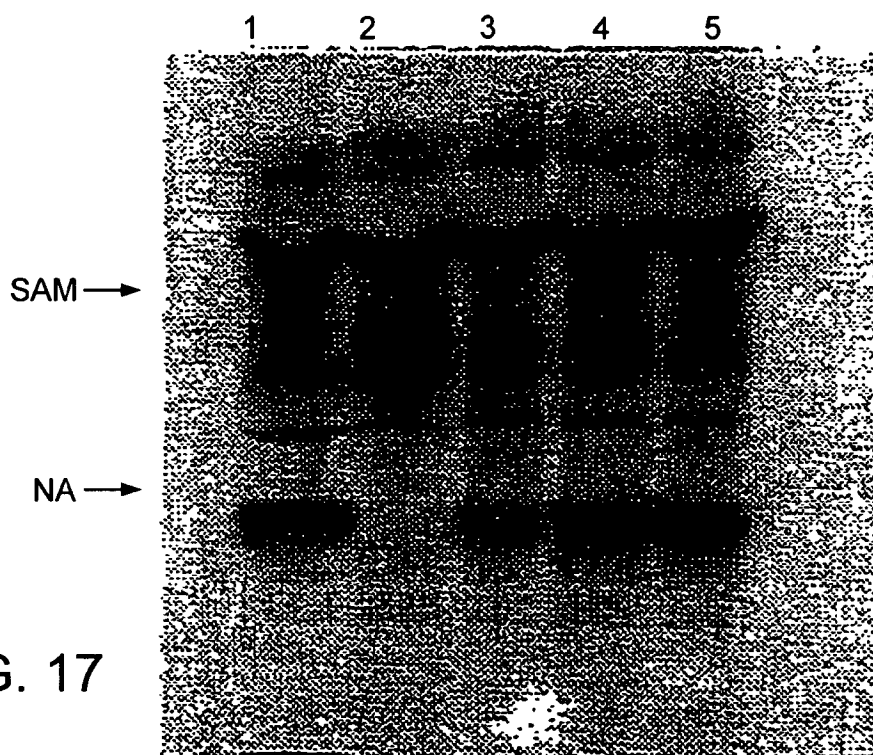


FIG. 17

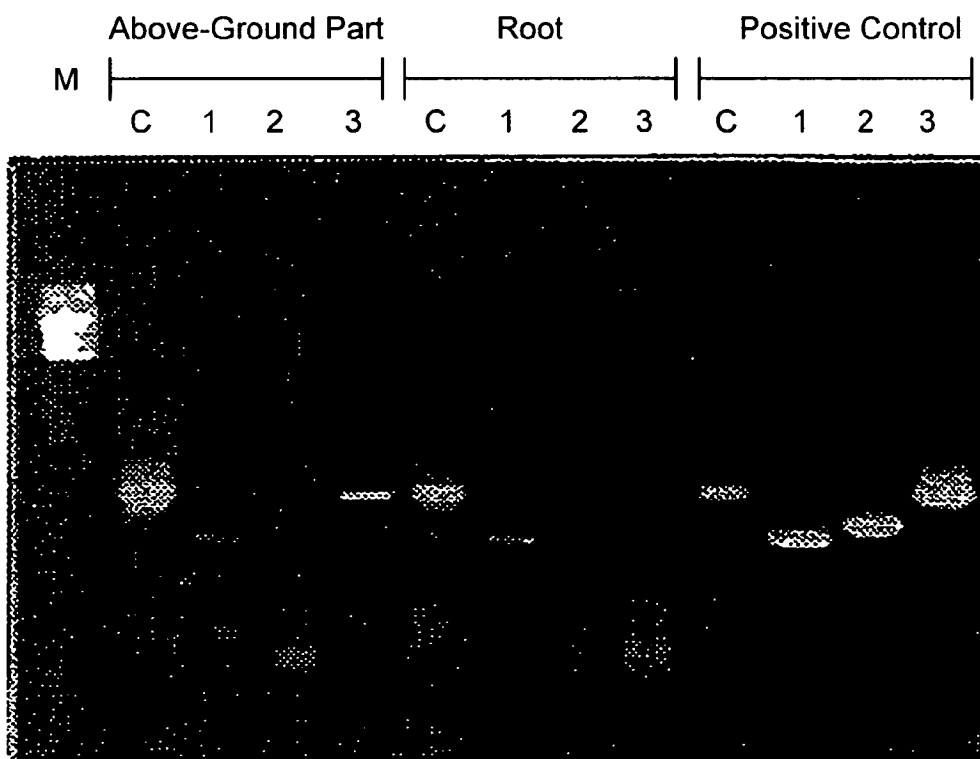


FIG. 18

NICOTIANAMINE SYNTHASE AND GENE ENCODING THE SAME

TECHNICAL FIELD

[0001] The present invention relates to a nicotianamine synthase involved in the mugineic acid biosynthetic pathway, the amino acid sequence thereof, a gene encoding the same, a vector, a process for producing nicotianamine by using the same, plants transformed by the gene encoding the nicotianamine synthase, and an antibody against the nicotianamine synthase.

BACKGROUND ART

[0002] Gramineous plants that absorb by chelating the insoluble state Fe(III) in soil using mugineic acid and adopt so called the Strategy-II mechanism of Fe acquisition secrete Fe chelators (phytosiderophores) from their roots to solubilize sparingly soluble Fe in the rhizosphere (Roemheld, 1987). The amount of the secreted phytosiderophores increases under Fe-deficiency stress. The mugineic acid family is the only examples of phytosiderophores known so far (Takagi, 1976). Tolerance to Fe deficiency in gramineous plants is thought to depend on a quantity of mugineic acid family secreted by plants (Takagi et al. 1984, Roemheld and Marschner 1986, Marschner et al. 1987, Mori et al. 1987, Kawai et al. 1988, Mori et al. 1988, Mihashi and Mori 1989, and Shingh et al. 1993).

[0003] The biosynthetic pathway of mugineic acid in plants is shown in FIG. 1. S-adenosylmethionine is synthesized from methionine by S-adenosylmethionine synthase. Subsequently, three molecules of S-adenosylmethionine are combined to form one molecule of nicotianamine by nicotianamine synthase. The generated nicotianamine is then converted to 3"-keto acid by nicotianamine aminotransferase, and 2'-deoxymugineic acid is synthesized by the subsequent action of a reductase. A further series of hydroxylation steps produces the other mugineic acid derivatives including mugineic acid from the deoxymugineic acid (Mori and Nishizawa 1987, Shojima et al. 1989, Shojima et al. 1990 and Ma and Nomoto 1993).

[0004] A compound in FIG. 1, a compound in the lower right, wherein R₁ and R₂ are hydrogen and R₃ is hydroxyl, is mugineic acid. A compound wherein R₁ is hydrogen and R₂ and R₃ are hydroxyl, is 3-hydroxymugineic acid. Also a compound wherein R₂ is hydrogen and R₁ and R₃ are hydroxyl, is 3-epihydroxymugineic acid.

[0005] Three S-adenosylmethionine synthase genes were isolated from barley roots, but these genes were not induced by Fe deficiency (Takizawa et al. 1996). A gene *Ids3*, which is obtained from the barley by differential screening, is suspected to be a gene, which converts deoxymugineic acid to mugineic acid by hydroxylation and is strongly induced by Fe-deficiency (Nakanishi et al. 1993). Further, nicotianamine aminotransferase was purified and isolated from Fe-deficient barley roots, and two nicotianamine aminotransferase genes, Naat-A and Naat-B, were isolated (Takahashi et al. 1997). Naat-A expression was induced in Fe-deficient roots.

[0006] The synthesis of nicotianamine from S-adenosylmethionine is similar to polyamine synthesis from decarboxy-S-adenosylmethionine. In contrast to polyamine synthase, however, nicotianamine synthase catalyzes the combination of three S-adenosylmethionine molecules and the azetidine ring formation at the same time (FIG. 1). Such

the nicotianamine synthase is a novel type of enzyme. Previously, we reported the partial purification of nicotianamine synthase from the roots of Fe-deficient barley and expression pattern of the activity (Higuchi et al. 1994, Higuchi et al. 1995, Kanazawa et al. 1995, Higuchi et al. 1996a and Higuchi et al. 1996b). Since nicotianamine synthase is easily decomposed during extraction and purification, it has been difficult to purify sufficient quantities for amino acid sequencing.

[0007] The present invention has an object to provide a plant, especially gramineous plant, highly tolerant to Fe-deficiency, as a result of isolating and purifying a nicotianamine synthase, being cloned the gene of this enzyme, determining the base sequence and amino acid sequence thereof, and using said enzyme.

DISCLOSURE OF INVENTION

[0008] The present invention relates to a nicotianamine synthase shown in SEQ ID NO: 1 comprising amino acid sequence shown in SEQ ID NO: 1, or amino acid sequence having deletion in a part thereof, being substituted by the other amino acids or being added with the other amino acids.

[0009] The present invention relates to the gene encoding said amino acid sequence of nicotianamine synthase.

[0010] The present invention also relates to a vector comprising containing said gene, and a transformant transformed by the said vector.

[0011] The present invention relates to a process for production of nicotianamine using the said transformant.

[0012] The present invention further relates to plants, especially gramineous plants, to which said gene is introduced, and fruits obtained by growing said plants.

[0013] The present invention relates to a process for extraction of said nicotianamine synthase in the presence of thiol protease inhibitor, preferably E-64.

[0014] Further, the present invention relates to an antibody against said nicotianamine synthase.

BRIEF DESCRIPTION OF DRAWING

[0015] FIG. 1 shows the biosynthetic pathway of mugineic acid family.

[0016] FIG. 2 shows a comparison of nicotianamine synthase purification from Fe-dependent and control barley roots.

[0017] FIG. 3 shows a preparative SDS-PAGE (sodium dodecyl sulfate-polyacrylamide gel electrophoresis, hereinafter designates as SDS-PAGE) around 30-35 kDa. The horizontal bar indicates relative enzyme activity detected from the gels.

[0018] FIG. 4 shows elution pattern of nicotianamine synthase activity from the gel-filtration column.

[0019] The large closed circles (●) indicates enzyme activity.

[0020] FIG. 5 shows a comparison with a six partial amino acid sequence determined by nicotianamine synthase originated from barley and similar sequence of gramineous plants obtained by computer search of the database. Identical amino acid residue is shown in a ":".

[0021] FIG. 6 shows full length of HvNAS1 cDNA and amino acid sequence deduced therefrom. The underlined sequences indicate the identical partial amino acid sequences of fragments in the above FIG. 5. Numbers of the nucleotide sequence are indicated to the right of each row. Amino acid numbers are indicated on the left of each row.

[0022] FIG. 7 shows comparison of the deduced amino acid sequences of the above 7 cDNA obtained from barley. Asterisks "*" indicates identical amino acid residues in all sequences.

[0023] FIG. 8 shows results of thin layer chromatographic (TLC) analysis of nicotianamine synthase activity obtained from *E. coli* crude extract expressing a fused protein of maltose binding protein—HvNAS1.

[0024] FIG. 9 shows Northern-hybridization analysis of HvNAS1 as a probe.

[0025] FIG. 10 shows Southern-hybridization analysis of HvNAS1 as a probe.

[0026] FIG. 11 shows Western-blot analysis of crude enzyme used for detection of nicotianamine synthase activity.

[0027] FIG. 12 shows Western-blot analysis of total protein extracted by trichloroacetic acid/acetone.

[0028] FIG. 13 shows comparison of nicotianamine synthase purification from Fe-deficient barley and control barley after DEAE-Sepharose FF.

[0029] FIG. 14 shows comparison of nicotianamine synthase purification from Fe-deficient barley and control barley after Ether Toyopearl 650M.

[0030] FIG. 15 shows results of thin layer chromatographic (TLC) analysis of nicotianamine synthase activity obtained from *E. coli* crude extract expressing a fused protein of maltose binding protein-OsNAS1.

[0031] FIG. 16 shows Northern-hybridization analysis of OsNAS1 as a probe.

[0032] FIG. 17 shows results of thin layer chromatographic (TLC) analysis of nicotianamine synthase activity obtained from *E. coli* crude extract expressing a fused proteins of maltose binding protein-AtNAS1, AtNAS2 or AtNAS3.

[0033] FIG. 18 shows results of RT-PCR of total RNA extracted from the aboveground parts and roots of *Arabidopsis thaliana*. Right group indicates positive control.

BEST MODE FOR CARRYING OUT THE INVENTION

[0034] We have tried to isolate nicotianamine synthase (Higuchi et al. *Plant & Soil*, Vol. 165, p. 173-179, 1994), and since nicotianamine synthase was easily decomposed and was difficult to isolate and purify, we were unable to obtain sufficient amounts of protein to determine its partial amino acid sequence. Subsequently, it was found that a thiol protease inhibitor E-64 (hereinafter designates as E-64) was very effective in suppressing degradation of nicotianamine synthase (Higuchi et al. *Plant & Soil*, Vol. 178, p. 171-177, 1996 a).

[0035] In the present invention, as a result that frozen roots were crushed to a fine powder in liquid N₂ and then rapidly homogenized with buffer containing 0.1 mM thiol protease inhibitor E-64, nicotianamine synthase protein could be isolated and its gene could also be isolated.

[0036] Further, the enzyme of the present invention recovered its activity by removal of SDS after SDS-PAGE treatment, but the rate of recovery was very low (Higuchi et al. *Plant & Soil*, Vol. 165, p. 173-179, 1994). Consequently, degree of purification should be increased up before treatment of SDS-PAGE. Then the column chromatography procedures were further improved.

[0037] We have also found that the enzyme of the present invention is relatively hydrophobic and a buffer containing a mild surface active agent CHAPS increased the rate of recovery. Several ion-exchange chromatography carriers were

tested, and DEAE-Sepharose FF and DEAE Sephacel were found to be the most effective. In addition to TSK gel Butyl Toyopearl, another hydrophobic chromatography carrier, TSK gel Ether Toyopearl 650M, effectively removed impurities of the 30-35 kDa.

[0038] The enzyme of the present invention has been reported that it was the peptide of 30-35 kDa, the activity of which was recovered by removing SDS after SDS-PAGE treatment, and the activity was detected as a broad molecular weight range of 30-35 kDa (refer to FIG. 3). FIG. 3 shows a result of preparative SDS-PAGE in the fractions showing enzyme activity. SDS-PAGE was carried out using 11% acrylamide slab gels. A portion of the gel was stained with Coomassie brilliant blue and the rest of the gel was stained with Cu. The gel containing proteins between 30-35 kDa in size was cut into seven fragments (indicated by the short lines). The thick bars in FIG. 3 indicate relative enzymatic activities detected from each gel fragment.

[0039] In order to identify nicotianamine synthase peptide from the proteins having these molecular weights, the peptides, which were contained in the nicotianamine synthase fractions, purified from Fe-deficient and control barley roots were compared using SDS-PAGE. From each barley root 200 g, the present enzyme was purified according to the method described in example 3 hereinbelow.

[0040] The enzyme activity of the control was a quarter of the Fe-deficient roots.

[0041] The peptide composition of the active enzyme fraction from each purification step of the present enzyme was analyzed and compared by SDS-PAGE, and results are shown in FIG. 2, FIG. 13 and FIG. 14. FIG. 2, FIG. 13 and FIG. 14 show comparison with the active fraction from the purification step of Fe-deficient barley roots 200 g [in the figure, shown with (-)], and the active fraction from the purification step of the control barley roots 200 g [in the figure, shown with (+)]. SDS-PAGE was carried out using 12.5% acrylamide slab gels (Laemmli, *Nature* Vol. 227, p. 680-685, 1970). Gels were stained with Coomassie brilliant blue. FIG. 2 shows a step before DEAE-Sepharose. The upper row shows enzyme from Fe-deficient barley roots and the lower row shows enzyme from control roots. In each lane, lanes 1, crude extract, 200#g of protein; lanes 2, after Butyl Toyopearl 650M, 100 µg of protein; lanes 3, after hydroxyapatite, 20 µg of protein; and lanes 4, after Butyl Toyopearl 650M, 15 µg of protein, are shown.

[0042] FIG. 13 shows after DEAE-Sepharose FF, each lane, 25 µg of protein. FIG. 14 shows after Ether Toyopearl 650M; in which left shows inactive fraction, and right shows active fraction, and 1/25 of each fraction is electrophoresed.

[0043] As a result, almost no difference was observed in both Fe-deficient and control roots before DEAE-Sepharose step (refer to FIG. 2). After the DEAE-Sepharose step it became clear that the 30- and 31-kDa peptides were induced by Fe-deficiency (refer to FIG. 13). After the Ether Toyopearl step, the 31 kDa peptide was eliminated from the active fraction. The 32 and 33 kDa peptides were found to be newly induced by Fe-deficiency (refer to FIG. 14). Activities were detected from the 32 and 33 kDa peptides, but no activity was detected from 30 kDa peptide (refer to FIG. 3).

[0044] Molecular weight of the enzyme of the present invention was determined by gel-filtration.

[0045] Estimated molecular weight of nicotianamine synthase by gel-filtration was reported to be 40,000-50,000

(Higuchi et al. Plant & Soil, Vol. 165, p. 173-179, 1994). But this did not correspond with the value estimated by SDS-PAGE.

[0046] In the present study, the buffer containing CHAPS effectively increased the resolution and molecular weight of the present enzyme was estimated to be 35,000 (refer to FIG. 4). this corresponds well to the value estimated by SDS-PAGE.

[0047] FIG. 4 shows elution pattern of nicotianamine synthase from the gel-filtration column. The black circles (●) indicate the enzyme activity and the solid line indicates absorption at 280 nm. The active fraction after hydroxyapatite chromatography was applied to a Sephacryl S300HR (Pharmacia) column (1.5 cm×71 cm, 125 ml), equilibrated with developing buffer (50 mM Tris, 1 mM EDTA, 0.1 M KCl, 0.05% CHAPS, 0.1 mM p-APMSF and 3 mM DTT, pH 8.0). Molecular weight markers used were thyroglobulin (Mr 670,000), γ -globulin (Mr 158,000), ovalbumin (Mr 44,000), and myoglobin (Mr 17,000). The linear flow was 10 cm/hour.

[0048] Partial amino acid sequence was determined from purified nicotianamine synthase.

[0049] The above explained 30 kDa, 32 kDa and 33 kDa peptides were purified from 1 kg of Fe-deficient barley roots by using a method in example 3 hereinbelow. These were partially degraded using a method in example 4 hereinbelow. Although 32- and 33-kDa peptides could not be completely separated from each other, these might have similar sequence or 32 kDa peptide was presumed to be the degradation product of 33 kDa peptide, and both of them were degraded in together.

[0050] The determined partial amino acid sequences indicated that these peptides were very similar in each other (FIG. 5). Further, since the molecular weights of the 33 kDa and 32 kDa (1) fragments had almost unchanged-molecular weight as compared with before degradation, this sequence might be N-terminal region of the present enzyme. As a result of computer search of the database, a gene of unknown function having very similar sequence to these sequences was found to exist in sativa and *Alabidopsis thaliana*. Especially, EST-cDNA clones D23792 and D24790 of sativa were very similar with 80.0% identity in a 33-amino acid overlap in the former and 68.4% identity in a 19-amino acid overlap in the latter (FIG. 5).

[0051] FIG. 5 shows a comparison with a six partial amino acid sequence determined by nicotianamine synthase originated from barley and similar sequence of graminaceous plants obtained by computer search of the database. Identical amino acid residue is shown in “:”. The part of nucleotide sequences indicated by the arrows was applied for the sequences of primer used in PCR.

[0052] Cloning and nucleotide sequences of cDNA clones encoding nicotianamine synthase were performed and determined.

[0053] PCR amplification of total cDNA prepared from Fe-deficient barley roots using degenerate primers designed from the partial amino acid sequence obtained from the method explained hereinbefore was performed, but the objective DNA could not amplified. Then the primers having single nucleotide sequence (shown by arrows in FIG. 5) from sequences of *Oryza sativa*, D23792 and D24790, were synthesized and PCR amplification was performed. The 205 bp fragment was amplified by PCR using NF and NR primers and the 274 bp fragment was amplified by PCR using IF and IR primers, and these contained the objective sequences. A cDNA library prepared using poly (A)⁺ RNA from Fe-deficient barley roots was screened and 19 positive clones using

the 205 bp fragment probe and 88 positive clones using the 274 fragment bp probe were obtained.

[0054] Among the thus obtained clones, the clone designated as HvNAS1, contained a translated region of 985 bp and amino acid sequence deduced therefrom was 328 amino acids residue, with deduced molecular weight of 35,144. This corresponded well with the value estimated by SDS-PAGE and gel-filtration. The partial amino acid sequences of the 32 kDa and 33 kDa peptides were included totally in HvNAS1 (FIG. 6).

[0055] FIG. 6 shows full length of HvNAS1 cDNA and amino acid sequence deduced therefrom. The underlined sequences indicate the identical partial amino acid sequences of fragments in the above FIG. 5. Numbers of the nucleotide sequence are indicated to the right of each row. Amino acid numbers are indicated on the left of each row.

[0056] The predicted pI of 5.2 matched the value estimated by native isoelectric focusing electrophoresis well. The six clones having very similar sequence other than HvNAS1, i.e. HvNAS2, HvNAS3, HvNAS4, HvNAS5, HvNAS6 and HvNAS7, were also obtained (Table 1, FIG. 7).

[0057] FIG. 7 shows comparison of the deduced amino acid sequences of the above 7 cDNA obtained from barley. Asterisks “*” indicates identical amino acid residues in all sequences.

[0058] The nucleotide sequences of these clones are shown in SEQ ID NO: 2 (HvNAS1), SEQ ID NO: 4 (HvNAS2), SEQ ID NO: 6 (HvNAS3), SEQ ID NO: 8 (HvNAS4), SEQ ID NO: 10 (HvNAS5), SEQ ID NO: 12 (HvNAS6) and SEQ ID NO: 14 (HvNAS7), respectively. The amino acid sequences of these amino acid sequences are shown in. SEQ ID NO: 1 (HvNAS1), SEQ ID NO: 3 (HvNAS2), SEQ ID NO: 5 (HvNAS3), SEQ ID NO: 7 (HvNAS4), SEQ ID NO: 9 (HvNAS5), SEQ ID NO: 11 (HvNAS6) and SEQ ID NO: 13 (HvNAS7), respectively.

TABLE 1

Clone	Properties of nas clones					
	Number of Amino Acid Residues	Molecular Weight	pI	Identity to nas 1 (%)	Identity to nas 2 (%)	Identity to nas 4 (%)
HvNAS 1	328	35144	5.20	—	—	—
HvNAS 2	336	35839	5.07	72	—	—
HvNAS 3	336	36013	5.47	72	95	—
HvNAS 4	330	35396	4.91	73	89	—
HvNAS 5	283	30148	5.22	61	61	59
HvNAS 6	329	35350	5.07	74	89	88
HvNAS 7	330	35244	4.98	70	86	91

[0059] The partial amino acid sequences determined from the 30 kDa peptide were all included in HvNAS5. The 5'- and 3'-non-translated regions of these clones were not similar with each other.

[0060] D23792 and D24790 similar to nicotianamine synthase of *Oryza sativa* were found with about 80% identity to HvNAS1. AC003114 and AB005245 of *Arbidopsis thaliana* were found with about 45% identity to HvNAS1.

[0061] The obtained HvNAS1 protein was expressed in *E. coli*.

[0062] The PCR amplification of HvNAS1 ORF was cloned with vector pMAL-c2 to express HvNAS1 fused with C-terminal of maltose binding protein. The expression of fused protein is strongly induced by IPTG.

[0063] The crude extract was obtained from the transformed *E. coli*, and nicotianamine synthase activity was assayed in the state of the fused protein. The crude extract

from the strain transformed with only the vector could not be detected the activity, whereas in case of inserted with HvNAS1 ORF, the activity was detected. Result is shown in FIG. 8.

[0064] FIG. 8 shows results of thin layer chromatographic (TLC) analysis of nicotianamine synthase obtained from *E. coli* crude extract expressing a fused protein of maltose binding protein-HvNAS1. In FIG. 8, lane 1: a standard nicotianamine synthase; lane 2: *E. coli* expressing maltose binding protein (SAM); and lane 3: *E. coli* expressing maltose binding protein-HvNAS1 fused protein.

[0065] Northern hybridization analysis conducted by the method described in example 7 hereinbelow indicated that this gene was strongly induced in Fe-deficient roots (FIG. 9). This coincides with expression pattern of the present enzyme activity (Higuchi et al. 1994). FIG. 9 shows a result of Northern hybridization analysis using HvNAS1 as a probe. Total RNA was extracted from after one week of Fe-deficient-treatment and control barley leaves and roots, and in each lane, 5 µg of RNA were electrophoresed.

[0066] Southern hybridization analysis of the barley genome DNA was performed according to the method described in example 8 hereinafter mentioned. Cutting of DNA with BamHI, EcoRI or HindIII produced plurality of fragments, however none of clones obtained at present could be digested by BamHI and EcoRI, consequently nicotianamine synthase gene might exist with multiple copies in genomes of barley and rice (FIG. 10).

[0067] FIG. 10 shows Southern hybridization analysis of HvNAS1 as a probe.

[0068] Genomic DNAs from barley and rice were digested with BamHI (lanes B), EcoRI (lanes R) and HindIII (lanes H) and 10 µg thereof were electrophoresed in each lane.

[0069] Further, using antigen prepared by the method described in example 9 hereinbelow, Western-blot analysis was performed according to the method described in example 10. It was found that the present enzyme protein was rapidly decomposed during the operation in the crude extract prepared for detecting the present enzyme activity (FIG. 11). The staining patterns coincided with the fact that the present enzyme activity was detected on the broad range between 30-35 kDa after SDS-PAGE (refer to FIG. 3).

[0070] FIG. 11 shows Western-blot analysis of crude enzyme used for detection of activity. SDS-PAGE was performed using 12.5% acrylamide slab gel. Protein 100 µg was electrophoresed.

[0071] The crude extract obtained from denatured protein according to the method described in example 10 hereinbelow was detected as almost single band with 35-36 kDa (FIG. 12). This value coincided with the deduced value from the amino acid sequence.

[0072] FIG. 12 shows Western-blot analysis of total protein extracted by trichloroacetic acid/acetone. SDS-PAGE was performed using 12.5% acrylamide slab gel. Protein 100 µg was electrophoresed. Proteins 200 µg extracted from roots and proteins 500 µg extracted from leaves were electrophoresed.

[0073] Western-blot analysis after 2-dimension electrophoresis reveals to detect several spots. This coincided with the fact of obtaining plurality of nicotianamine synthase gene. All spots were induced in Fe-deficient roots.

[0074] As a result that cDNA library from Fe-deficient rice roots poly (A)+RNA was screened using probes prepared by cutting HvNAS1 with restriction enzymes ApaI and XhoI, 20 clones were obtained. These clones were divided into 3 types of clones according to their sequences, and among them, only one type contains ORF full length, which was

designated as OsNAS1. Nucleotide sequence of OsNAS1 is shown in SEQ ID NO: 16 and amino acid sequence is shown in SEQ ID NO: 15.

[0075] PCR amplification of OsNAS1 ORF was cloned with a vector pMAL-c2 to express a form fused with maltose binding protein C-terminal. The fused protein is strongly induced its expression by IPTG.

[0076] Crude extract from the transformed *E. coli* with the fused protein was obtained and nicotianamine synthase activity was assayed in the state of the fused protein. The same activity with HvNAS1 was detected. Result is shown in FIG. 15. FIG. 15 shows results of thin layer chromatographic (TLC) analysis of nicotianamine synthase obtained from *E. coli* crude extract expressing a fused protein of maltose binding protein-OsNAS1. In FIG. 15, lane 1: a standard nicotianamine (NA); lane 2: an extract from *E. coli* expressing maltose binding protein-OsNAS1 fused protein; and lane 3: an extract from *E. coli* expressing maltose binding protein-HvNAS1 fused protein.

[0077] Northern hybridization analysis conducted by the method described in example 7 hereinbelow indicated that in contrast to barley, the expression was induced in rice by Fe-deficient treatment not only in roots but also in leaves (FIG. 16). FIG. 16 shows a result of Northern hybridization analysis using OsNAS1 ORF as a probe. Total RNA was extracted from after two weeks of Fe-deficient treatment and control rice leaves and roots, and in each lane, 5 µg of RNA were electrophoresed.

[0078] Nucleotide sequence of *Arabidopsis thaliana* similar to HvNAS1 obtained by computer search of the database was used as a primer. PCR amplification for genome DNA of *Arabidopsis thaliana* resulted to obtain three nicotianamine synthase genes.

[0079] These were designated as AtNAS1, AtNAS2 and AtNAS3.

[0080] Nucleotide sequence of these genes are shown in SEQ ID NO: 18 (AtNAS1), SEQ ID NO: 20 (AtNAS2) and SEQ ID NO: 22 (AtNAS3). These amino acid sequences are shown in SEQ ID NO: 17 (AtNAS1), SEQ ID NO: 19 (AtNAS2) and SEQ ID NO: 21 (AtNAS3).

[0081] AtNAS1, AtNAS2 and AtNAS3 ORF were amplified with PCR and were cloned with a vector pMAL-c2. Each of them was tried to be expressed in the form of fusing with maltose binding protein C-terminal. The expression of the fused protein was strongly induced by IPTG.

[0082] Crude extract from the transformed *E. coli* with the fused protein was obtained and nicotianamine synthase activity was assayed in the state of the fused protein. The activity was detected. Result is shown in FIG. 17. FIG. 17 shows results of TLC analysis of nicotianamine synthase activity obtained from *E. coli* crude extract expressing a fused protein of maltose binding protein—ATNAS. In FIG. 17, lanes 1: a standard nicotianamine (NA) and S-adenosylmethionine; lanes 2: an extract from *E. coli* expressing only maltose binding protein; lanes 3: an extract from *E. coli* expressing maltose binding protein-AtNAS1 fused protein; lanes 4: an extract from *E. coli* expressing maltose binding protein-AtNAS2 fused protein; and lanes 5: an extract from *E. coli* expressing maltose binding protein-AtNAS3 fused protein.

[0083] RT-PCR was conducted according to the method described in example 11 hereinbelow. It was found that AtNAS1 was expressed in the roots and the aboveground parts of *Arabidopsis thaliana*, whereas AtNAS2 was expressed neither in the roots nor in the aboveground parts, and AtNAS3 was expressed only in the roots (FIG. 18). In FIG. 18, lane M shows molecular weight marker. Gene expression was conducted in the aboveground parts, roots and

positive controls. In the figure, lanes C: AtNAS1 and AtNAS2 ORF full length were amplified; lanes 1: AtNAS1 specific amplification fragments; lanes 2: AtNAS2 specific amplification fragments; and lanes 3: AtNAS3 specific amplification fragments.

[0084] The amount of secreted mugineic acid is reported increased up to 20 mg mugineic acid/g roots dry weight/day (Takagi, 1993). Crude nicotianamine synthase activity detected by the present inventors was sufficient to fulfill it. Since the present enzyme proteins exist in more than several types and 30 kDa peptide without activity exists, it can be speculated that as a result of aggregation of these peptides, the constructed structure, which is preferable for binding with 3 molecules of S-adenosylmethionine, reveals maximum activity. The molecular weight estimated by gel-filtration was 35,000 (FIG. 4).

[0085] Increase in activity by re-aggregation of subunits has not been observed at present. Since the fused protein with maltose binding protein and subunits showed its activity, we have at present an idea that the present enzyme might be a monomer. However, the possibility that large activity can be revealed by constructing multimer, can not completely denied.

[0086] The reaction mechanism synthesizing nicotianamine from S-adenosylmethionine may be similar to methyl transfer reaction using S-adenosylmethionine as a methyl donor, and a reaction synthesizing spermidine and spermine from decarboxylated S-adenosylmethionine. The common catalytic domain of these enzymes has been discussed in relation to equivalent amino acids configuration occupying similar positions in higher-order structures (Hashimoto et al. 1998 and Schluckebier et al. 1995).

[0087] In future, catalytic domain may be elucidated as the results of comparison with nicotianamine synthase from other plant species or X-ray crystallography.

[0088] Induction of nicotianamine synthase activity by Fe-deficiency is a specific phenomenon in graminaceous plants, and is essential for mass production of mugineic acid family. *Oryza sativa* is a plant, in which secretion of mugineic acid family is the least among major graminaceous plants, consequently it is very weak for Fe-deficiency in calcareous soil.

[0089] Consequently, as a result of creating transformant *Oryza sativa* having tolerance to Fe-deficiency by introducing nicotianamine synthase gene of the present invention into the graminaceous plants, especially *Oryza sativa*, and expressing large amount at the Fe-deficiency, cultivation of rice in the calcareous soil can be possible.

[0090] Heretofore, in the graminaceous plants, nicotianamine has been thought to have only a role as a precursor for synthesis of mugineic acid family. However, since the present invention has elucidated that nicotianamine synthase gene constituted the multiple gene family, it may play other important roles in the graminaceous plants.

[0091] In plants, which lack the ability to secrete mugineic acid family, except for graminaceous plants, it has been proposed that nicotianamine plays a key role as an endogenous chelator of divalent metal cations, such as Fe²⁺, Cu²⁺, Zn²⁺ and Mn²⁺, and that it contributes to the homeostasis of those metals (Stephan et al. 1994). Consequently, it may play the same role in the graminaceous plants.

[0092] Nicotianamine synthase activity is not induced in dicots, and expression of gene of the present invention may not be induced by Fe-deficiency. We have cloned nicotianamine synthase genes of *Arabidopsis thaliana*. Composition

of promoter regions in these genes can elucidate the mechanism of gene expression caused by Fe-deficiency, and the gene of the present invention may play important function not only in the graminaceous plants but also in the dicots.

[0093] SEQ ID NO: 1 shows amino acid sequence of nicotianamine synthase of the present invention.

[0094] The present invention includes nicotianamine synthase having amino acid sequence shown in SEQ ID NO: 1. However, the present invention is not limited within the above nicotianamine synthase. The nicotianamine synthase of the present invention includes, unless it loses nicotianamine synthase activity, the peptides, in which a part of the amino acid sequence of said peptide is deleted, preferably 50% or less, more preferably 30% or less, or more further preferably 10% or less in the total amino acids, or is substituted by other amino acids, or to which other amino acids are further added, or in which these deletion, substitution and addition may be combined.

[0095] Nucleotide sequence coding nicotianamine synthase of the present invention is shown in SEQ ID NO: 2.

[0096] The present invention also includes not only a gene coding nicotianamine synthase shown in SEQ ID NO: 2 but also genes coding nicotianamine synthase mentioned herein-above.

[0097] The vector of the present invention introducing the above gene is not specifically limited, and various vectors can be introduced. Preferable vector is the expression vector.

[0098] Various cells can be transformed conventionally by using recombinant vector of the present invention. Mass production of nicotianamide can be performed by using the thus obtained transformant. These methods are well known in the person skilled in the art.

[0099] Examples of hosts for introducing the gene of the present invention are bacteria, yeasts and cells. Preferable host is plants, especially the graminaceous plant.

[0100] Method for introducing gene is not limited. It can be made by using vector or can be directly introduce in genome.

[0101] Antibody of the present invention against nicotianamine synthase can be prepared conventionally by using nicotianamine synthase of the present invention. Antibody can be a polyclonal antibody or, if necessary, monoclonal antibody.

[0102] Further, a selective breeding of plants, preferably graminaceous plants, can be made by using gene of the present invention. Especially, the gene of the present invention can be applied for improvement of varieties, which can grow even in Fe-deficient soil.

EXAMPLES

[0103] The following examples illustrate the present invention, but are not construed as limiting the present invention.

Example 1

Preparation of Plant Material

[0104] Seeds of barley (*Hordeum vulgare* L. cv Ehimehadakamugi No. 1) were germinated on wet filter paper and transferred into the standard hydroponic culture solution (Mori and Nishizawa, 1987) in a glass house at natural temperature under natural light. The pH of the hydroponic culture solution was adjusted at 5.5 by 0.5 N HCl everyday. When the third leaves developed, the plants were transferred to the hydroponic culture solution without containing Fe. The pH was maintained at 7.0 by 0.5 N NaOH everyday. The control

plants were also cultured in the standard culture solution continuously. The culture solution was renewed once in every week. Two weeks after starting Fe-deficient treatment, when severe iron chlorosis significantly appeared on the 4th and 5th leaves, roots were harvested and frozen in liquid N₂ and stored at -80° C. until use.

Example 2

Assay of Nicotianamine Synthase Activity

[0105] Modified assay method reported previously by the present inventors (Higuchi et al. 1996a) was used. Enzyme solutions were equilibrated with reaction buffer [50 mM Tris, 1 mM EDTA, 3 mM dithiothreitol (hereinafter designates as DTT), 10 μM (p-aminodiphenyl) methanesulfonyl fluoride (hereinafter designates as p-APMSF) and 10 μM trans-epoxysuccinyl-leucylamido-(4-guanidino) butane (hereinafter designates, as E-64), pH 8.7]. Buffer exchange was performed by using ultrafiltration unit, Ultrafree C3LGC NMWL10000 (Millipore Co.). S-adenosylmethionine labeled with ¹⁴C in carboxyl group (Amersham Inc.) was added to the enzyme solution at the final concentration of 20 μM and kept at 25° C. for 15 minutes. The reaction products were separated by thin layer chromatography on silica gel LK6 (Whatman Inc.) using developer (phenol: butanol: formic/acid: water=12:3:2:3). Radioactivity of the reaction products was detected by image Analyzer BAS-2000 (Fuji Film Co.). The protein content was assayed by Bradford method using Protein Assay Kit (Bio Rad Inc.).

Example 3

Purification of Nicotianamine Synthase

[0106] The following operations were performed at 4° C. and E-64 was added to fractions containing nicotianamine synthase at the final concentration of 10 μM.

[0107] The frozen roots were crushed into a fine powder in liquid N₂ and homogenized in a household juicer with 200 ml of extraction buffer [0.2 M Tris, 10 mM EDTA, 5% (v/v) glycerol, 10 mM DTT, 0.1 mM E-64, 0.1 mM p-APMSF and 5% (w/v) insoluble polyvinylpyrrolidone (VP), pH 8.0] per 100 g of roots. The homogenate was centrifuged for 30 minutes at 22,500×g to obtain supernatant. Ammonium sulfate was added to the supernatant to yield a final concentration of 0.4 M and allowed to stand for 1 hour. Again, the mixture was centrifuged for 30 minutes at 22,500×g to obtain supernatant.

[0108] The supernatant was loaded onto a TSK gel Butyl Toyopearl 650M column (10 ml bed volume per 100 g of roots), equilibrated with the adsorption buffer [20 mM Tris, 1 mM EDTA, 3 mM DTT, 0.4 M (NH₄)₂SO₄ and 0.1 mM p-APMSF, pH 8.0] and eluted with elution buffer [10 mM Tris, 1 mM EDTA, 3 mM DTT, 0.1 mM p-APMSF, 5% glycerol and 0.05% 3-[(3-chloramidopropyl) dimethyl-ammonio] propanesulfonic acid (hereinafter designates as CHAPS), pH 8.0].

[0109] KCl was added to the active fraction to give a final concentration of 0.4 M, and 1 M potassium phosphate buffer (pH 8.0) was added to a final concentration of 1 mM of KCl. A hydroxyapatite 100-350 mesh (Nacalai Tesque), equilibrated with the adsorption buffer (1 mM K-P, 10 mM KCl, 3 mM DTT and 0.1 mM p-APMSF, pH 8.0), was prepared at 10 ml per protein 100 mg and the fractions containing nicotianamine synthase were loaded. Nicotianamine synthase was passed through without adsorption. The passed through fraction was loaded onto TSK gel Butyl Toyopearl 650M column (1 ml bed volume per 10 mg of protein), and nicotianamine synthase was eluted in the manner described above.

[0110] The active fraction was loaded onto a DEAE-Sepharose FF column (5 ml bed volume per 25 mg of protein, Pharmacia) equilibrated with the adsorption buffer (20 mM Tris, 1 mM EDTA, 3 mM DTT, 0.1 mM p-APMSF and 0.05% CHAPS, pH 8.0) and eluted with stepwise gradient elution of potassium chloride concentration of 0.05 M, 0.1 M, 0.15 M and 0.2 M. Nicotianamine synthase was eluted at 0.15 M of KCl concentration.

[0111] The active fraction was loaded onto the Ether Toyopearl 650M column (10 ml bed volume per 100 g of roots), equilibrated with adsorption buffer [20 mM Tris, 1 mM EDTA, 3 mM DTT, 1.2 M (NH₄)₂SO₄ and 0.1 mM p-APMSF, pH 8.0]. Nicotianamine synthase was not adsorbed and passed through from the column. The passed through fraction was loaded onto TSK gel Butyl Toyopearl 650M column and fractions containing nicotianamine synthase was eluted. The peptides in the active fraction containing nicotianamine synthase, which was purified by the above column chromatographic treatments, were separated by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (hereinafter designates as SDS-PAGE) using 11% acrylamide slab gels. After SDS-PAGE the gel was stained with 0.3 M copper chloride (Dzandu et al. 1988), and the separated bands were cut out. The gel fragments were destained with 0.25 M EDTA/0.25 M Tris (pH 9.0) and homogenized, with the extraction buffer (1% SDS, 25 mM Tris and 192 mM glycine). Each homogenate was electroeluted with SDS-free buffer (25 mM Tris and 192 mM glycine) and peptide was recovered.

Example 4

Determination of Partial Amino Acid Sequence

[0112] The isolated nicotianamine synthase was digested chemically with cyanogen bromide (Gross 1967).

[0113] After SDS-PAGE treatment, 10-fold volume of 70% (v/v) formic acid and 1% (w/v) cyanogen bromide were added to gel fragments containing nicotianamine synthase and decomposed at 4° C. for overnight. After completion of digestion, the liquid part was collected and dried in vacuo. The dried substance was dissolved in SDS-PAGE sample buffer, and allowed to stand at room temperature for overnight, then the digested product was separated by SDS-PAGE using 16.5% acrylamide gel containing Tricine (Schagger and Jagow, 1987). The peptides were transferred onto a PVDF membrane by electroblotting (Towbin et al. 1979) and stained with amido black. The stained bands were cut out and the amino acid sequence was determined from N-terminal side of each peptide by Edman degradation in gas-phase sequencer (model 492A protein sequencer, Applied Biosystems Inc.).

Example 5

Cloning of Nicotianamine Synthase Genes

[0114] PCR amplification was conducted for cDNA originated from Fe-deficient barley roots using primers, which were synthesized based on the obtained partial amino acid sequence. A pYH23 cDNA library prepared from the poly (A)⁺RNA of Fe-deficient barley roots was screened with the thus obtained DNA fragments of PCR product, which was labeled with [α-³²P]dATP using the random primer kit (Takara Shuzo Co.), as the primers. The isolated cDNA clones were sequenced by cycle sequencing kit (Shimadzu Bunko Co.) using Shimadzu DNA sequencer DSQ-2000L.

[0115] PCR amplification was conducted for genomic DNA of *Arabidopsis thaliana* using primers, which were synthesized based on nucleotide sequences of AC003114 and

AB005245 of *Arabidopsis thaliana*. The thus obtained DNA fragments were sequenced by cycle sequencing kit (Shimadzu Bunko Co.) using Shimadzu DNA sequencer DSQ-1000L.

[0116] The determined nucleotide sequence is shown in SEQ ID NO: 2.

Example 6

Expression of NAS1 Protein in *E. coli*

[0117] A fragment, in which EcoRI site was introduced into the upstream of the first ATG of the HvNAS1 cDNA and PstI and BamHI sites were introduced into the downstream of the stop codon of the HvNAS1 cDNA, was amplified by PCR. The thus obtained amplified product was subcloned in the pBluescriptII SK— using EcoRI site and BamHI site, and the correct nucleotide sequence was confirmed. The fragment between EcoRI site and PstI site was cloned into pMAL-c2 to make expression in the form of fusing the HvNAS1 to the C-terminal of maltose binding protein.

[0118] A fragment, in which EcoRI site was introduced into the upstream of the first ATG of the OsNAS1 and HindIII site was introduced into the downstream of the stop codon of the OsNAS1, was amplified by PCR. The thus obtained amplified product was subcloned in the pBluescriptII SK— using EcoRI site and HindIII site, and the correct nucleotide sequence was confirmed. The fragment between EcoRI site and HindIII site was cloned into pMAL-c2 to make expression in the form of fusing the OsNAS1 to the C-terminal of maltose binding protein.

[0119] A fragment, in which EcoRI site was introduced into the upstream of the first ATG of the AtNAS1, AtNAS2 and AtNAS3 and XbaI site was introduced into the downstream of the stop codon of the AtNAS1, AtNAS2 and AtNAS3, was amplified by PCR. The thus obtained amplified products were subcloned in the pBluescriptII SK—, and the correct nucleotide sequences were confirmed. The fragment between EcoRI site and XbaI site was cloned into pMAL-c2 to make expression in the form of fusing the AtNAS1, AtNAS2 and AtNAS3 to the C-terminal of maltose binding proteins, respectively.

[0120] *E. coli* strain XL1-Blue was used as a host for expressing the said fused protein. pMAL-c2-HvNAS1 and pMAL-c2, respectively, were introduced into XL1-Blue. The thus obtained recombinant bacteria were cultured in LB medium containing ampicillin and tetracycline, each 50 µg/ml, at 37° C. until the OD 600 of the culture reached 0.5. Isopropyl 6-D-thiogalactopyranoside (IPTG) was added to the final concentration of 0.3 mM, and continuously cultured at 37° C. for 3 hours, and collected bacterial cells. Cells were suspended in 10 mM Tris buffer containing 0.2 M NaCl, 1 mM EDTA, 3 mM DTT and 0.1 mM E-64, pH 7.4 and frozen with liquid nitrogen. This was melted in ice water and ultrasonication for 15 seconds was repeated for 10 times. Nicotianamine synthase activity of the thus obtained crude extract was assayed according to the method described in example 2 and the enzyme activity was confirmed.

Example 7

Northern Hybridization

[0121] Northern hybridization of barley RNA was performed using DNA fragment, which was prepared by cutting HvNAS1 cDNA with HindIII and NotI and labeled with [α -³²P] dATP, as a probe. Total RNA was extracted from

barley (Naito et al. 1988). The extracted RNA was separated by 1.4% agarose gel electrophoresis, and blotted onto Hybond-N⁺ membranes (Amersham). Northern hybridization of rice RNA was performed using OsNAS1 ORF, which was labeled with [α -³²P]dATP, as a probe. Total RNA was extracted from rice. The extracted RNA was separated by 1.4% agarose gel electrophoresis, and blotted onto Hybond-N⁺ membranes (Amersham). The membrane was hybridized with the probe in 0.5 M Church phosphate buffer (Church and Gilbert 1984), 1 mM EDTA, 7% (w/v) SDS with 100 µg/ml salmon sperm DNA at 65° C. for overnight. The membrane was washed with buffer containing 40 mM Church phosphate buffer and 1% (w/v) SDS at 65° C. for 10 minutes. After the washing was repeated once again, the membrane was washed with buffer containing 0.2×SSPE and 0.1% (w/v) SDS at 65° C. for 10 minutes. Radioactivity was detected using the image analyzer BAS-2000.

[0122] Results are shown in FIG. 9 and FIG. 16.

Example 8

Southern Hybridization

[0123] Genomic DNA was extracted from leaves of barley and rice. The extract was digested with BamHI, EcoRI or HindIII, separated on a 0.8% (w/v) agarose gel electrophoresis, and transferred onto Hybond-N⁺ membranes (Amersham). The hybridization was performed according to the method described in example 7 and radioactivity was detected.

[0124] Result is shown in FIG. 10.

Example 9

Preparation of Polyclonal Antibody

[0125] Total protein was extracted using trichloroacetic acid and acetone (Damerval et al. 1986). The plants were crashed in the liquid nitrogen until powder was obtained, and mixed with acetone containing 0.1% (v/v) 2-mercaptoethanol. The protein was precipitated by allowing to stand at -20° C. for 1 hour, and the precipitate was collected by centrifugation at 16,000×g for 30 minutes. The precipitate was suspended in acetone containing 0.1% (v/v) 2-mercaptoethanol and allowed to stand at -20° C. for 1 hour, then collected the precipitate by centrifugation at 16,000×g for 30 minutes. The precipitate was dried in vacuo, and dissolved in the sample buffer [9.5 M urea, 2% (w/v) Triton X-100 and 5% (v/v) 2-ME], then centrifuged at 16,000×g for 10 minutes to obtain the supernatant. The proteins contained in the supernatant were separated by SDS-PAGE or the denaturing two-dimensional electrophoresis (O'Farrell 1975) and transferred onto PVDF membrane. Western blotting analysis was performed by applying the primary antibody containing anti-nicotianamine synthase antibody prepared in example 1 and the secondary antibody containing horse radish binding anti-mouse IgG (H+L) goat antibody (Wako Pure Chemicals Co.) on the membrane and coloring with diaminobenzidine.

[0126] Result is shown in FIG. 12. SDS-PAGE was performed using 12.5% acrylamide slab gel. Protein 100 µg was electrophoresed. Proteins of roots 200 µg and leaves 500 µg were electrophoresed.

Example 11

RT-PCR

[0127] Total RNA was extracted from *Arabidopsis thaliana*. RT-PCR was performed with 1 µg RNA as a tem-

plate by using the EZ rTth RNA PCR kit (Parkin Elmer Inc.). Specific primers for AtNAS1, AtNAS2 and AtNAS3, respectively, were used.

[0128] Result is shown in FIG. 18.

INDUSTRIAL APPLICABILITY

[0129] Various cells are transformed according to the conventional method by using recombinant vectors of the present

invention. Mass production of nicotianamine can be performed by using the obtained transformant. These methods can be performed according to the method known in the person skilled in the art.

[0130] Selective breeding of plants, preferably graminaceous plants can also be performed using genes of the present invention. Especially, genes of the present invention can be applied for improving varieties, which can grow on Fe-deficient soil.

SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 37

<210> SEQ ID NO 1

<211> LENGTH: 328

<212> TYPE: PRT

<213> ORGANISM: Hordeum vulgare

<400> SEQUENCE: 1

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1           5           10           15

Gly Ile Gln Ala Ala Ile Ala Glu Leu Pro Ser Leu Ser Pro Ser Pro
20          25          30

Glu Val Asp Arg Leu Phe Thr Asp Leu Val Thr Ala Cys Val Pro Pro
35          40          45

Ser Pro Val Asp Val Thr Lys Leu Ser Pro Glu His Gln Arg Met Arg
50          55          60

Glu Ala Leu Ile Arg Leu Cys Ser Ala Ala Glu Gly Lys Leu Glu Ala
65          70          75          80

His Tyr Ala Asp Leu Leu Ala Thr Phe Asp Asn Pro Leu Asp His Leu
85          90          95

Gly Leu Phe Pro Tyr Tyr Ser Asn Tyr Val Asn Leu Ser Arg Leu Glu
100         105        110

Tyr Glu Leu Leu Ala Arg His Val Pro Gly Ile Ala Pro Ala Arg Val
115        120        125

Ala Phe Val Gly Ser Gly Pro Leu Pro Phe Ser Ser Leu Val Leu Ala
130        135        140

Ala His His Leu Pro Glu Thr Gln Phe Asp Asn Tyr Asp Leu Cys Gly
145        150        155        160

Ala Ala Asn Glu Arg Ala Arg Lys Leu Phe Gly Ala Thr Ala Asp Gly
165        170        175

Val Gly Ala Arg Met Ser Phe His Thr Ala Asp Val Ala Asp Leu Thr
180        185        190

Gln Glu Leu Gly Ala Tyr Asp Val Val Phe Leu Ala Ala Leu Val Gly
195        200        205

Met Ala Ala Glu Glu Lys Ala Lys Val Ile Ala His Leu Gly Ala His
210        215        220

Met Val Glu Gly Ala Ser Leu Val Val Arg Ser Ala Arg Pro Arg Gly
225        230        235        240

Phe Leu Tyr Pro Ile Val Asp Pro Glu Asp Ile Arg Arg Gly Gly Phe
245        250        255

Glu Val Leu Ala Val His His Pro Glu Gly Glu Val Ile Asn Ser Val
260        265        270

Ile Val Ala Arg Lys Ala Val Glu Ala Gln Leu Ser Gly Pro Gln Asn

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275	280	285	
Gly Asp Ala His Ala Arg Gly Ala Val Pro Leu Val Ser Pro Pro Cys			
290	295	300	
Asn Phe Ser Thr Lys Met Glu Ala Ser Ala Leu Glu Lys Ser Glu Glu			
305	310	315	320
Leu Thr Ala Lys Glu Leu Ala Phe			
325			
<210> SEQ ID NO 2			
<211> LENGTH: 1295			
<212> TYPE: DNA			
<213> ORGANISM: Hordeum vulgare			
<220> FEATURE:			
<221> NAME/KEY: CDS			
<222> LOCATION: (55)..(1041)			
<400> SEQUENCE: 2			
gcggttcagag gcttcacagag ttcttcocggt caccaagaag catttgatca taac atg			57
Met			
1			
gat gcc cag aac aag gag gtc gct gct ctg atc gag aag atc gcc ggt			105
Asp Ala Gln Asn Lys Glu Val Ala Ala Leu Ile Glu Lys Ile Ala Gly			
5	10	15	
atc cag gcc gcc atc gcc gag ctg ccg tgg ctg agc ccg tcc ccc gag			153
Ile Gln Ala Ala Ile Ala Glu Leu Pro Ser Leu Ser Pro Ser Pro Glu			
20	25	30	
gtc gac agg ctc ttc acc gac ctc gtc acg gcc tgc gtc ccg ccg agc			201
Val Asp Arg Leu Phe Thr Asp Leu Val Thr Ala Cys Val Pro Pro Ser			
35	40	45	
ccc gtc gac gtg acg aag ctc agc ccg gag cac cag agg atg cgg gag			249
Pro Val Asp Val Thr Lys Leu Ser Pro Glu His Gln Arg Met Arg Glu			
50	55	60	65
gct ctc atc cgc ttg tgc tcc gcc gcc gag ggg aag ctc gag gcg cac			297
Ala Leu Ile Arg Leu Cys Ser Ala Ala Glu Gly Lys Leu Glu Ala His			
70	75	80	
tac gcc gac ctg ctc gcc acc ttc gac aac ccg ctc gac cac ctc ggc			345
Tyr Ala Asp Leu Leu Ala Thr Phe Asp Asn Pro Leu Asp His Leu Gly			
85	90	95	
ctc ttc ccg tac tac agc aac tac gtc aac ctc agc agg ctg gag tac			393
Leu Phe Pro Tyr Tyr Ser Asn Tyr Val Asn Leu Ser Arg Leu Glu Tyr			
100	105	110	
gag ctc ctg gcg cgc cac gtg ccg ggc atc gcg ccg gcg cgc gtc gcc			441
Glu Leu Leu Ala Arg His Val Pro Gly Ile Ala Pro Ala Arg Val Ala			
115	120	125	
ttc gtc ggc tcc gcc ccg ctg ccg ttc agc tgg ctc gtc ctc gcc gcg			489
Phe Val Gly Ser Gly Pro Leu Pro Phe Ser Ser Leu Val Leu Ala Ala			
130	135	140	145
cac cac ctg ccc gag acc cag ttc gac aac tac gac ctg tgc ggc gcg			537
His His Leu Pro Glu Thr Gln Phe Asp Asn Tyr Asp Leu Cys Gly Ala			
150	155	160	
gcc aac gag cgc gcc agg aag ctg ttc ggc gcg acg gcg gac ggc gtc			585
Ala Asn Glu Arg Ala Arg Lys Leu Phe Gly Ala Thr Ala Asp Gly Val			
165	170	175	
ggc gcg cgt atg tgg ttc cac acg gcg gac gtc gcc gac ctc acc cag			633
Gly Ala Arg Met Ser Phe His Thr Ala Asp Val Ala Asp Leu Thr Gln			
180	185	190	
gag ctc ggc gcc tac gac gtg gtc ttc ctc gcc gcg ctc gtc ggc atg			681
Glu Leu Gly Ala Tyr Asp Val Val Phe Leu Ala Ala Leu Val Gly Met			

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195	200	205	
gca gcc gag gag aag gcc aag gtg att gcc cac ctg ggc gcg cac atg			729
Ala Ala Glu Glu Lys Ala Lys Val Ile Ala His Leu Gly Ala His Met			
210	215	220	225
gtg gag ggg gcg tcc ctg gtc gtg cgg agc gca cgg ccc cgc ggc ttt			777
Val Glu Gly Ala Ser Leu Val Val Arg Ser Ala Arg Pro Arg Gly Phe			
230	235	240	
ctt tac ccc att gtc gac ccg gag gac atc agg cgg ggt ggg ttc gag			825
Leu Tyr Pro Ile Val Asp Pro Glu Asp Ile Arg Arg Gly Gly Phe Glu			
245	250	255	
gtg ctg gcc gtg cac cac ccg gaa ggt gag gtg atc aac tct gtc atc			873
Val Leu Ala Val His His Pro Glu Gly Glu Val Ile Asn Ser Val Ile			
260	265	270	
gtc gcc cgt aag gcc gtc gaa gcg cag ctc agt ggg ccg cag aac gga			921
Val Ala Arg Lys Ala Val Glu Ala Gln Leu Ser Gly Pro Gln Asn Gly			
275	280	285	
gac gcg cac gca cgg ggc gcg gtg ccg ttg gtc agc ccg cca tgc aac			969
Asp Ala His Ala Arg Gly Ala Val Pro Leu Val Ser Pro Pro Cys Asn			
290	295	300	305
ttc tcc acc aag atg gag gcg agc gcg ctt gag aag agc gag gag ctg			1017
Phe Ser Thr Lys Met Glu Ala Ser Ala Leu Glu Lys Ser Glu Glu Leu			
310	315	320	
acc gcc aaa gag ctg gcc ttt tga ttgaagagtg cgcgtggtca ttctgtcgcc			1071
Thr Ala Lys Glu Leu Ala Phe			
325			
tgcgatcgtg gtaactttcc tactcgtgtg tgttttgatg tttgtgectg taagagttat			1131
gcttccggcc ttgtgctggt aatttacacg cgttacatgt agtacttgta tttatactg			1191
gaataacggt atgtaacata aatattagtg ggatttgaag tgtaatgcta aataataaga			1251
aaacttgatg cagacattca aaaaaaaaaa aaaaaaaaaa aaaa			1295

<210> SEQ ID NO 3

<211> LENGTH: 335

<212> TYPE: PRT

<213> ORGANISM: Hordeum vulgare

<400> SEQUENCE: 3

Met Ala Ala Gln Asn Asn Gln Glu Val Asp Ala Leu Val Glu Lys Ile			
1	5	10	15
Thr Gly Leu His Ala Ala Ile Ala Lys Leu Pro Ser Leu Ser Pro Ser			
20	25	30	
Pro Asp Val Asp Ala Leu Phe Thr Glu Leu Val Thr Ala Cys Val Pro			
35	40	45	
Pro Ser Pro Val Asp Val Thr Lys Leu Gly Pro Glu Ala Gln Glu Met			
50	55	60	
Arg Glu Gly Leu Ile Arg Leu Cys Ser Glu Ala Glu Gly Lys Leu Glu			
65	70	75	80
Ala His Tyr Ser Asp Met Leu Ala Ala Phe Asp Lys Pro Leu Asp His			
85	90	95	
Leu Gly Met Phe Pro Tyr Tyr Asn Asn Tyr Ile Asn Leu Ser Lys Leu			
100	105	110	
Glu Tyr Glu Leu Leu Ala Arg Tyr Val Pro Gly Gly Tyr Arg Pro Ala			
115	120	125	
Arg Val Ala Phe Ile Gly Ser Gly Pro Leu Pro Phe Ser Ser Phe Val			
130	135	140	

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Leu Ala Ala Arg His Leu Pro Asp Thr Met Phe Asp Asn Tyr Asp Leu
 145 150 155 160
 Cys Gly Ala Ala Asn Asp Arg Ala Ser Lys Leu Phe Arg Ala Asp Arg
 165 170 175
 Asp Val Gly Ala Arg Met Ser Phe His Thr Ala Asp Val Ala Asp Leu
 180 185 190
 Ala Gly Glu Leu Ala Lys Tyr Asp Val Val Phe Leu Ala Ala Leu Val
 195 200 205
 Gly Met Ala Ala Glu Asp Lys Ala Lys Val Ile Ala His Leu Gly Ala
 210 215 220
 His Met Ala Asp Gly Ala Ala Leu Val Val Arg Ser Ala His Gly Ala
 225 230 235 240
 Arg Gly Phe Leu Tyr Pro Ile Val Asp Pro Gln Asp Ile Gly Arg Gly
 245 250 255
 Gly Phe Glu Val Leu Ala Val Cys His Pro Asp Asp Asp Val Val Asn
 260 265 270
 Ser Val Ile Ile Ala Gln Lys Ser Lys Asp Val His Ala Asp Gly Leu
 275 280 285
 Gly Ser Gly Arg Gly Ala Gly Gly Gln Tyr Ala Arg Gly Thr Val Pro
 290 295 300
 Val Val Ser Pro Pro Cys Arg Phe Gly Glu Met Val Ala Asp Val Thr
 305 310 315 320
 Gln Asn His Lys Arg Asp Glu Phe Ala Asn Ala Glu Val Ala Phe
 325 330 335

<210> SEQ ID NO 4

<211> LENGTH: 1342

<212> TYPE: DNA

<213> ORGANISM: Hordeum vulgare

<400> SEQUENCE: 4

```

ctcctgtgcc tgtcctgagg taccaagaac accagtgaac tggctgcccc gaacaaccag    60
gaggtggatg ccctggtgga gaagatcacc gggctccatg cgcgaatcgc caagctgccg    120
tcgctcagcc catccccgga cgtcgacgcg ctcttcacgg agctggtcac ggcgtgcggt    180
ccaccgagtc cagtggacgt gaccaagetc gggccggagg cgcaggagat gcgggagggc    240
ctcatccgcc tatgtccga ggcgagggg aagctggagg cgcactactc cgacatgctc    300
gccgccttcg acaagccgct ggatcacctc ggcattgttc cctactacaa caactacatc    360
aaactcagca agctcgagta cgagctctct gcccgtacg tgcctggcgg ctatcgcccg    420
gcgcgcgtcg cgttcacatg ctcgagcccg ctgcccgtca gctcctttgt cctggccgcg    480
cgccacctgc ccgacacat gttcgacaac tatgacctgt gcggtgcggc caacgatcgc    540
gccagcaagc tcttcgcgc ggatcgcgac gtgggtgccc gcatgtcgtt ccacacggcc    600
gacgtcgcgg acctcgccgg cgagctcgcc aagtaecgac ttgtcttctt ggccgcactc    660
gtcggcatgg ccgccgagga caaggcgaag gtgatcgcgc acctcggcgc acacatggca    720
gacggggcgg cctcgtctgt gcgcagcgca cacggagcgc gggggttctt gtaccgatc    780
gtcgaccccc aggacatcgg ccgaggcggg ttcgaggtgc tggccgtgtg ccatccccgac    840
gacgacgtgg tgaactcctg catcatcgca cagaagtcca aggacgtgca tgccgatgga    900
cttggcagcg ggcgtggtgc cgggtgacag tacgcgcccg gcacggtgcc tgttgtcagc    960

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cccccgtagc ggctcggcga gatgggtggcg gacgtgaccc agaaccacaa gagagacgag 1020
tttgccaacg ccgaagtggc cttttgatcg ttcgctgcga ggggtgtgcat ccatgatcca 1080
tccatcacctc gttctgtgat tgcatacaagc ttgcaatcgt atgcatttca agtcacgtgt 1140
tgcttctatc caataatgta cgtgtgggtgt ttacacgcga atgtcttgta gacctttgta 1200
tgtgtacaag tgaattttaa ttcacaagta catataatgg tcaccattga aaagatgttt 1260
agtgtgtgtt ttccaatata tgtttggtga aggttcatca tctaataaaa tatgtttgga 1320
acccaaaaaa aaaaaaaaaa aa 1342

```

<210> SEQ ID NO 5

<211> LENGTH: 335

<212> TYPE: PRT

<213> ORGANISM: Hordeum vulgare

<400> SEQUENCE: 5

```

Met Ala Ala Gln Asn Asn Asn Lys Asp Val Ala Ala Leu Val Glu Lys
1           5           10           15
Ile Thr Gly Leu His Ala Ala Ile Ala Lys Leu Pro Ser Leu Ser Pro
20          25          30
Ser Pro Asp Val Asp Ala Leu Phe Thr Glu Leu Val Thr Ala Cys Val
35          40          45
Pro Pro Ser Pro Val Asp Val Thr Lys Leu Gly Pro Glu Ala Gln Glu
50          55          60
Met Arg Glu Gly Leu Ile Arg Leu Cys Ser Glu Ala Glu Gly Lys Leu
65          70          75          80
Glu Ala His Tyr Ser Asp Met Leu Ala Ala Phe Asp Asn Pro Leu Asp
85          90          95
His Leu Gly Ile Phe Pro Tyr Tyr Ser Asn Tyr Ile Asn Leu Ser Lys
100         105         110
Leu Glu Tyr Glu Leu Leu Ala Arg Tyr Val Arg Arg His Arg Pro Ala
115         120         125
Arg Val Ala Phe Ile Gly Ser Gly Pro Leu Pro Phe Ser Ser Phe Val
130         135         140
Leu Ala Ala Arg His Leu Pro Asp Thr Met Phe Asp Asn Tyr Asp Leu
145         150         155         160
Cys Gly Ala Ala Asn Asp Arg Ala Ser Lys Leu Phe Arg Ala Asp Thr
165         170         175
Asp Val Gly Ala Arg Met Ser Phe His Thr Ala Asp Val Ala Asp Leu
180         185         190
Ala Ser Glu Leu Ala Lys Tyr Asp Val Val Phe Leu Ala Ala Leu Val
195         200         205
Gly Met Ala Ala Glu Asp Lys Ala Lys Val Ile Ala His Leu Gly Ala
210         215         220
His Met Ala Asp Gly Ala Ala Leu Val Val Arg Ser Ala His Gly Ala
225         230         235         240
Arg Gly Phe Leu Tyr Pro Ile Val Asp Pro Gln Asp Ile Gly Arg Gly
245         250         255
Gly Phe Glu Val Leu Ala Val Cys His Pro Asp Asp Asp Val Val Asn
260         265         270
Ser Val Ile Ile Ala Gln Lys Ser Lys Glu Val His Ala Asp Gly Leu
275         280         285

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Gly Ser Ala Arg Gly Ala Gly Arg Gln Tyr Ala Arg Gly Thr Val Pro
 290 295 300

Val Val Ser Pro Pro Cys Arg Phe Gly Glu Met Val Ala Asp Val Thr
 305 310 315 320

Gln Asn His Lys Arg Asp Glu Phe Ala Asn Ala Glu Val Ala Phe
 325 330 335

<210> SEQ ID NO 6
 <211> LENGTH: 1314
 <212> TYPE: DNA
 <213> ORGANISM: Hordeum vulgare

<400> SEQUENCE: 6

```

ctacttcact cacactagtg cccagaaaga aggctgcaat ggctgcccag aacaacaaca    60
aggatgtcgc tgccctgggtg gagaagatca ccgggctcca cgccgccatc gccaaagtgc    120
cgtcgctcag cccatccccg gacgtcgacg cgctcttcac cgagctggtc acggcgtgcg    180
ttcccccgag ccccgtagac gtgaccaagc tcggccccga ggcgcaggag atgcggggagg    240
gcctcatccg cctctgtctc gaggccgagg ggaagtggga ggcgcactac tccgacatgc    300
tcggcgcctt cgacaacccg ctggatcacc tcggcatctt cccctactac agcaactaca    360
tcaacctcag caagtggag tacgagctcc tggcacgcta cgtccggcgg catcgcccgg    420
cccgcgtcgc gttcatcggc tccggcccgc tgcggttcag ctcccttctc ctggcgcgcg    480
gccacctgcc cgacacatg tttgacaact acgacctttg cggcgcgccc aacgatcgcg    540
ccagcaagct cttccgcgcg gacacggacg tgggtgcccc catgtcgttc cacacggccc    600
acgtcgcgga cctcgccagc gagctcgcca agtacgacgt cgtcttctct gcccgcctcg    660
tcggcatggc cgccgaggac aaggccaagg tgatcgcgca cctcgggcga cacatggcag    720
acggggcggc cctcgctcgt cgcagcgcac acggagcgcg cgggttctctg taccgattg    780
tcgacccccg ggacatcggc cgcggcgggt tcgaggtgct ggccgtgtgc caccgccgacg    840
acgacgtggt gaactccgtc atcatcgcac agaagtccaa ggaggtgcat gccgatggac    900
ttggcagcgc gcgtggtgcc ggtcgacagt acgcgcgcg caccggtgccg gttgtcagcc    960
ccccgtgcag gttcgggtgag atggtggcgg atgtgaccca gaaccacaag agagacgagt   1020
ttgccaacgc cgaagtggcc ttttgatcga tcgtcgccaa gggacaataa atgaactggtg   1080
atgtggttagg gtaatttgc tacctcgctg cttgatcgct tgcaatatgt gcacattttc   1140
ctactaccgc tgcttatgca tttcaagcca tgtgatgttg gtatccaata aagtatgtgt   1200
agggtttaca cgcaaatgtc tttacacctt gtacgtgtaa gtgttgacaa cgatgaattt   1260
cagttcacia ttaataaata gtataatgga ttcaaaaaaa aaaaaaaaaa aaaa         1314

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<210> SEQ ID NO 7
 <211> LENGTH: 329
 <212> TYPE: PRT
 <213> ORGANISM: Hordeum vulgare

<400> SEQUENCE: 7

Met Asp Gly Gln Ser Glu Glu Val Asp Ala Leu Val Gln Lys Ile Thr
 1 5 10 15

Gly Leu His Ala Ala Ile Ala Lys Leu Pro Ser Leu Ser Pro Ser Pro
 20 25 30

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Asp Val Asp Ala Leu Phe Thr Asp Leu Val Thr Ala Cys Val Pro Pro
 35 40 45
 Ser Pro Val Asp Val Thr Lys Leu Ala Pro Glu Ala Gln Ala Met Arg
 50 55 60
 Glu Gly Leu Ile Arg Leu Cys Ser Glu Ala Glu Gly Lys Leu Glu Ala
 65 70 75 80
 His Tyr Ser Asp Met Leu Ala Ala Phe Asp Asn Pro Leu Asp His Leu
 85 90 95
 Gly Val Phe Pro Tyr Tyr Ser Asn Tyr Ile Asn Leu Ser Lys Leu Glu
 100 105 110
 Tyr Glu Leu Leu Ala Arg Tyr Val Pro Gly Arg His Arg Pro Ala Arg
 115 120 125
 Val Ala Phe Ile Gly Ser Gly Pro Leu Pro Phe Ser Ser Tyr Val Leu
 130 135 140
 Ala Ala Arg His Leu Pro Asp Thr Val Phe Asp Asn Tyr Asp Leu Cys
 145 150 155 160
 Gly Ala Ala Asn Asp Arg Ala Thr Arg Leu Phe Arg Ala Asp Lys Asp
 165 170 175
 Val Gly Ala Arg Met Ser Phe His Thr Ala Asp Val Ala Asp Leu Thr
 180 185 190
 Asp Glu Leu Ala Thr Tyr Asp Val Val Phe Leu Ala Ala Leu Val Gly
 195 200 205
 Met Ala Ala Glu Asp Lys Ala Lys Val Ile Ala His Leu Gly Ala His
 210 215 220
 Met Ala Asp Gly Ala Ala Leu Val Ala Arg His Gly Ala Arg Gly Phe
 225 230 235 240
 Leu Tyr Pro Ile Val Asp Pro Gln Asp Ile Gly Arg Gly Gly Phe Glu
 245 250 255
 Val Leu Ala Val Cys His Pro Asp Asp Asp Val Val Asn Ser Val Ile
 260 265 270
 Ile Ala Gln Lys Ser Asn Asp Val His Glu Tyr Gly Leu Gly Ser Gly
 275 280 285
 Arg Gly Gly Arg Tyr Ala Arg Gly Thr Val Val Pro Val Val Ser Pro
 290 295 300
 Pro Cys Arg Phe Gly Glu Met Val Ala Asp Val Thr Gln Lys Arg Glu
 305 310 315 320
 Glu Phe Ala Asn Ala Glu Val Ala Phe
 325

<210> SEQ ID NO 8

<211> LENGTH: 1249

<212> TYPE: DNA

<213> ORGANISM: Hordeum vulgare

<400> SEQUENCE: 8

```

ccactaccga ctaccgtagt accgtgcttc agagctcadc actggtcagg taccaagaag    60
acataaaaat ggacggccag agcgaggagg tgcagccct tgtccagaag atcaccggcc    120
tccacgccgc catcgccaag ctgccctgc tcagcccgtc cccggacgtc gacgcgtct    180
tcaccgacct ggtcaccgcy tgcgtgcccc cgagccccgt ggacgtgacc aagctcgccc    240
cggaggcgca ggcgatgcyg gagggcctca tccgcctctg ctccgaggcc gagggcaagc    300
tggaggcgca ctactccgac atgctcgccg ccttcgacaa cccgctcgac cacctcggcg    360

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tcttccccta ctacagcaac tacatcaacc tcagcaagct tgagtacgag ctctctcgcg 420
gtactcgtgcc cggcaggcat cgcccggccc gcgtcgccct catcggtccc ggcccgtgce 480
cgttcagctc ctactgcctc gccgcgcgcc acctgcccga caccgtgttc gacaactacg 540
acctgtgcgg cgccggccaac gaccgcgcga ccaggctgtt ccgcgcggac aaggacgtcg 600
gcgcccgcac gtcgttccac accgccgacg tcgctggacct caccgacgag ctcgctacgt 660
acgacgtcgt cttcctggcc gcgctcgtgg gcatggccgc cgaggacaag gccaaagtga 720
tcgcgcaact tggcgcgcac atggcggacg gggcggccct cgttgcgcgg caeggcgcgc 780
gtgggttccct ctaccgacg gtcgatcccc aggacatcgg tcgaggcggg ttcgaggtgc 840
tcgcccgtgtg tcaccccgcac gacgacgtgg tgaactccgt catcatcgca caaaagagca 900
acgacgtgca cgagtatgga cttggcagcg ggcgtggtgg acggtacgcg cgaggcacgg 960
tggtgccggg ggtcagccca cctgcaggt tcggcgagat ggtggcagac gtgaccaga 1020
agagagagga gtttgccaac gcggaagtgg ccttctgatt gctgctgaat cgcttctgat 1080
cgtacgtggt aatTTTTCTA ctactcctcc tctaccacc acctatcacc tatgtatgca 1140
tttcaagtcg tgtgtgtttt gtatccaata atgtaagtga gatgtttaca cgcgcaaaaa 1200
aaaaaaaaa aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa 1249

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<210> SEQ ID NO 9

<211> LENGTH: 282

<212> TYPE: PRT

<213> ORGANISM: *Hordeum vulgare*

<400> SEQUENCE: 9

```

Met Glu Ala Glu Asn Gly Glu Val Ala Ala Leu Val Glu Lys Ile Thr
 1             5             10             15
Gly Leu His Ala Ala Ile Ser Lys Leu Pro Ala Leu Ser Pro Ser Pro
20             25             30
Gln Val Asp Ala Leu Phe Thr Glu Leu Val Ala Ala Cys Val Pro Ser
35             40             45
Ser Pro Val Asp Val Thr Lys Leu Gly Pro Glu Ala Gln Glu Met Arg
50             55             60
Gln Asp Leu Ile Arg Leu Cys Ser Ala Ala Glu Gly Leu Leu Glu Ala
65             70             75             80
His Tyr Ser Asp Met Leu Thr Ala Leu Asp Ser Pro Leu Asp His Leu
85             90             95
Gly Arg Phe Pro Tyr Phe Asp Asn Tyr Val Asn Leu Ser Lys Leu Glu
100            105            110
His Asp Leu Leu Ala Gly His Val Ala Ala Pro Ala Arg Val Ala Phe
115            120            125
Ile Gly Ser Gly Pro Leu Pro Phe Ser Ser Leu Phe Leu Ala Thr Tyr
130            135            140
His Leu Pro Asp Thr Arg Phe Asp Asn Tyr Asp Arg Cys Ser Val Ala
145            150            155            160
Asn Gly Arg Ala Met Lys Leu Val Gly Ala Ala Asp Glu Gly Val Arg
165            170            175
Ser Arg Met Ala Phe His Thr Ala Glu Val Thr Asp Leu Thr Ala Glu
180            185            190
Leu Gly Ala Tyr Asp Val Val Phe Leu Ala Ala Leu Val Gly Met Thr

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195 200 205

Ser Lys Glu Lys Ala Asp Ala Ile Ala His Leu Gly Lys His Met Ala
210 215 220

Asp Gly Ala Val Leu Val Arg Glu Ala Leu His Gly Ala Arg Ala Phe
225 230 235 240

Leu Tyr Pro Val Val Glu Leu Asp Asp Val Gly Arg Gly Gly Phe Gln
245 250 255

Val Leu Ala Val His His Pro Ala Gly Asp Glu Val Phe Asn Ser Phe
260 265 270

Ile Val Ala Arg Lys Val Lys Met Ser Ala
275 280

<210> SEQ ID NO 10
 <211> LENGTH: 1044
 <212> TYPE: DNA
 <213> ORGANISM: Hordeum vulgare

<400> SEQUENCE: 10

```

gtgacatgga ggccgaaaac ggcgaggtgg ctgctctggt cgagaagatc accggtctcc      60
acgccgccat ctccaagctc ccggcactaa gcccgctctcc tcaagtcgac gcgctcttca      120
ccgagctggt tgcggcgtgc gtcccatcaa gcccggtgga cgtgaccaag ctcggcccgg      180
aggcgcagga gatcgcgcag gacctcatcc gtctctgctc ggccgcccag gggctgctcg      240
aggcgcacta ctccgacatg ctcaccgctg tggacagccc gctcggaccac ctcggcccgt      300
tccttactt  cgacaactac gtcaacctca gcaagctcga gcaagatctt ctggcaggtc      360
acgtggcggc cccggcccgc gtggcggttca tggggtcggg gccactgccg ttcagctcgc      420
tcttccttgc gacgtaccac ctgccggaca cccggttcga caactacgac cgggtgcagcg      480
tggcgaatgg ccgggcgatg aagctggtcg gcgcggcgga cgagggcgtg cgatcacgca      540
tggcggtcca cacggccgaa gtcacggacc tcacggctga gctcggcgct tacgacgtgg      600
tcttcctggc cgcgctcgtg ggaatgacgt ccaaggagaa ggccgacgcc atagcgcact      660
tggggaagca catggcagat ggggcggtgc tcgtgcgcga agcgtgcac ggggcgcgag      720
cgttcctgta tcctgtcgtg gagctggacg atgtcgggcg tgggtgggttc caagtgtggt      780
ccgtgcacca cctcgcaggc gatgaggtgt tcaactcatt catagttgcc cggaagtgta      840
aatgagtgc  ttaaattaag aaaaggtgga gcctgtctgc ttgtgcaaat ggtgtctcac      900
attgataata accagatgat accctgcaca ttgatggggg tactgcagta tgtttcaatg      960
aggtctggtt gtatcaaata tgagtatttg gcttaataat atcagcgaat atgtttogat     1020
taaaaaaaaa aaaaaaaaaa aaaa                                             1044

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<210> SEQ ID NO 11
 <211> LENGTH: 328
 <212> TYPE: PRT
 <213> ORGANISM: Hordeum vulgare

<400> SEQUENCE: 11

Met Asp Ala Gln Asn Lys Glu Val Asp Ala Leu Val Gln Lys Ile Thr
1 5 10 15

Gly Leu His Ala Ala Ile Ala Lys Leu Pro Ser Leu Ser Pro Ser Pro
20 25 30

Asp Val Asp Ala Leu Phe Thr Asp Leu Val Thr Ala Cys Val Pro Pro

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```

tacatcaacc tcagcaagct ggagtacgag ctccctggcgc gctacgtgcc gggcggcacc 420
gcccggcccc ctgtcgcggt catcggtccc gcccgcgtgc cgttcagctc ctacgtcctc 480
gcccgtcgcc acctgcccga cgcgatgttc gacaactacg acctgtgtag cgcggccaac 540
gaccgtgcga gcaagctggt ccgcgccggac aaggacgtgg gcgcccgcac gtctttccac 600
accgcccagc tagcggacct caccgcgag ctccgcgcgt acgacgtcgt cttcctggcc 660
gcgctcgtgg gcacgggtgc cgaggacaag gccaaggtga ttccgcacct cggcgcgcac 720
atggcggacg gggcgccctc cgtcgtgcgc agtgcgcagg cacgtggggt cctctacccg 780
atcgtcgatc cccaggacat cgtcgcaggc gggtttgagg tgctggccgt gtgtcacccc 840
gacgatgacg tggtaactc cgtcatcacc gcacacaagt ccaaggacgt gcacgccaat 900
gaacgtccca acggcgctgg tggacagtac cggggcgcgg taccgggtgt cagcccgcgc 960
tgcaggttcg gtgagatggt ggcggacgtg acccacaaga gagaggagt caccaacgcg 1020
gaagtggcct tctgatcgtt gcgagggaat gaaaatgaag gtggacgtgt gtggtcagca 1080
tccatcagtg gctgcctgct tcacgccttg caatcgtact actacctacc tatgcagttc 1140
aagtcagtgt ttgtcaatgt aagtgtgatg ttactactag tctatgaaag gcagggcaga 1200
cgagggtagt gtgccaagta acagtggtgc attataggtg taagtgttga gaataagacc 1260
atctttgttc acaaatagta tgatgtaac ggtgtcatat tcgtattgag tacatttgtc 1320
aagttggttg ctaaaaaaaaa aaaaaaaaaa aa 1352

```

<210> SEQ ID NO 13

<211> LENGTH: 329

<212> TYPE: PRT

<213> ORGANISM: Hordeum vulgare

<400> SEQUENCE: 13

```

Met Asp Ala Gln Ser Lys Glu Val Asp Ala Leu Val Gln Lys Ile Thr
 1          5          10          15

Gly Leu His Ala Ala Ile Ala Lys Leu Pro Ser Leu Ser Pro Ser Pro
20          25          30

Asp Val Asp Ala Leu Phe Thr Asp Leu Val Thr Ala Cys Val Pro Pro
35          40          45

Ser Pro Val Asp Val Thr Lys Leu Ala Pro Glu Ala Gln Ala Met Arg
50          55          60

Glu Gly Leu Ile Arg Leu Cys Ser Glu Ala Glu Gly Lys Leu Glu Ala
65          70          75          80

His Tyr Ser Asp Met Leu Ala Ala Phe Asp Asn Pro Leu Asp His Leu
85          90          95

Gly Val Phe Pro Tyr Tyr Ser Asn Tyr Ile Asn Leu Ser Lys Leu Glu
100         105         110

Tyr Glu Leu Leu Ala Arg Tyr Val Pro Gly Gly Ile Ala Pro Ala Arg
115         120         125

Val Ala Phe Ile Gly Ser Gly Pro Leu Pro Phe Ser Ser Tyr Val Leu
130         135         140

Ala Ala Arg His Leu Pro Asp Thr Val Phe Asp Asn Tyr Val Pro Val
145         150         155         160

Arg Ala Ala Asn Asp Arg Ala Thr Arg Leu Phe Arg Ala Asp Lys Asp
165         170         175

Val Gly Ala Arg Met Ser Phe His Thr Ala Asp Val Ala Asp Leu Thr

```

-continued

180	185	190
Asp Glu Leu Ala Thr Tyr Asp Val Val Phe Leu Ala Ala Leu Val Gly		
195	200	205
Met Ala Ala Glu Asp Lys Gly Gln Gly Asp Pro His Leu Gly Ala His		
210	215	220
Met Ala Asp Gly Ala Ala Leu Val Arg Ser Ala His Gly Ala Arg Gly		
225	230	235 240
Phe Leu Tyr Pro Ile Val Asp Pro Gln Asp Ile Gly Arg Gly Gly Phe		
245	250	255
Glu Val Leu Ala Val Cys His Pro Asp Asp Asp Val Val Asn Ser Val		
260	265	270
Ile Ile Ala Gln Lys Ser Lys Asp Met Phe Ala Asn Gly Pro Arg Asn		
275	280	285
Gly Cys Gly Gly Arg Tyr Ala Arg Gly Thr Val Pro Val Val Ser Pro		
290	295	300
Pro Cys Arg Phe Gly Glu Met Val Ala Asp Val Thr Gln Lys Arg Glu		
305	310	315 320
Glu Phe Ala Lys Ala Glu Val Ala Phe		
325		

<210> SEQ ID NO 14
 <211> LENGTH: 1371
 <212> TYPE: DNA
 <213> ORGANISM: Hordeum vulgare
 <220> FEATURE:
 <221> NAME/KEY: modified_base
 <222> LOCATION: (8)
 <223> OTHER INFORMATION: a, c, g, t, unknown or other

<400> SEQUENCE: 14

```

ggagcggnac gcgtggcgga ggtgggcact accgtagtac cgtgcctcag agctcatcac      60
tggtcaggta ccaagaagac ataaaaatgg acgccagag caaggaggtc gacgcccttg      120
tccagaagat caccggcctc cacgccgcca tcgccaagct gccctcgctc agcccgtccc      180
cggacgtcga cgcgctcttc accgacctgg tcaccgctg cgtgcccccg agcccgtgg      240
acgtgaccaa gctcgccccg gaggcgacag cgatgcggga gggcctcacc cgcctctgct      300
ccgagggcca gggcaagctg gagggcact actccgacat gctcgccgcc ttcgacaacc      360
cgctcgacca cctcggcgtc tcccctact acagcaacta catcaacctc agcaagctcg      420
agtacgagct cctcgcgcgc tacgtgcccg gcggcatcgc cccggcccgc gtcgcttca      480
tcggctccgg cccgctcccg ttcagctcct acgtcctcgc cgcgcgccac ctgcccgaca      540
ccgtgttcga caactacgta cctgtgcgcg cggccaacga ccgcgcgacc aggctgttcc      600
gcgcggaaca ggacgtcggc gcccgcatgt cgttccacac cgccgacgtc ggggacctca      660
ccgacgagct cgctacgtac gacgtcgtct tcctggccgc gctcgtgggc atggccgccc      720
aggacaaggg ccaaggtgat ccgcacctg gcgcgcacat ggcgacggg gcggccctcg      780
tccgcagcgc gcacggggcg cgtgggttcc tetaccgat cgtcgatccc caagacattg      840
gtcgaggcgg gttcagaggc ctcgccgtgt gtcaccccga cgacgacgtg gtgaactccg      900
tcatcatcgc gcagaagtct aaggacatgt ttgccaatgg acctcgcaac ggggtgtggtg      960
gacggtacgc gcgaggcacg gtcggggtg tcagcccgcc ctgcaggttc ggcgagatgg     1020
tggcagacgt gaccocagaag agagaggagt ttgccaggc ggaagtggcc ttctgattgc     1080
  
```

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```

tcgcagggtca ccatccgtat gccgctgcta cctttcaata tcttgcaatc gtaggtggcg 1140
atcttcttac tcttgttacg acctttcaaa tcatatgttg tttgtaccca ataagtgaag 1200
tgtgttgctt acacgcgcat gtcttgtaga ctcggtctct agaaggcagg gcagatcaag 1260
agactgtgca aaggaaaaga aatgtgtgtt gttgtaggtg tatgagttgg gagtaagatg 1320
attctagttc acaaaaaaaaa aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa a 1371

```

```

<210> SEQ ID NO 15
<211> LENGTH: 324
<212> TYPE: PRT
<213> ORGANISM: Oryza sativa

```

```

<400> SEQUENCE: 15

```

```

Met Glu Ala Gln Asn Gln Glu Val Ala Ala Leu Val Glu Lys Ala Gly
 1           5           10           15
Leu His Ala Ala Ser Lys Leu Pro Ser Leu Ser Pro Ser Ala Glu Val
20           25           30
Asp Ala Leu Phe Thr Asp Leu Val Thr Ala Cys Val Pro Ala Ser Pro
35           40           45
Val Asp Val Ala Lys Leu Gly Pro Glu Ala Gln Ala Met Arg Glu Glu
50           55           60
Leu Arg Leu Cys Ser Ala Ala Glu Gly His Leu Glu Ala His Tyr Ala
65           70           75           80
Asp Met Leu Ala Ala Phe Asp Asn Pro Leu Asp His Leu Ala Arg Phe
85           90           95
Pro Tyr Tyr Gly Asn Tyr Val Asn Leu Ser Lys Leu Glu Tyr Asp Leu
100          105          110
Leu Val Arg Tyr Val Pro Gly Ala Pro Thr Arg Val Ala Phe Val Gly
115          120          125
Ser Gly Pro Leu Pro Phe Ser Ser Leu Val Leu Ala Ala His His Leu
130          135          140
Pro Asp Ala Val Phe Asp Asn Tyr Asp Arg Cys Gly Ala Ala Asn Glu
145          150          155          160
Arg Ala Arg Arg Leu Phe Arg Gly Ala Asp Glu Gly Leu Gly Ala Arg
165          170          175
Met Ala Phe His Thr Ala Asp Val Ala Thr Leu Thr Gly Glu Leu Gly
180          185          190
Ala Tyr Asp Val Val Phe Leu Ala Ala Leu Val Gly Met Ala Ala Glu
195          200          205
Glu Lys Ala Gly Val Ala His Leu Gly Ala His Met Ala Asp Gly Ala
210          215          220
Ala Leu Val Val Arg Thr Ala His Gly Ala Arg Gly Phe Leu Tyr Pro
225          230          235          240
Val Asp Pro Glu Asp Val Arg Arg Gly Gly Phe Asp Val Leu Ala Val
245          250          255
Cys His Pro Glu Asp Glu Val Asn Ser Val Val Ala Arg Lys Val Gly
260          265          270
Ala Ala Ala Ala Ala Ala Ala Ala Arg Arg Asp Glu Leu Ala Asp Ser
275          280          285
Arg Gly Val Val Leu Pro Val Val Gly Pro Pro Ser Thr Cys Cys Lys
290          295          300

```

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Val Glu Ala Ser Ala Val Glu Lys Ala Glu Glu Phe Ala Ala Asn Lys
305 310 315 320

Glu Leu Ser Val

<210> SEQ ID NO 16
<211> LENGTH: 1372
<212> TYPE: DNA
<213> ORGANISM: *Oryza sativa*

<400> SEQUENCE: 16

```

ctccatttgg ttgtcatttt caactataat ccaccacaac tcgtgcaaca tcagctcact    60
cgtgttccca accgcgacaa agcttcacag atggaggctc agaaccaaga ggtcgctgcc    120
ctggtcgaga agatcgccgg cctccacgcc gccatctcca agctgcccgc gctgagccca    180
tccgccgagg tggacgcgct cttcaccgac ctcgtcacgg cgtgctccc ggcgagcccc    240
gtcgacgtgg ccaagctcgg cccggaggcg caggcgtatgc gggaggagct catccgctc    300
tgctccgccc ccgagggcca cctcgaggcg cactacgccc acatgctcgc cgcttcgac    360
aaccgcctcg accacctcgc ccgcttcccg tactacggca actacgtcaa cctgagcaag    420
ctggagtacg acctcctcgt ccgctacgtc cccggcattg cccccaccg cgtcgcttc    480
gtcgggtcgg gcccgctgcc gttcagctcc ctcgctcctc ctgcccacca cctgcccggc    540
gcggtgttcc acaactacga ccgggtcggc gcggccaacg agcgggagag gaggtgttc    600
cgcggcgccc acgagggcct cggcgccgcg atggcgctcc acaccgccga cgtggcgacc    660
ctgacggggg agctcgccgc gtacgacgtc gtgttcctgg cggcgctcgt gggcatggcg    720
gccgaggaga aggccggggt gatcgcgcac ctgggcgccc acatggcgga cggcgcgggc    780
ctcgctcgtc ggacggcgca cggggcgccc ggggtcctgt acccgatcgt cgatcccag    840
gacgtcaggc gtggcggggt cgacgttctg gcggtgtgcc acccgagga cgaggtgatc    900
aactccgtca tcgtcgcccg caaggtcggg gccgcgccc ccgcgcgccc ggcgcgcaga    960
gacgagctcg cggactcgcg cggcgtgggt ctgccggtgg tcgggcccgc gtcccagtc    1020
tgcaaggtgg aggcgagcgc ggttgagaag gcagaagagt ttgcccaca caaggagctg    1080
tccgtctaac agcccgaaga tcgaaaggcg cactatatta tggcaataaa tcatttgatt    1140
atacttatgc tgcatttgcg aagctaaggt atactatgca agccatatgt ttgtgttcgt    1200
acgtgttgtt tgggacgtac agttgtgttg ttgtacgtcg tgaagtactg aagtgttcac    1260
agtagatcac aagttcacag caatcaatga ggacctgta agccagtgt aacgaggaac    1320
atgccatctg tgtatgacag tgagaaatta tataagaaaa acattttgtg ac    1372

```

<210> SEQ ID NO 17
<211> LENGTH: 320
<212> TYPE: PRT
<213> ORGANISM: *Arabidopsis thaliana*

<400> SEQUENCE: 17

```

Met Ala Cys Gln Asn Asn Leu Val Val Lys Gln Ile Ile Asp Leu Tyr
 1          5          10          15
Asp Gln Ile Ser Lys Leu Lys Ser Leu Lys Pro Ser Lys Asn Val Asp
 20          25          30
Thr Leu Phe Gly Gln Leu Val Ser Thr Cys Leu Pro Thr Asp Thr Asn
 35          40          45

```

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Ile	Asp	Val	Thr	Asn	Met	Cys	Glu	Glu	Val	Lys	Asp	Met	Arg	Ala	Asn
50					55					60					
Leu	Ile	Lys	Leu	Cys	Gly	Glu	Ala	Glu	Gly	Tyr	Leu	Glu	Gln	His	Phe
65					70					75					80
Ser	Thr	Ile	Leu	Gly	Ser	Leu	Gln	Glu	Asp	Gln	Asn	Pro	Leu	Asp	His
85					90					95					
Leu	His	Ile	Phe	Pro	Tyr	Tyr	Ser	Asn	Tyr	Leu	Lys	Leu	Gly	Lys	Leu
100					105					110					
Glu	Phe	Asp	Leu	Leu	Ser	Gln	His	Ser	Ser	His	Val	Pro	Thr	Lys	Ile
115					120					125					
Ala	Phe	Val	Gly	Ser	Gly	Pro	Met	Pro	Leu	Thr	Ser	Ile	Val	Leu	Ala
130					135					140					
Lys	Phe	His	Leu	Pro	Asn	Thr	Thr	Phe	His	Asn	Phe	Asp	Ile	Asp	Ser
145					150					155					160
His	Ala	Asn	Thr	Leu	Ala	Ser	Asn	Leu	Val	Ser	Arg	Asp	Pro	Asp	Leu
165					170					175					
Ser	Lys	Arg	Met	Ile	Phe	His	Thr	Thr	Asp	Val	Leu	Asn	Ala	Thr	Glu
180					185					190					
Ala	Leu	Asp	Gln	Tyr	Asp	Val	Val	Phe	Leu	Ala	Ala	Leu	Val	Gly	Met
195					200					205					
Asp	Lys	Glu	Ser	Lys	Val	Lys	Ala	Ile	Glu	His	Leu	Glu	Lys	His	Met
210					215					220					
Ala	Pro	Gly	Ala	Val	Leu	Met	Leu	Arg	Arg	Ala	His	Ala	Leu	Arg	Ala
225					230					235					240
Phe	Leu	Tyr	Pro	Ile	Val	Asp	Ser	Ser	Asp	Leu	Lys	Gly	Phe	Gln	Leu
245					250					255					
Leu	Thr	Ile	Tyr	His	Pro	Thr	Asp	Asp	Val	Val	Asn	Ser	Val	Val	Ile
260					265					270					
Ala	Arg	Lys	Leu	Gly	Gly	Pro	Thr	Thr	Pro	Gly	Val	Asn	Gly	Thr	Arg
275					280					285					
Gly	Cys	Met	Phe	Met	Pro	Cys	Asn	Cys	Ser	Lys	Ile	His	Ala	Ile	Met
290					295					300					
Asn	Asn	Arg	Gly	Lys	Lys	Asn	Met	Ile	Glu	Glu	Phe	Ser	Thr	Ile	Glu
305					310					315					320

<210> SEQ ID NO 18

<211> LENGTH: 963

<212> TYPE: DNA

<213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 18

atggccttgcc	aaaacaatct	cgttgtgaag	caaatcatcg	acttgtagca	ccaaatctca	60
aagctcaaga	gcttaaaacc	ttccaaaaat	gtcgacactt	tgttcggaca	actcgtgtcc	120
acgtgcttac	ccacggatag	aaacatcgat	gtcacaataa	tgtgtgaaga	agtcaaagac	180
atgagagcta	atctcatcaa	gctttgtggt	gaagccgaag	gttatttggg	gcaaaccttc	240
tccacaattt	tgggatcttt	acaagaagac	caaaaccac	ttgaccattt	acacatcttt	300
ccttactact	ccaactacct	caagctaggc	aagctcgagt	tcgatctcct	gagccaacac	360
tcaagccatg	tccccaccaa	gattgccttc	gtgggttcgg	gtccgatgcc	tctcacatcc	420
atcgtattgg	ccaagtttca	cctcccacac	acgacgttcc	acaactttga	catcgactca	480
cacgcaaaca	cactcgtctc	aaactcgtc	tctcgcgacc	cggacctctc	aaaacgatg	540

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```

atcttcacaca caacggacgt actaaacgca accgaagccc ttgaccaata tgacgtcgtt    600
ttcttagcgg cgctttagg gatggacaaa gagtcaaagg tcaaagccat cgagcacttg    660
gagaaacaca tggctcctgg agctgttctt atgctaagga gggctcatgc tctcagagct    720
ttcttatatc caatcgttga ctgcgtctgat ctcaaaggct ttcaactctt gaccatctat    780
catccaaccg atgacgtggt taactcgggt gtgatcgcac gtaagctcgg tggtcggacc    840
acgccccggg ttaatggtac tcgtggatgc atgtttatgc cttgtaactg ctccaagatt    900
cacgcatca tgaacaaccg tgtaagaag aatatgatcg aggagttag taccatcgag    960
taa                                                                 963

```

<210> SEQ ID NO 19

<211> LENGTH: 320

<212> TYPE: PRT

<213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 19

```

Met Ala Cys Gln Asn Asn Leu Val Val Lys Gln Ile Met Asp Leu Tyr
1           5           10           15
Asn Gln Ile Ser Asn Leu Glu Ser Leu Lys Pro Ser Lys Asn Val Asp
20          25          30
Thr Leu Phe Arg Gln Leu Val Ser Thr Cys Leu Pro Thr Asp Thr Asn
35          40          45
Ile Asp Val Thr Glu Ile His Asp Glu Lys Val Lys Asp Met Arg Ser
50          55          60
His Leu Ile Lys Leu Cys Gly Glu Ala Glu Gly Tyr Leu Glu Gln His
65          70          75          80
Phe Ser Ala Ile Leu Gly Ser Phe Glu Asp Asn Pro Leu Asn His Leu
85          90          95
His Ile Phe Pro Tyr Tyr Asn Asn Tyr Leu Lys Leu Gly Lys Leu Glu
100         105         110
Phe Asp Leu Leu Ser Gln His Thr Thr His Val Pro Thr Lys Val Ala
115         120         125
Phe Ile Gly Ser Gly Pro Met Pro Leu Thr Ser Ile Val Leu Ala Lys
130         135         140
Phe His Leu Pro Asn Thr Thr Phe His Asn Phe Asp Ile Asp Ser His
145         150         155         160
Ala Asn Thr Leu Ala Ser Asn Leu Val Ser Arg Asp Ser Asp Leu Ser
165         170         175
Lys Arg Met Ile Phe His Thr Thr Asp Val Leu Asn Ala Lys Glu Gly
180         185         190
Leu Asp Gln Tyr Asp Val Val Phe Leu Ala Ala Leu Val Gly Met Asp
195         200         205
Lys Glu Ser Lys Val Lys Ala Ile Glu His Leu Glu Lys His Met Ala
210         215         220
Pro Gly Ala Val Val Met Leu Arg Ser Ala His Gly Leu Arg Ala Phe
225         230         235         240
Leu Tyr Pro Ile Val Asp Ser Cys Asp Leu Lys Gly Phe Glu Val Leu
245         250         255
Thr Ile Tyr His Pro Ser Asp Asp Val Val Asn Ser Val Val Ile Ala
260         265         270

```

-continued

Arg Lys Leu Gly Gly Ser Asn Gly Ala Arg Gly Ser Gln Ile Gly Arg
 275 280 285

Cys Val Val Met Pro Cys Asn Cys Ser Lys Val His Ala Ile Leu Asn
 290 295 300

Asn Arg Gly Met Glu Lys Asn Leu Ile Glu Glu Phe Ser Ala Ile Glu
 305 310 315 320

<210> SEQ ID NO 20
 <211> LENGTH: 963
 <212> TYPE: DNA
 <213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 20

```

atggcttgcc aaaacaatct cgttgtgaag caaatcatgg acttatacaa ccaaattcca    60
aacctcgaga gcttaaaacc atccaagaat gtcgacactt tggtcagaca acttgtgtcc    120
acgtgcttac caacggacac gaacatcgat gtcacagaga tacacgatga aaaagtcaaa    180
gacatgagat ctcatctcat caagctttgt ggtgaagccg aaggttattt agagcaacac    240
ttttcagcaa tcttaggctc ttttgaagac aaccctctaa accatttaca catcttcccc    300
tattacaaca actatctcaa actaggcaaa ctcgaaatcg atctcctttc tcagcacaca    360
acccatgtcc cgaccaaaagt cgcctttatt ggttcoggtc cgatgccact tacttccatc    420
gtcttgGCCa agttccacct ccccaacaca acgttccaca acttcogacat cgactcacac    480
gccaaacacac tcgcttcaaa cctcgtttct cgtgattctg acctttccaa acgcatgatt    540
ttccacacaa ctgatgtatt aaacgctaag gaggggtag accaatacga tgttgttttc    600
ttggcagctc ttgttgggat ggataaagag tcaaaggtea aagctattga gcatttagag    660
aagcatatgg cccttgagc tgtggtgatg ctaagaagtg ctcatggtct tagagctttc    720
ttgatccaa tcggtgactc ttgtgatcct aaagggttg aggtgtaac catttatcat    780
ccgtctgacg acgtggtaa ttcggtggtc atcgcacgta agcttggtgg ttcaaatgga    840
gctcgaggca gccagatcgg acggtgtgtg gttatgcctt gtaattgctc taaggccac    900
gcatcttga acaatcgtgg tatggagaag aatttgatcg aggagttag tgccatogag    960
taa
  
```

<210> SEQ ID NO 21
 <211> LENGTH: 320
 <212> TYPE: PRT
 <213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 21

Met Gly Cys Gln Asp Glu Gln Leu Val Gln Thr Ile Cys Asp Leu Tyr
 1 5 10 15

Glu Lys Ile Ser Lys Leu Glu Ser Leu Lys Pro Ser Glu Asp Val Asn
 20 25 30

Ile Leu Phe Lys Gln Leu Val Ser Thr Cys Ile Pro Pro Asn Pro Asn
 35 40 45

Ile Asp Val Thr Lys Met Cys Asp Arg Val Gln Glu Ile Arg Leu Asn
 50 55 60

Leu Ile Lys Ile Cys Gly Leu Ala Glu Gly His Leu Glu Asn His Phe
 65 70 75 80

Ser Ser Ile Leu Thr Ser Tyr Gln Asp Asn Pro Leu His His Leu Asn
 85 90 95

-continued

Ile Phe Pro Tyr Tyr Asn Asn Tyr Leu Lys Leu Gly Lys Leu Glu Phe
100 105 110

Asp Leu Leu Glu Gln Asn Leu Asn Gly Phe Val Pro Lys Ser Val Ala
115 120 125

Phe Ile Gly Ser Gly Pro Leu Pro Leu Thr Ser Ile Val Leu Ala Ser
130 135 140

Phe His Leu Lys Asp Thr Ile Phe His Asn Phe Asp Ile Asp Pro Ser
145 150 155 160

Ala Asn Ser Leu Ala Ser Leu Leu Val Ser Ser Asp Pro Asp Ile Ser
165 170 175

Gln Arg Met Phe Phe His Thr Val Asp Ile Met Asp Val Thr Glu Ser
180 185 190

Leu Lys Ser Phe Asp Val Val Phe Leu Ala Ala Leu Val Gly Met Asn
195 200 205

Lys Glu Glu Lys Val Lys Val Ile Glu His Leu Gln Lys His Met Ala
210 215 220

Pro Gly Ala Val Leu Met Leu Arg Ser Ala His Gly Pro Arg Ala Phe
225 230 235 240

Leu Tyr Pro Ile Val Glu Pro Cys Asp Leu Gln Gly Phe Glu Val Leu
245 250 255

Ser Ile Tyr His Pro Thr Asp Asp Val Ile Asn Ser Val Val Ile Ser
260 265 270

Lys Lys His Pro Val Val Ser Ile Gly Asn Val Gly Gly Pro Asn Ser
275 280 285

Cys Leu Leu Lys Pro Cys Asn Cys Ser Lys Thr His Ala Lys Met Asn
290 295 300

Lys Asn Met Met Ile Glu Glu Phe Gly Ala Arg Glu Glu Gln Leu Ser
305 310 315 320

<210> SEQ ID NO 22

<211> LENGTH: 963

<212> TYPE: DNA

<213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 22

```

atgggttgcc aagacgaaca attggtgcaa acaatatgcg atctctacga aaagatctca    60
aagcttgaga gtctaaaacc atccgaagat gtcaacattc tttcaagca gctcgtttcc    120
acatgcatac caccaaaacc taacatogat gtcaccaaga tgtgtgacag agtccaagag    180
attcgactta atctcatcaa gatttgtggt ctagccgaag gtcacttaga aaaccatttc    240
tcttcgatct tgacctctta ccaagacaac ccacttcac atttaaacat tttcccttat    300
tacaacaact atttgaact cggaaagtc gagttogacc tctcogaaca aaacctaaat    360
ggctttgtcc caaagagtgt ggctttcatt ggatctggtc ctcttctct cacttccatc    420
gttcttgctt cattccatct caaagacaca atctttcaca actttgacat cgaccatca    480
gcgaactcac tgcttctct tctggtttcc tetgatccag acatctctca acgcatgttc    540
ttccacaccg ttgatataat ggaogtgaca gagagcttaa agagctttga tgtcgtgttt    600
ctagctgctc ttgttggaaat gaacaaagag gagaaagtta aagtgatcga gcatctgcag    660
aaacacatgg ctctgtgtgc tgtgtctatg cttaggagtg ctcattgtcc gagagcgttt    720
ctttatccga tcgttgagcc gtgtgatctt caggggttcg aggttttgtc tatttatcac    780

```


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```

ccaacagatg atgttatcaa ctccgtggtg atctctaaaa agcatccagt tgtttcaatt      840
gggaatggtg gtggctctaa ttcatgcttg ctcaagcctt gcaactgttc caagaccac      900
gcgaaaatga acaagaacat gatgatcgag gagttcggag ctaggaggga acagttgtct      960
taa                                                                           963

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```

<210> SEQ ID NO 23
<211> LENGTH: 32
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
consensus sequence
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (3)
<223> OTHER INFORMATION: Variable amino acid residue
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (9)
<223> OTHER INFORMATION: Variable amino acid residue
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (16)
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<220> FEATURE:
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<223> OTHER INFORMATION: Variable amino acid residue
<220> FEATURE:
<221> NAME/KEY: MOD_RES
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<223> OTHER INFORMATION: Variable amino acid residue

<400> SEQUENCE: 23

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Leu Pro Xaa Leu Ser Pro Ser Pro Xaa Val Asp Arg Leu Phe Thr Xaa
1           5           10           15
Leu Val Xaa Ala Cys Val Pro Xaa Ser Pro Val Asp Val Thr Lys Leu
20          25          30

```

```

<210> SEQ ID NO 24
<211> LENGTH: 16
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
consensus sequence
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (7)
<223> OTHER INFORMATION: Variable amino acid residue
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (11)
<223> OTHER INFORMATION: Variable amino acid residue

<400> SEQUENCE: 24

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Leu Ile Arg Leu Cys Ser Xaa Ala Glu Gly Xaa Leu Glu Ala His Tyr
1           5           10           15

```

```

<210> SEQ ID NO 25
<211> LENGTH: 10
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
consensus sequence

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<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (7)
<223> OTHER INFORMATION: Variable amino acid residue

<400> SEQUENCE: 25

Pro Leu Asp His Leu Gly Xaa Phe Pro Tyr
1 5 10

<210> SEQ ID NO 26
<211> LENGTH: 13
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
consensus sequence

<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (4)
<223> OTHER INFORMATION: Variable amino acid residue

<400> SEQUENCE: 26

Val Ala Phe Xaa Gly Ser Gly Pro Leu Pro Phe Ser Ser
1 5 10

<210> SEQ ID NO 27
<211> LENGTH: 11
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
consensus sequence

<400> SEQUENCE: 27

Asp Val Val Phe Leu Ala Ala Leu Val Gly Met
1 5 10

<210> SEQ ID NO 28
<211> LENGTH: 12
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
consensus sequence

<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (5)
<223> OTHER INFORMATION: Variable amino acid residue
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (10)
<223> OTHER INFORMATION: Variable amino acid residue

<400> SEQUENCE: 28

Arg Gly Gly Phe Xaa Val Leu Ala Val Xaa His Pro
1 5 10

<210> SEQ ID NO 29
<211> LENGTH: 328
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
amino acid sequence of nicotianamine synthase

<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (1)..(10)
<223> OTHER INFORMATION: Variable amino acid residue

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<220> FEATURE:
<221> NAME/KEY: MOD_RES
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<223> OTHER INFORMATION: Variable amino acid residue
<220> FEATURE:
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<223> OTHER INFORMATION: Variable amino acid residue
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<223> OTHER INFORMATION: Variable amino acid residue
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<223> OTHER INFORMATION: Variable amino acid residue
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<223> OTHER INFORMATION: Variable amino acid residue
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<223> OTHER INFORMATION: Variable amino acid residue
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<223> OTHER INFORMATION: Variable amino acid residue
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<223> OTHER INFORMATION: Variable amino acid residue
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<223> OTHER INFORMATION: Variable amino acid residue
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<223> OTHER INFORMATION: Variable amino acid residue
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<223> OTHER INFORMATION: Variable amino acid residue
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<223> OTHER INFORMATION: Variable amino acid residue
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<223> OTHER INFORMATION: Variable amino acid residue
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<223> OTHER INFORMATION: Variable amino acid residue
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<223> OTHER INFORMATION: Variable amino acid residue

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<220> FEATURE:
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<223> OTHER INFORMATION: Variable amino acid residue
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<222> LOCATION: (94)..(94)
<223> OTHER INFORMATION: Variable amino acid residue
<220> FEATURE:
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<222> LOCATION: (97)..(98)
<223> OTHER INFORMATION: Variable amino acid residue
<220> FEATURE:
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<223> OTHER INFORMATION: Variable amino acid residue
<220> FEATURE:
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<222> LOCATION: (106)..(107)
<223> OTHER INFORMATION: Variable amino acid residue
<220> FEATURE:
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<223> OTHER INFORMATION: Variable amino acid residue
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<223> OTHER INFORMATION: Variable amino acid residue
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<222> LOCATION: (117)..(128)
<223> OTHER INFORMATION: Variable amino acid residue
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<223> OTHER INFORMATION: Variable amino acid residue
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<223> OTHER INFORMATION: Variable amino acid residue
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<222> LOCATION: (164)..(165)
<223> OTHER INFORMATION: Variable amino acid residue
<220> FEATURE:
<221> NAME/KEY: MOD_RES
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<223> OTHER INFORMATION: Variable amino acid residue
<220> FEATURE:
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<222> LOCATION: (170)..(179)
<223> OTHER INFORMATION: Variable amino acid residue
<220> FEATURE:
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<222> LOCATION: (182)..(182)
<223> OTHER INFORMATION: Variable amino acid residue

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<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (184)..(184)
<223> OTHER INFORMATION: Variable amino acid residue
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<222> LOCATION: (186)..(194)
<223> OTHER INFORMATION: Variable amino acid residue
<220> FEATURE:
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<223> OTHER INFORMATION: Variable amino acid residue
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<223> OTHER INFORMATION: Variable amino acid residue
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<223> OTHER INFORMATION: Variable amino acid residue
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<223> OTHER INFORMATION: Variable amino acid residue
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<223> OTHER INFORMATION: Variable amino acid residue
<220> FEATURE:
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<223> OTHER INFORMATION: Variable amino acid residue
<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (272)..(276)
<223> OTHER INFORMATION: Variable amino acid residue

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<220> FEATURE:
<221> NAME/KEY: MOD_RES
<222> LOCATION: (278)..(328)
<223> OTHER INFORMATION: Variable amino acid residue

<400> SEQUENCE: 29

Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Leu Xaa Xaa Lys Ile Xaa
1           5           10           15
Xaa Xaa Xaa Xaa Xaa Ile Xaa Xaa Leu Xaa Xaa Leu Xaa Pro Xaa Xaa
20          25          30
Xaa Xaa Xaa Xaa Leu Phe Xaa Xaa Leu Val Xaa Xaa Cys Xaa Pro Xaa
35          40          45
Xaa Xaa Xaa Asp Val Xaa Xaa Xaa Xaa Xaa Xaa Xaa Gln Xaa Met Arg
50          55          60
Xaa Xaa Leu Ile Xaa Xaa Cys Xaa Xaa Ala Glu Xaa Xaa Leu Glu Xaa
65          70          75          80
His Xaa Xaa Xaa Xaa Leu Xaa Xaa Xaa Asp Xaa Pro Leu Xaa His Leu
85          90          95
Xaa Xaa Phe Pro Tyr Xaa Xaa Asn Tyr Xaa Xaa Leu Xaa Xaa Leu Glu
100         105         110
Xaa Xaa Leu Leu Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
115         120         125
Ala Phe Xaa Gly Ser Gly Pro Leu Pro Xaa Xaa Ser Xaa Xaa Leu Ala
130         135         140
Xaa Xaa His Leu Xaa Xaa Xaa Xaa Phe Xaa Asn Xaa Xaa Xaa Xaa Xaa
145         150         155         160
Xaa Ala Asn Xaa Xaa Ala Xaa Xaa Leu Xaa Xaa Xaa Xaa Xaa Xaa Xaa
165         170         175
Xaa Xaa Xaa Arg Met Xaa Phe Xaa Thr Xaa Xaa Xaa Xaa Xaa Xaa
180         185         190
Xaa Xaa Leu Xaa Xaa Xaa Asp Val Val Phe Leu Ala Ala Xaa Val Gly
195         200         205
Met Xaa Xaa Xaa Xaa Lys Xaa Xaa Xaa Xaa Xaa His Leu Xaa Xaa His
210         215         220
Met Xaa Xaa Gly Ala Xaa Leu Xaa Xaa Xaa Xaa Xaa Xaa Xaa Arg Xaa
225         230         235         240
Phe Leu Tyr Pro Xaa Val Xaa Xaa Xaa Xaa Xaa Xaa Xaa Gly Phe
245         250         255
Xaa Val Leu Xaa Val Xaa His Pro Xaa Xaa Xaa Val Xaa Asn Ser Xaa
260         265         270
Xaa Xaa Xaa Xaa Lys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
275         280         285
Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
290         295         300
Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
305         310         315         320

Xaa Xaa Xaa Xaa Xaa Xaa Xaa
325

<210> SEQ ID NO 30
<211> LENGTH: 19
<212> TYPE: PRT
<213> ORGANISM: Hordeum vulgare

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<400> SEQUENCE: 30

Asp Ala Gln Asn Lys Glu Val Ala Ala Leu Ile Glu Lys Ile Ala Gly
 1 5 10 15

Ile Gln Ala

<210> SEQ ID NO 31

<211> LENGTH: 7

<212> TYPE: PRT

<213> ORGANISM: Hordeum vulgare

<400> SEQUENCE: 31

Arg Glu Ala Leu Ile Arg Leu
 1 5

<210> SEQ ID NO 32

<211> LENGTH: 71

<212> TYPE: PRT

<213> ORGANISM: Oryza sativa

<400> SEQUENCE: 32

Met Glu Ala Gln Asn Gln Glu Val Ala Ala Leu Val Glu Lys Ile Ala
 1 5 10 15

Gly Leu His Ala Ala Ile Ser Lys Leu Pro Ser Leu Ser Pro Ser Ala
 20 25 30

Glu Val Asp Ala Leu Phe Thr Asp Leu Val Thr Ala Cys Val Pro Ala
 35 40 45

Ser Pro Val Asp Val Ala Lys Leu Gly Pro Glu Ala Gln Ala Met Arg
 50 55 60

Glu Glu Leu Ile Arg Leu Cys
 65 70

<210> SEQ ID NO 33

<211> LENGTH: 111

<212> TYPE: PRT

<213> ORGANISM: Oryza sativa

<400> SEQUENCE: 33

Tyr Val Asn Leu Ser Lys Leu Glu Tyr Asp Leu Leu Val Arg Tyr Val
 1 5 10 15

Pro Gly Ile Ala Pro Thr Arg Val Ala Phe Val Gly Ser Gly Pro Leu
 20 25 30

Pro Phe Ser Ser Leu Val Leu Ala Ala His His Leu Pro Asp Ala Val
 35 40 45

Phe Asp Asn Tyr Asp Arg Cys Gly Ala Ala Asn Glu Arg Ala Arg Arg
 50 55 60

Leu Phe Arg Gly Ala Asp Glu Gly Leu Gly Ala Arg Met Ala Phe His
 65 70 75 80

Thr Gly Asp Val Ala Thr Leu Thr Gly Glu Leu Gly Ala Tyr Asp Val
 85 90 95

Val Phe Leu Ala Thr Leu Val Gly Met Ala Ala Glu Glu Lys Pro
 100 105 110

<210> SEQ ID NO 34

<211> LENGTH: 64

<212> TYPE: PRT

<213> ORGANISM: Hordeum vulgare

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<400> SEQUENCE: 34

Ser Phe His Thr Ala Asp Val Ala Asp Leu Thr Gln Glu Leu Gly Ala
 1 5 10 15

Tyr Asp Val Val Phe Leu Ala Ala Leu Val Asp Met Ala Ala Glu Glu
 20 25 30

Lys Ala Lys Val Ile Ala His Leu Gly Ala His Met Val Glu Gly Ala
 35 40 45

Ser Leu Val Val Tyr Ser Ala His Gly Ala Arg Gly Phe Leu Tyr Pro
 50 55 60

<210> SEQ ID NO 35

<211> LENGTH: 19

<212> TYPE: PRT

<213> ORGANISM: Hordeum vulgare

<400> SEQUENCE: 35

Ala Phe His Thr Ala Glu Val Thr Asp Leu Thr Ala Glu Leu Gly Ala
 1 5 10 15

Tyr Asp Val

<210> SEQ ID NO 36

<211> LENGTH: 29

<212> TYPE: PRT

<213> ORGANISM: Hordeum vulgare

<400> SEQUENCE: 36

Ala Asp Gly Ala Val Leu Val Ala Arg Ser Ala His Gly Ala Arg Ala
 1 5 10 15

Phe Leu Tyr Pro Val Val Glu Leu Asp Asp Val Gly Arg
 20 25

<210> SEQ ID NO 37

<211> LENGTH: 20

<212> TYPE: PRT

<213> ORGANISM: Hordeum vulgare

<400> SEQUENCE: 37

Pro Glu Asp Ile Arg Arg Gly Gly Phe Glu Val Leu Ala Val His His
 1 5 10 15

Pro Glu Gly Glu
 20

1.-26. (canceled)

27.-35. (canceled)

36. An isolated or purified polypeptide comprising an amino acid sequence that is at least 90% identical to SEQ ID NO:15.

37. The isolated or purified polypeptide of claim 36 comprising an amino acid sequence that is at least 95% identical to SEQ ID NO: 15.

38. The isolated or purified polypeptide of claim 36, wherein the polypeptide consists of the amino acid sequence set forth as SEQ ID NO: 15.

39. The isolated or purified polypeptide of any one of claims 36-38, wherein the polypeptide has nicotianamine synthase activity.

40. An isolated or purified *Oryza* enzyme exhibiting nicotianamine synthase activity, wherein:

- a. the enzyme is:
 - i. isolated or purified from *Oryza*; or
 - ii. expressed directly or indirectly from a nucleic acid isolated or purified from *Arabidopsis*; or
 - iii. expressed directly or indirectly from a chimeric nucleic acid at least partially isolated or purified from *Arabidopsis*;
- b. the enzyme comprises a polypeptide having at least 90% identity with an amino acid sequence of SEQ ID NO: 15,
- c. the enzyme has more than 25% of the relative nicotianamine synthase activity of the enzyme of SEQ ID NO:15.

* * * * *