



(11) **EP 1 930 346 B1**

(12) **EUROPEAN PATENT SPECIFICATION**

(45) Date of publication and mention of the grant of the patent:  
**19.10.2011 Bulletin 2011/42**

(21) Application number: **06796940.2**

(22) Date of filing: **29.08.2006**

(51) Int Cl.:  
**C07K 16/00** <sup>(2006.01)</sup> **A61K 39/395** <sup>(2006.01)</sup>  
**A61P 35/00** <sup>(2006.01)</sup> **C12P 21/08** <sup>(2006.01)</sup>  
**C07K 16/28** <sup>(2006.01)</sup> **C07K 16/10** <sup>(2006.01)</sup>  
**C07K 16/18** <sup>(2006.01)</sup>

(86) International application number:  
**PCT/JP2006/316960**

(87) International publication number:  
**WO 2007/026689 (08.03.2007 Gazette 2007/10)**

(54) **ANTIBODY PRODUCED USING OSTRICH AND METHOD FOR PRODUCTION THEREOF**  
MIT EINEM STRAUSS PRODUZIERTER ANTIKÖRPER UND PRODUKTIONSVERFAHREN DAFÜR  
ANTICORPS OBTENU EN SE SERVANT D'UNE AUTRUCHE ET SON PROCÉDÉ DE PRODUCTION

(84) Designated Contracting States:  
**DE FR GB**

(30) Priority: **29.08.2005 JP 2005246993**

(43) Date of publication of application:  
**11.06.2008 Bulletin 2008/24**

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- **BLIGNAUT A ET AL: "Antibody responses to La Sota strain vaccines of Newcastle disease virus in ostriches (*Struthio camelus*) as detected by enzyme-linked immunosorbent assay" AVIAN DISEASES, vol. 44, no. 2, April 2000 (2000-04), pages 390-398, XP008102082 ISSN: 0005-2086**
- **SHIVAPRASAD H L ET AL: "Turlock-like bunyavirus associated with encephalomyelitis and myocarditis in an ostrich chick." JOURNAL OF VETERINARY DIAGNOSTIC INVESTIGATION : OFFICIAL PUBLICATION OF THE AMERICAN ASSOCIATION OF VETERINARY LABORATORY DIAGNOSTICIANS, INC SEP 2002, vol. 14, no. 5, September 2002 (2002-09), pages 363-370, XP002515165 ISSN: 1040-6387**
- **DATABASE BIOSIS [Online] BIOSCIENCES INFORMATION SERVICE, PHILADELPHIA, PA, US; May 2006 (2006-05), SAKAI KOUJI ET AL: "Serological and virological studies of Newcastle disease and avian influenza in slaughter-age ostriches (*Struthio camelus*) in Japan" XP002515166 Database accession no. PREV200600490842 & JOURNAL OF VETERINARY MEDICAL SCIENCE, vol. 68, no. 5, May 2006 (2006-05), pages 491-494, ISSN: 0916-7250**
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Remarks:

The file contains technical information submitted after the application was filed and not included in this specification

**Description**Technical Field

**[0001]** The present invention relates to an antibody produced using an ostrich and a method for production thereof. The present invention is useful not only for mass production of antibodies for medical use (that is, antibodies for treatment or diagnosis), but also for other various fields where antibodies may be used.

Background Art

**[0002]** Conventional animals used for production of antibodies are, for example, the mouse, the rat, the sheep and the goat. These mammals, however, have their proteins which are highly similar to the human proteins and therefore, it is often difficult to produce an antibody that suppresses function of a specific human protein by using these animals. In other words, even if these animals are immunized against a human molecule (for example, a human protein) that relates to a certain disease, it was often difficult to produce a desired antibody (particularly, an antibody for treatment) within these animals.

**[0003]** On the other hand, the production of anti-human antibodies for treatment or diagnosis by using the chicken, which is far apart from the human, has been recently attempted (for example, see the Patent document 1 listed below). However, this method has the problem that it is difficult to produce a large amount of homogeneous antibodies, because the chicken has very little blood. Moreover, a conventional method for producing a monoclonal antibody has the problem that many cell cultures, media for the cell cultures and complicated work are needed in order to produce a sufficient amount of antibodies.

[Patent document 1] Japanese Laid-Open Patent Publication No. 2001-238676

**[0004]** BLIGNAUT A ET AL, AVIAN DISEASES, vol. 44, no. 2, April 2000, p. 390-398 discloses the production of polyclonal antibodies binding to the Newcastle disease virus after immunisation of ostriches.

**[0005]** SHIVAPRASAD H L ET AL, JOURNAL OF VETERINARY DIAGNOSTIC INVESTIGATION, September 2002, vol. 14, no. 5, p. 363-370 describes Turlock-like bunyavirus associated with encephalomyelitis and myocarditis in an ostrich chick. The neutralisation properties of an ostrich serum for said bunyavirus are described in table 2.

**[0006]** DE GIROLAMO, VETERINARY RESEARCH COMMUNICATIONS, vol. 27, September 2003, p. 591-593 discloses proteins called S-100 which belong to the group of calcium-binding proteins being highly conserved throughout phylogenesis. Some of the proteins of the S-100 family are from human sources. The authors describe antibodies reacting with S-100 isolated from ostriches.

Disclosure of Invention

**[0007]** The present invention is made to solve the above-mentioned problems and intended to provide a novel method for producing a large amount of homogeneous antibodies in a simple manner (particularly, a method for producing antibodies which were difficult to produce by conventional methods using the mammals).

**[0008]** In accordance with the present invention an antibody is produced by administering to an ostrich a human CD 166 or CD 146 protein or a peptide fragment thereof as an antigen, said antibody specifically binding to said human protein or a peptide fragment thereof.

**[0009]** In a further embodiment of the invention, an antibody is produced by administering to an ostrich a viral protein of IBV (Infectious Bronchitis Virus), SARS Virus or Avian Influenza Virus, or a peptide fragment thereof as an antigen, said antibody specifically binding to said protein or a peptide fragment thereof, and said antibody is for examination of IBV (Infectious Bronchitis Virus), SARS Virus or Avian Influenza Virus.

**[0010]** In order to solve the above-mentioned problems, the present inventor focused on use of the ostrich of a large-size bird. As a result of research and analysis, the present inventor has found the following points, which led to the present invention;

(1) By immunizing the ostrich, it becomes possible to easily produce an antibody detecting an antigen protein (or part thereof) highly reserved among the mammals, which was difficult to produce by conventional methods using the mouse and the rabbit.

(2) Using the antibody produced by using the ostrich, western blot analysis and immunocytochemistry were carried out. As a result, the antibody was able to specifically detect the target protein (e.g. CD166 protein) which is a cancer marker and thus, the antibody can be used as an antibody for diagnosis.

(3) The above anti-CD166 antibody, produced by using the ostrich, also had antitumor effect of suppressing tumor growth. Therefore, the antibody can be also used as an antibody for treatment.

**[0011]** That is, the present invention provides an antibody produced by using the ostrich and method for production thereof. The ostrich is now becoming a farm animal (used for food) and the number of ostrich farms is also increasing in Japan. Food for keeping the ostrich can be purchased on the market, and its nutrition efficiency is very high. The ostrich can reach about as much as 120kg in weight and it is possible to collect a large amount of blood from a single ostrich, which corresponds to an amount of almost 100 chickens' blood. That is, it becomes possible to produce a large amount of homogeneous antibodies for treatment (or diagnosis) by using a single ostrich, which were difficult to produce by using the mammals such as the mouse and the rat. Thus, the present invention can overcome the problem of lot-to-lot

variation when producing polyclonal antibodies by using other animals. In addition, by using an infant ostrich, the immunization can be realized by a small amount of antigens, and then it becomes possible to produce a large amount of antibodies from a single ostrich by growing up to be adult for about ten months and additional immunization.

**[0012]** Thus, by using the ostrich, it becomes possible to easily produce such antibodies, specifically detecting proteins (or part thereof), that were difficult to produce by conventional methods. In addition, the antibody of the present invention has an advantage in commercial use (as an antibody for diagnosis and treatment), since it can be produced in large quantities and homogeneously without lot-to-lot variation.

**[0013]** Here, "antibody" means the protein that specifically binds to and detects an antigen. The kinds of antibodies are not particularly limited (for example, an antibody may be IgG, IgM, IgA, IgD, IgE or IgY). The antibody of the present invention may be monovalent or multivalent. Also, the antibody may be labeled with a fluorescent substance, a radioactive substance, an enzyme or other proteins (such as avidin). Or, the antibody of the present invention may be not labeled and a secondary antibody, binding to and detecting a primary antibody of the present invention, may be used in order to detect a target molecule.

**[0014]** In this DESCRIPTION, "ostrich" means one of the birds that belong to the *Struthioniformes*. Among the birds, it is preferable to use the *Struthio camelus* that belongs to the *Struthionidae*. A female or male ostrich may be used. Here, "using an ostrich" means that the ostrich is used as an animal to which an antigen is administered for the production of an antibody. A substance, which is administered as an antigen, is not particularly limited. Proteins, peptides (including nature peptides, synthetic peptides and recombinant peptides), polysaccharides and other biological substances are available as an antigen. Also, a method for preparing and administering an antigen is not particularly limited, although preferable embodiments are described later.

**[0015]** In the Examples described later, the antibody to human CD166 protein was produced by immunizing (vaccinating) an infant ostrich against a synthetic peptide, which was used as antigen. The synthetic peptide consisted of 20 amino acids, which are located in the fourth Ig loop of the extracellular region and highly reserved among the mammals. Thus produced anti-CD166 ostrich antibody turned out to be able to specifically detect human CD166 protein which is a cancer marker (FIGS. 1 and 3) and thus, it can be used as an antibody for diagnosis of cancer. Moreover, it was found out that the above anti-CD 166 ostrich antibody had antitumor effect of suppressing tumor growth (FIG. 4), which means that the above antibody can be also used as an antibody for treatment of cancer. Thus, by using the ostrich, it becomes possible to easily produce an antibody for medical use (that is, an antibody for diagnosis or treatment), which

was difficult to produce by conventional methods using the mammals.

**[0016]** Of course, in the production of antibody to human CD166 protein, a used antigen is not limited to the above-mentioned synthetic peptide. The whole human CD166 protein, or part thereof consisting of a specific sequence (preferably about 10-20 residues), can be also used as an antigen. By a similar method, anti-CD 146 antibodies (antibodies to CD 146 antigens) may be produced. Here, CD (Cluster of differentiation) means an antigen of a cell surface (for example, an adhesive molecule), to which a specific number is denominated according to the CD classification. The present invention, i.e., a method for producing an antibody by using the ostrich, can realize easy production of a large amount of homogeneous anti-human CD 166 and 146 antibodies with high detecting ability.

**[0017]** As described above, use of an infant ostrich has the advantage that the immunization can be realized by a small amount of antigens. Therefore, it is preferable to produce the antibody of the present invention by firstly immunizing an infant ostrich. Specifically, an infant ostrich, which is firstly immunized, may be preferably at not less than the age of two months and half, and at less than the age of ten months. More preferably, an infant ostrich, which is firstly immunized, may be at not less than the age of three months, and at less than the age of seven months. It is much more preferable to firstly immunize an infant ostrich at not less than the age of three months, and at less than the age of five months.

**[0018]** The following are descriptions of advantages and preferable embodiments of the production of an antibody by using the ostrich;

(1) The ostrich grows up rapidly from its hatch (about 0.8kg in weight) and will reach 60kg at the age of six months and 120kg at the age of ten months (adult). This means that, by starting the immunization from an infant bird, small amount of antigens and several times of additional immunizations are sufficient to obtain a large amount of antibodies.

(2) The following is one example of methods for production of antibodies; a synthetic peptide can be used as an antigen and the antigen dose for immunization is 100 $\mu$ g per once. The ostrich is first immunized at the age of three months and then, immunized at the same antigen dose every month to grow up to be adult. (In the Example described later, the ostrich was immunized every other week, and three times in total.) In the first immunization, 0.5mL of solution with 100 $\mu$ g antigen is mixed with 0.5mL of complete adjuvant, and the mixture is vaccinated into the breast muscle of the ostrich. From the second immunization, 0.5mL of solution with 100 $\mu$ g antigen is mixed with 0.5mL of incomplete adjuvant, and the mixture is vaccinated into the neck muscle of the ostrich. Blood of the immunized ostrich is collected 2-5 days after the final immunization and antibodies

are purified from the antiserum in the blood by using a protein G column.

(3) For measurement of the antibody titer, it is preferable to collect blood from the wing vein of the immunized ostrich until the age of four months and from the jugular vein after that age. (After the age of five months, the ostrich becomes brutal.)

(4) It is preferable to start the immunization after the age of three months, since an infant ostrich is weak to stress before the age of two months.

(5) As a result of immunizing an ostrich by the method described in the above (2), the rise of the antibody titer was already confirmed at the time of the third immunization (at the age of five months). The antibody titer reached the highest value at the time of the fifth immunization (at the age of seven months). It is possible to produce antibodies at this stage by collecting a large amount of blood of the immunized ostrich. However, the highest value of the antibody titer can be maintained by extension of breeding period and additional immunization until the adult (120kg in weight). Therefore, it is possible to produce much more antibodies at the adult.

(6) The antibody can be continuously collected as long as the immunized ostrich is not dead and thus, it is possible to continuously provide antibodies.

(7) Antibodies of the bird can move to its egg. The egg of the immunized ostrich may include antibodies with high concentration. That is, the egg may be used in order to collect antibodies continuously, easily and in large quantities, which is also described later.

(8) It is preferable to use a protein G in the purification of antibodies IgG obtained by immunizing an ostrich, since a protein A is weak at binding to these antibodies.

**[0019]** The following are descriptions of further advantages and preferable embodiments of the production of an antibody by using the ostrich, and examples of industrial application of the produced antibody.

By using the ostrich, the present invention can produce high sensitive antibodies in large quantities homogeneously and in a simple manner, as compared with the production by using the chicken or other animals. For example, two kinds of antibodies, i.e., the antibody to human CD146 and the antibody to human CD166, were produced by using the ostrich and the chicken. Then, antigen-detecting abilities of two kinds of ostrich antibodies were respectively compared with those of two kinds of antibodies produced by using the chicken. The result was that both ostrich antibodies had higher antigen-detecting abilities than the chicken antibodies (FIGS. 8 and 9) and thus, the ostrich antibodies have qualitative advantage.

**[0020]** As described in the above (7), it is possible to produce a large amount of antibodies continuously and easily, by immunizing a female ostrich against an antigen and then purifying antibodies (IgY) from the yolk of the egg produced by that female ostrich. For example, a fe-

male ostrich, producing eggs, was immunized every week against Collagen Type I as an antigen. As a result, four weeks after the first immunization, the production of antibodies was confirmed in the yolk of the egg produced by that female ostrich (FIGS. 5 and 6). In a similar way, a female ostrich, producing eggs, was immunized every week against an inactivated vaccine of IBV (Infectious Bronchitis Virus). As a result, six weeks after the first immunization, the production of antibodies was confirmed in the yolk of the egg produced by that female ostrich (FIG. 7).

**[0021]** Moreover, by adopting the method for purification of IgY, 2-4g of antibodies (IgY) were purified per the yolk of a single egg produced by the female ostrich of six weeks after the first immunization. During the ostrich egg producing period ranging from April to September, a single female ostrich produces about 100 eggs, from which about 400g of antibodies (IgY) can be produced at maximum. This means that, for example if producing a diagnosis kit for testing virus infection, a single female ostrich can produce a large amount of antibodies that can be used as diagnosis kits for testing about 4-40 million samples, at the time of six months after the first immunization by adjusting the timing of immunization.

**[0022]** The method for purification of IgY from an ostrich egg yolk may adopt essentially the same method as that for purification of IgY from a chicken egg yolk (for example, see J. Immunol. Methods 46: 63-68, 1981.; Agric. Biol. Chem. 54: 2531-2535, 1990.; Immunological Investigations 19: 253-258, 1990.). Commercial kit for purification of antibodies (IgY) from a chicken egg yolk may be used for purification of antibodies (IgY) from an ostrich egg yolk, although it is preferable to increase up to about 1.5 times the amount of all the reagents (for example, dextran sulfate/TBS), as compared with purification from a chicken egg yolk, since the ostrich egg yolk includes more lipids.

**[0023]** The antibody of the present invention can be used in various fields and its use is not particularly limited. The antibody of the present invention can be used as antibodies presently used in various fields, in addition to antibodies for medical use as described above. Especially, the antibody of the present invention can be mass-produced by using an ostrich. Therefore, by applying the present invention to the production of antibodies for examination (used as a kit for examining cancer and a kit for examining viral infection, etc.), it becomes possible to easily and promptly prepare a large amount of examination kits from a single ostrich, which are required to examine enormous samples in the examinations of physical condition, infectious disease and food poisoning.

**[0024]** According to the invention it becomes possible to produce a large amount of antibodies against SARS (Severe Acute Respiratory Syndrome) virus by using an ostrich, since the above-mentioned IBV (Infectious Bronchitis Virus) is a coronavirus close to the SARS virus. It is also possible to produce a large amount of antibodies for detection of Avian Influenza Virus. In addition, the

antibody of the present invention can be used as an industrial antibody. That is, it can be used as antibodies for removal of pathogenic substances, allergens or other antigens, which can be applied in various industrial products such as an air conditioner filter, a face mask and a water filter.

**[0025]** Thus, the present invention can be used as an industrial antibody for removal of virus by applying to an air conditioner filter, a face mask and other products, in addition to using as antibodies for detection of virus or antibodies for examination of diseases. Of course, it is possible to apply to an antibody for detection and/or removal of pathogenic substances (such as bacteria causing infectious disease or food poisoning), allergens in food (such as buckwheat and ovalbumin) and pathogenic substances within drinking water.

**[0026]** The antibody of the present invention may be produced as a polyclonal antibody or a monoclonal antibody. General production method of such antibody is described in, for example, "Antibodies: A laboratory manual" written by Harlow et al., Cold Spring Harbor Laboratory, New York (1988) and "Monoclonal antibody: Hybridoma and ELISA" written by Iwasaki et al., Kodansha (1991). The following is a brief explanation of the production method of a polyclonal antibody and a monoclonal antibody.

#### (1) PRODUCTION OF POLYCLONAL ANTIBODIES

**[0027]** The polyclonal antibody may be produced by the following method; firstly, a substance is administered to an ostrich as an antigen. The substance may be administered alone or with other substance such as a carrier and a diluent. The substance may be administered to, for example, the breast muscle and the neck muscle of the ostrich, as described above, which enables the production of antibodies in the ostrich. To improve the productivity of antibodies, the substance may be administered with Freund's complete or incomplete adjuvant. The antigen may be administered once only. However, the antigen is usually administered once every 2-6 weeks and administered about 2-10 times in total.

**[0028]** Antibody titer to the antigen in the ostrich serum may be measured by a liquid phase method or a solid phase method. In the liquid phase method, for example, labeled antigen is reacted to the antiserum and then, the activity of labeling agents binding to antibodies is measured. In the solid phase method, for example, the antigen is immobilized to an inner wall of each well in 96 hole plate and then, the diluted serum is added to each well, in order to bind antibodies to the antigen, followed by removal of extra antibodies by washing the solution in each well. Then, antibodies binding to the inner wall of each well are measured.

**[0029]** Polyclonal antibodies of the present invention may be purified by a method for separation and purification of immunoglobulin. Such method includes salting-out method, alcohol precipitation method, isoelectric pre-

cipitation method, electrophoresis, adsorption and desorption by an ion exchanger such as DEAE, ultracentrifugal method, gel filtration technique and antigen binding solid phase method, in addition to specific purification method for obtaining antibodies by collecting antibodies with an activated adsorbent such as a protein G and then dissociating the antibodies from the adsorbent.

#### (2) PRODUCTION OF MONOCLONAL ANTIBODIES

**[0030]** The monoclonal antibody may be produced by the following method; firstly, among ostriches which have been immunized against an antigen, the ostriches having antibodies are selected. After collecting the spleen or the lymph node of these ostriches 2-5 days after the final immunization, an antibody-producing cell contained in such organ is fused with a myeloma cell, so as to prepare a hybridoma producing an antibody of the present invention.

**[0031]** The fusion technique may be carried out according to any of known methods, for example, Kayler and Milstein method (Nature Vol. 256, Page 495 (1975)). As a fusion promoting agent, polyethylene glycol (PEG) and Sendai virus are exemplified, and preferably PEG and Sendai virus are exemplified, and preferably PEG may be used. As the myeloma cell, NS-1, P3U1, SP2/0 and AP-1 are exemplified, and preferably P3U1 may be used. In the fusion, preferable ratio of the number of the antibody-producing cells (such as spleen cells) and the number of the myeloma cells is usually about at 1:1-20:1. The cell fusion may be efficiently performed by adding PEG (preferably PEG1000-PEG6000) in the concentration of about 10-80%, and incubating for 1-10 minutes usually at 20-40°C (preferably at 30-37°C).

**[0032]** Antibody-producing hybridoma may be screened by various methods. Such methods include the following methods. For example, a supernatant of cultured hybridoma is added to a solid phase (such as a microplate) in which an antigen or its partial peptide is immobilized directly or with a carrier. Next, added is an anti-ostrich immunoglobulin antibody or a protein G, which is labeled with a fluorescent substance or an enzyme etc., in order to detect a monoclonal antibody of the present invention, binding to the solid phase. In another method, the antibody-producing hybridoma is added to a solid phase in which an anti-ostrich immunoglobulin antibody or a protein G is immobilized. Next, added is an antigen labeled with a fluorescent substance or an enzyme etc., in order to detect a monoclonal antibody binding to the solid phase.

**[0033]** Screening of a monoclonal antibody of the present invention and culture for such screening may be performed by adding HAT (hypoxanthine, aminopterin and thymidine) to a medium for culture of animal cells (such as RPMI1640) including 10-20% fetal bovine serum. Antibody titer of a supernatant of cultured hybridoma may be measured by the same method as the above-mentioned method used for measurement of the antibody titer in the antiserum. Monoclonal antibodies of the

present invention may be separated and purified by the same method as the above-mentioned method used for separation and purification of polyclonal antibodies.

**[0034]** Thus, the monoclonal antibody of the present invention may be produced by culturing a hybridoma cell in an animal body or *in vitro*, and then collecting monoclonal antibodies from a body fluid or a culture.

**[0035]** Of course, the method for producing polyclonal and monoclonal antibodies of the present invention are not limited to the above-mentioned methods, and various known methods and techniques may be applied. Newly developed methods and techniques may be also applied.

#### Brief Description of Drawings

##### **[0036]**

FIG. 1 is a figure showing the result of detecting human CD166 protein by western blot analysis using each antiserum of the mouse, the rabbit and the ostrich, after immunizing these animals against human CD166 peptide.

FIG. 2 is a figure showing the result of confirming the existence of purified ostrich IgG by SDS-PAGE.

FIG. 3 is a figure showing the result of detecting human CD166 protein in human lung cancer cells (A549 cells), by immunocytochemical technique using the anti-CD166 ostrich antibodies.

FIG. 4 is a graph showing the result of investigation of *in vivo* antitumor effect of the anti-CD166 ostrich antibodies administered to a nude mouse into which human lung cancer cells have been implanted. The sign "\*" shows a significant difference compared with the case where PBS was administered. \* P<0.01 (Student-t)

FIG. 5 is a figure showing the result of examining the production of antibodies in the sera of immunized female ostriches (producing eggs) and in the yolks of their eggs by precipitation reaction in agarose gel.

In this examination, female ostriches have been immunized against Collagen Type I. In this figure, A: the antigen (Collagen Type I), 1: the ostrich serum before immunization, 2: the ostrich serum one week after immunization, 3: the ostrich serum two weeks after immunization, 4: the ostrich serum three weeks after immunization, 5: the extract from the ostrich egg yolk four weeks after immunization, 6: PBS. The white precipitation line occurred by the antigen-antibody reaction shows the production of antibodies.

FIG. 6 is a figure showing the result of examining the production of antibodies in the egg yolks of immunized female ostriches by precipitation reaction in agarose gel. In this examination, female ostriches have been immunized against Collagen Type I. In this figure, A: the antigen (Collagen Type I), 1: the extract from the ostrich egg yolk before immunization, 2: the extract from the ostrich egg yolk one week after immunization, 3: the extract from the ostrich

egg yolk two weeks after immunization, 4: the extract from the ostrich egg yolk three weeks after immunization, 5: the extract from the ostrich egg yolk four weeks after immunization, 6: PBS. The white precipitation line occurred by the antigen-antibody reaction shows the production of antibodies.

FIG. 7 is a figure showing the result of examining the production of antibodies in the sera of immunized female ostriches (producing eggs) and in the yolks of their eggs by precipitation reaction in agarose gel. In this examination, female ostriches have been immunized against an inactivated vaccine of IBV (Infectious Bronchitis Virus). In this figure, A: the antigen (IBV), 1: PBS, 2: the ostrich serum before immunization, 3: the ostrich serum four weeks after immunization, 4: the ostrich serum six weeks after immunization, 5: the extract from the ostrich egg yolk six weeks after immunization, 6: the extract from the ostrich egg yolk before immunization. The white precipitation line occurred by the antigen-antibody reaction shows the production of antibodies.

FIG. 8 is a figure showing the result of investigation in which anti-CD146 antibodies were produced respectively by using the ostrich and the chicken, and then antigen-detecting abilities of both antibodies were compared. (a) result of immunoblot analysis using each antibody, (b) graph showing relative intensity of both antibodies. The sign "\*" shows a significant difference compared with the antibody produced by using the chicken. \* P<0.01 (Student-t)

FIG. 9 is a figure showing the result of investigation in which anti-CD166 antibodies were produced respectively by using the ostrich and the chicken, and then antigen-detecting abilities of both antibodies were compared. (a) result of immunoblot analysis using each antibody, (b) graph showing relative intensity of both antibodies. The sign "\*" shows a significant difference compared with the antibody produced by using the chicken. \* P<0.01 (Student-t)

#### Examples

**[0037]** Examples of the present invention are described below in reference to the drawings.

[Example 1]

**[0038]** Produced was the antibody against human CD166 protein which may be used as a marker of various cancers such as lung cancer, melanoma and prostate cancer. In this production, a synthetic peptide was used as the antigen; i.e., twenty amino acids in the fourth Ig loop of the extracellular region, which are highly reserved among the mammals. The sequence of the twenty amino acids is all the same among the human, the mouse and the rabbit. Using the synthetic peptide, the mouse (BAL-Bc), the rabbit (white kind of Japan) and the ostrich (an infant at the age of three months) were immunized every

other week (total three times) and then, each blood was collected.

**[0039]** The first immunization was a mixture of the liquid (0.5mL) with 100 $\mu$ g of antigen and 0.5mL of complete adjuvant, and the mixture was injected for vaccination into the breast muscle of the ostrich and into the dorsal skin of the rabbit respectively. 0.4mL of this mixture was injected for vaccination into the dorsal skin of the mouse. From the second immunization, a mixture of the liquid (0.5mL) with 100 $\mu$ g of antigen and 0.5mL of incomplete adjuvant was used for vaccination by injecting the mixture into the neck muscle of the ostrich and into the dorsal skin of the rabbit respectively. 0.4mL of this mixture was injected for vaccination into the dorsal skin of the mouse. Then, western blotting was carried out using antisera obtained from these animals, to the fraction of cell membrane of human lung cancer. The antisera were obtained two weeks after the final immunization. As a result, only the ostrich was able to produce antibodies which can detect human CD166 protein (FIG. 1).

**[0040]** In the western blotting, labeled secondary antibody was used. The method for production of this secondary antibody is as follows; firstly, immunoglobulin G (IgG) was purified from the blood of non-immunized ostrich by using a protein G, and confirmed by SDS-PAGE (FIG. 2). Then, the rabbit was immunized against this ostrich IgG. Specifically, the rabbit (white kind of Japan) was immunized every other week (total four times) against 200 $\mu$ g of the ostrich IgG per once. The whole blood was collected from the rabbit two weeks after the final time, to produce anti-ostrich IgG rabbit polyclonal antibody (serum). Purification procedure was carried out according to a normal method using a protein A. Thus purified antibody was labeled with FITC or HRP and used as the secondary antibody.

**[0041]** The antibody was purified from the above-mentioned ostrich antiserum by the following method; firstly, the ostrich antiserum was loaded onto a protein G column. After it was washed with PBS, the antibody was eluted with a mixture (pH 9.0) of a citrate buffer and a glycine buffer, and then neutralized with 2M Tris/HCl. Thus, the antibody was obtained and used in the following experiments as an anti-CD166 ostrich antibody.

[Example 2]

**[0042]** Using the anti-CD166 ostrich antibody produced by Example 1 as a primary antibody, immunofluorescence assay was carried out to cultured human lung cancer cells (A549 cells). As a result, the antibody strongly reacted to the cell membranes of the lung cancer cells (FIG. 3). In this experiment, at first, the A549 cells were cultured during a whole day on a coverslip, and then fixed with the Zamboni solution. Next, the anti-CD166 ostrich antibody (1mg/mL) produced by Example 1 was diluted to 1000 times with PBS, and the antibody was used as a primary antibody in the reaction with the A549 cells for one hour at 37°C. Then, the reaction using a secondary

antibody was carried out. FITC labeled anti-ostrich IgG rabbit polyclonal antibody (1mg/mL) produced by Example 1 was diluted to 1000 times with PBS, and the antibody was used as a secondary antibody in the reaction with the A549 cells for one hour at 37°C. After the reaction, the A549 cells were observed with fluorescence microscope.

[Example 3]

**[0043]** 10<sup>6</sup> of cultured human lung cancer cells (A549 cells) were subcutaneously transplanted to each of five nude mice. From five days after the transplant, the anti-CD166 ostrich antibody (120 $\mu$ g) produced by Example 1 was intraperitoneously administered every week, and tumor volume of each mouse was measured three weeks after the transplant. Only PBS was administered to a control mouse. As a result, it turned out that tumor growth was significantly suppressed in the group to which the anti-CD166 ostrich antibody was administered (FIG. 4).

[Example 4]

**[0044]** In order to investigate the production of antibodies into an egg yolk, a female ostrich at the age of two years and six months, producing eggs, was immunized every week against rat Collagen Type I. Then, the production of antibodies was examined in the serum at one to three weeks after the first immunization, and in the yolk extract at four weeks after the first immunization, by precipitation reaction in agarose gel. The following is a detailed explanation of this experiment.

**[0045]** Antigen dose and method for immunization: Antigen solution (4mg/0.5mL) was mixed with 0.5mL of complete adjuvant, and injected for vaccination into the dorsal skin of the ostrich. As an additional immunization, the antigen solution (4mg/0.5mL) was mixed with 0.5mL of incomplete adjuvant, and injected for vaccination into the dorsal skin of the ostrich.

Purification of serum: Blood was collected from the jugular vein of the ostrich, and left for one hour. After that, the serum was separated by centrifugal separation (15000 rpm).

Yolk extract: Only yolk (250-350mL) was collected from the ostrich egg, and stirred. Only 10mL of this liquid egg yolk was used and the remainder was preserved in a freezer. 10mL of the liquid egg yolk was mixed with 75mL of TBS (20mM Tris-HCl(pH7.5), 0.15M NaCl, 0.5% NaN<sub>3</sub>). After 5mL of 10% dextran sulfate/TBS was added, the mixture was stirred for 30 minutes. Moreover, after 10mL of 1M CaCl<sub>2</sub>/TBS was added, the mixture was stirred and left for two hours or more. Supernatants were collected after the centrifugation for 30 minutes by 10000 rpm. Ammonium sulfate was added to the supernatants so as to become 40% in the final concentration and left for 12 hours or more. Then, precipitation was collected after the centrifugation by 10000 rpm. This precipitation was resuspended in 10mL of TBS, and dialyzed with

TBS.

Precipitation reaction in agarose gel: Agarose was added to 0.8% NaCl solution so as to become 1% in the final concentration and completely dissolved by heating. The solution was mounted onto a slide glass and cooled so as to form the structure having one hole in the center and six holes in the surroundings. 30 $\mu$ l of the antigen solution (2mg/ml) was dropped to the center hole, while 30 $\mu$ l of PBS, the serum or the yolk extract was dropped to the surrounding holes. After 12 hours, the existence of any precipitation line was observed.

**[0046]** FIG. 5 shows the result. The signs and numbers in this figure are as follows;

- A: the antigen (Collagen Type I)
- 1: the ostrich serum before immunization
- 2: the ostrich serum one week after immunization
- 3: the ostrich serum two weeks after immunization
- 4: the ostrich serum three weeks after immunization
- 5: the extract from the ostrich egg yolk four weeks after immunization
- 6: PBS.

**[0047]** In a similar way, the production of antibodies in the egg yolks of immunized female ostriches was examined by the precipitation reaction in agarose gel. In this examination, female ostriches were immunized every week against rat Collagen Type I. Then, examined was the production of antibodies in the yolk extract made from the ostrich at one to four weeks after the first immunization. FIG. 6 shows the result. The signs and numbers in this figure are as follows;

- A: the antigen (Collagen Type I)
- 1: the extract from the ostrich egg yolk before immunization
- 2: the extract from the ostrich egg yolk one week after immunization
- 3: the extract from the ostrich egg yolk two weeks after immunization
- 4: the extract from the ostrich egg yolk three weeks after immunization
- 5: the extract from the ostrich egg yolk four weeks after immunization
- 6: PBS

**[0048]** In FIGS. 5 and 6, the white precipitation line occurred by the antigen-antibody reaction shows the production of antibodies (also in FIG. 7). As shown in these figures, antibodies are produced both in the blood of the immunized ostrich at two weeks after immunization and in the ostrich egg yolk at four weeks after immunization. Thus, the production of antibodies was confirmed in the ostrich egg yolk.

**[0049]** The production of antibodies in the sera of female ostriches (producing eggs) and in the yolks of their eggs was further examined by the precipitation reaction in agarose gel. In this examination, female ostriches were

immunized every week against an inactivated vaccine of IBV (Infectious Bronchitis Virus). Then, examined was the production of antibodies in the sera and the egg yolks. FIG. 7 shows the result. The signs and numbers in this figure are as follows;

- A: the antigen (IBV)
- 1: PBS
- 2: the ostrich serum before immunization
- 3: the ostrich serum four weeks after immunization
- 4: the ostrich serum six weeks after immunization
- 5: the extract from the ostrich egg yolk six weeks after immunization
- 6: the extract from the ostrich egg yolk before immunization

As shown in FIG. 7, antibodies are produced in the sera of the immunized ostriches at four and six weeks after the first immunization and in the extract from the ostrich egg yolk at six weeks after the first immunization. Thus, the production of antibodies was confirmed in the ostrich egg yolk.

[Example 5]

**[0050]** In order to compare antigen-detecting abilities between the ostrich antibody and the chicken antibody, anti-CD146 antibodies were produced respectively by using the ostrich and the chicken.

**[0051]** In this experiment, female chickens and female ostriches were immunized against human CD146 proteins every other week. Then, these sera were collected six weeks after immunization, in order to purify antibodies. The antibodies were respectively labeled with HRP equivalent (0.1mg). 10ng of the CD146 protein was subjected to SDS-PAGE and transferred to a PVDF membrane. Then, the PVDF membrane was subjected to reaction with each of the above-mentioned antibodies. After the reaction, the PVDF membrane was washed and subjected to light emitting reaction with ECL, followed by exposure to a roentgen film.

**[0052]** FIG. 8 shows the result. (a) in this figure shows the result of immunoblot using respective antibodies produced from two chickens and two ostriches. As shown, the ostrich antibodies had higher antigen-detecting ability than the chicken antibodies. (b) in this figure is a graph showing relative intensity of both antibodies. The relative intensity is an average respectively of three chickens and three ostriches. The relative intensity was calculated as follows; i.e., the density of each band on the roentgen film was measured by a densitometry, and the relative intensity was calculated in which the intensity of area with no bands was set as "1". As shown, the ostrich antibodies had higher relative intensity than the chicken antibodies.

**[0053]** In a similar way, anti-CD 166 antibodies were produced respectively by using the ostrich and the chicken, in order to compare antigen-detecting abilities of both antibodies.

In this experiment, female chickens and female ostriches were immunized against human CD166 proteins every other week. Then, these sera were collected six weeks after immunization, in order to purify antibodies. The antibodies were respectively labeled with HRP equivalent (0.1mg). 20ng of the CD166 protein was subjected to SDS-PAGE and transferred to a PVDF membrane. Then, the PVDF membrane was subjected to reaction with each of the above-mentioned antibodies. After the reaction, the PVDF membrane was washed and subjected to light emitting reaction with ECL, followed by exposure to a roentgen film.

**[0054]** FIG. 9 shows the result. (a) in this figure shows the result of immunoblot using respective antibodies produced from two chickens and two ostriches. As shown, the ostrich antibodies had higher antigen-detecting ability than the chicken antibodies. (b) in this figure is a graph showing relative intensity of both antibodies. The relative intensity is an average respectively of three chickens and three ostriches. The relative intensity was calculated as follows; i.e., the density of each band on the roentgen film was measured by a densitometry, and the relative intensity was calculated in which the intensity of area with no bands was set as "1". As shown, the ostrich antibodies had higher relative intensity than the chicken antibodies. Thus, the ostrich antibodies had higher detection ability than the chicken antibodies, and it was confirmed by this Example that the ostrich antibody has qualitative advantage.

#### Industrial Applicability

**[0055]** As described, the present invention relates to an antibody produced using an ostrich and a method for production thereof. The present invention is useful not only for mass production of antibodies for medical use, which are antibodies for treatment or diagnosis, but also for other various fields where antibodies may be used.

#### **Claims**

1. An antibody produced by administering to an ostrich a human CD166 or CD 146 protein or a peptide fragment thereof as an antigen , said antibody specifically binding to said human protein or a peptide fragment thereof.
2. The antibody according to claim 1 , wherein said antibody is an antibody for medical use which is for treatment or diagnosis.
3. The antibody according to claim 2 , wherein said antibody is an antibody for medical use which is for treatment of cancer.
4. An antibody produced by administering to an Ostrich a viral protein of IBV (Infectious Bronchitis Virus),

SARS Virus or Avian Influenza Virus, or a peptide fragment thereof as an antigen , said antibody specifically binding to said protein or a peptide fragment thereof, and said antibody is for examination of IBV (Infectious Bronchitis Virus), SARS Virus or Avian Influenza Virus.

5. The antibody according to any one of claims 1 to 4, wherein said antibody is produced by firstly immunizing an infant ostrich .
6. the antibody according to any one of claims 1 to 4, wherein said antibody is produced from a yolk of an egg of a female ostrich, which has been immunized .
7. A method for producing the antibody according to anyone of claims 1 to 4, comprising a step of immunizing an ostrich against a mammalian , human or viral protein or peptide fragment thereof, as an antigen, and a step of producing a polyclonal or monoclonal antibody.
8. The method according to claim 7 , comprising a step of firstly immunizing an infant ostrich.
9. The method according to claim 7, comprising a step of immunizing a female ostrich, and a step of producing an antibody from a yolk of an egg of said female ostrich.
10. A diagnosis kit for examination, comprising the antibody according to any one of claims 1 to 4.
11. An air-conditioner filter, face mask or water filter comprising the antibody according to any one of claims 1 to 4.
12. The antibody according to claim 1 to 4 , wherein said antibody is an antibody industrially used for removing a pathogenic substance, an allergen or other antigen.

#### **Patentansprüche**

1. Antikörper, hergestellt durch Verabreichung eines humanen CD 166 oder CD 146 Proteins oder eines Peptidfragments davon als Antigen an einen Strauß, wobei der Antikörper spezifisch an das humane Protein oder Peptidfragment davon bindet.
2. Antikörper gemäß Anspruch 1, wobei der Antikörper ein Antikörper für eine medizinische Verwendung ist, die eine Behandlung oder eine Diagnose ist.
3. Antikörper gemäß Anspruch 2, wobei der Antikörper ein Antikörper für eine medizinische Verwendung ist, die für die Behandlung von Krebs ist.

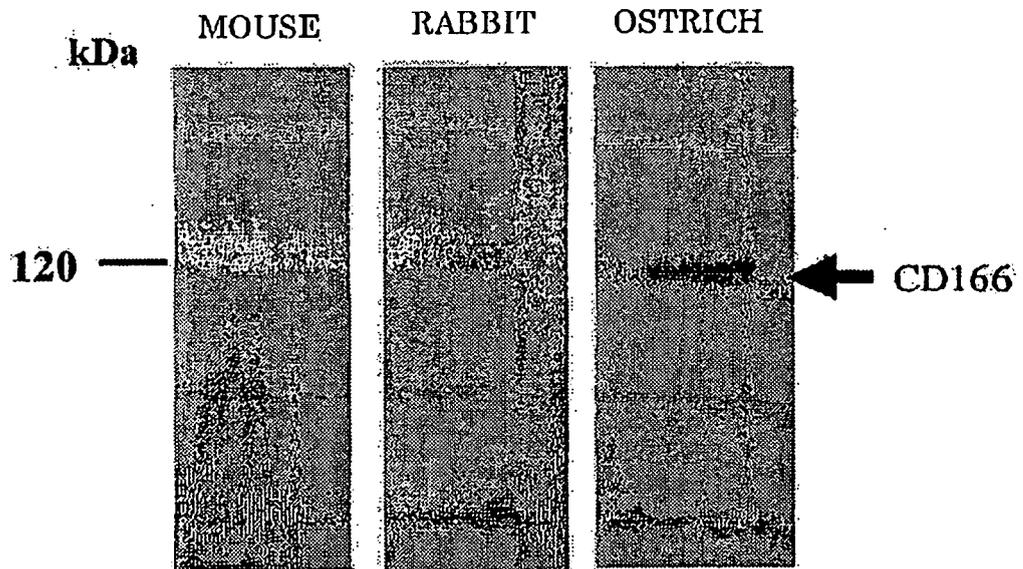
4. Antikörper, hergestellt durch Verabreichung eines viralen Proteins von IBV (infektiöses Bronchitisvirus), SARS Virus oder Vogelgrippevirus, oder eines Peptidfragments davon als Antigen an einen Strauß, wobei der Antikörper spezifisch an besagtes Protein oder einem Peptidfragment davon bindet, und der Antikörper für die Untersuchung von IBV (infektiösem Bronchitisvirus), SARS Virus oder Vogelgrippevirus ist.
5. Antikörper gemäß einem der Ansprüche 1 bis 4, wobei der Antikörper zunächst durch Immunisierung eines Straußkinds hergestellt wird.
6. Antikörper gemäß einem der Ansprüche 1 bis 4, wobei der Antikörper aus einem Dotter eines Eies eines weiblichen Straußes, der immunisiert wurde, hergestellt wird.
7. Verfahren zur Herstellung des Antikörpers gemäß einem der Ansprüche 1 bis 4, umfassend einen Schritt der Immunisierung eines Straußes gegen ein Säugetier-, humanes oder virales Protein oder Peptidfragment davon, als Antigen, und einen Schritt der Herstellung eines polyklonalen oder monoklonalen Antikörpers.
8. Verfahren gemäß Anspruch 7, umfassend einen Schritt von zunächst einer Immunisierung eines Straußenkinds.
9. Verfahren gemäß Anspruch 7, umfassend einen Schritt der Immunisierung eines weiblichen Straußes, und einen Schritt der Herstellung eines Antikörpers aus einem Dotter eines Eies des weiblichen Straußes.
10. Ein Diagnosekit für die Untersuchung, umfassend den Antikörper gemäß einem der Ansprüche 1 bis 4.
11. Klimaanlagefilter, Gesichtsmaske oder Wasserfilter, umfassend den Antikörper gemäß einem der Ansprüche 1 bis 4.
12. Antikörper gemäß Anspruch 1 bis 4, wobei der Antikörper ein Antikörper ist, der industriell für die Entfernung einer pathogenen Substanz, eines Allergens oder eines anderen Antigens verwendet wird.

#### Revendications

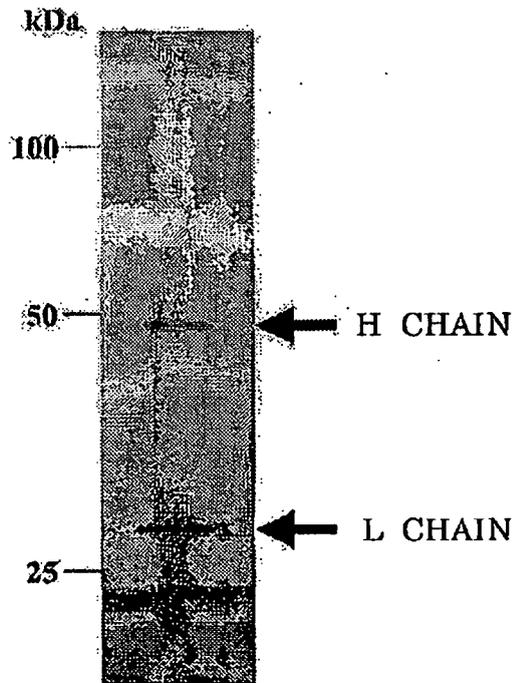
1. Anticorps produit en administrant à une autruche une protéine humaine CD 166 ou CD 146 ou un fragment peptidique de celle-ci comme antigène, ledit anticorps se liant spécifiquement à ladite protéine humaine ou audit fragment peptidique de celle-ci.

2. Anticorps selon la revendication 1, dans lequel ledit anticorps est un anticorps à usage médical destiné au traitement ou au diagnostic.
3. Anticorps selon la revendication 2, dans lequel ledit anticorps est un anticorps à usage médical destiné au traitement du cancer.
4. Anticorps produit en administrant à une autruche une protéine virale du virus de la bronchite infectieuse (IBV), du virus SARS ou du virus de la grippe aviaire ou un fragment peptidique de ceux-ci comme antigène, ledit anticorps se liant spécifiquement à ladite protéine ou audit fragment peptidique de celle-ci et ledit anticorps étant destiné à l'examen du virus de la bronchite infectieuse (IBV) ou du virus de la grippe aviaire.
5. Anticorps selon l'une des revendications 1 à 4, dans lequel ledit anticorps est produit en immunisant d'abord une autruche nouveau-née.
6. Anticorps selon l'une des revendications 1 à 4, dans lequel ledit anticorps est produit à partir du jaune d'un oeuf d'autruche qui a été immunisée.
7. Procédé pour produire l'anticorps selon l'une des revendications 1 à 4, comprenant l'étape d'immuniser une autruche contre une protéine de mammifère, une protéine humaine ou une protéine virale ou un fragment peptidique de celles-ci comme antigène et l'étape de produire un anticorps polyclonal ou monoclonal.
8. Procédé selon la revendication 7, comprenant l'étape d'immuniser d'abord une autruche nouveau-née.
9. Procédé selon la revendication 7, comprenant l'étape d'immuniser une autruche femelle et l'étape de produire un anticorps à partir du jaune d'un oeuf de ladite autruche.
10. Kit de diagnostic destiné à l'examen, comprenant l'anticorps selon l'une des revendications 1 à 4.
11. Filtre de climatisation, masque facial ou filtre d'eau comprenant l'anticorps selon l'une des revendications 1 à 4.
12. Anticorps selon la revendication 1 à 4, dans lequel ledit anticorps est un anticorps à usage industriel, utilisé pour supprimer une substance pathogène, un allergène ou un autre antigène.

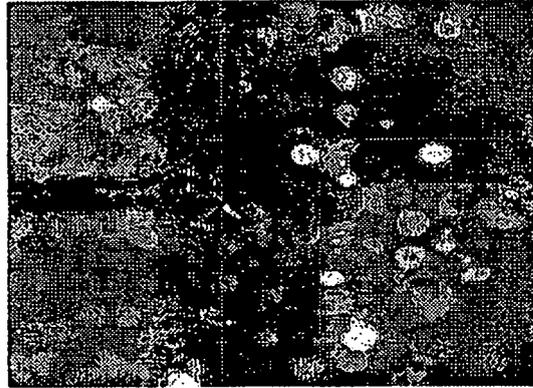
[Fig.1]



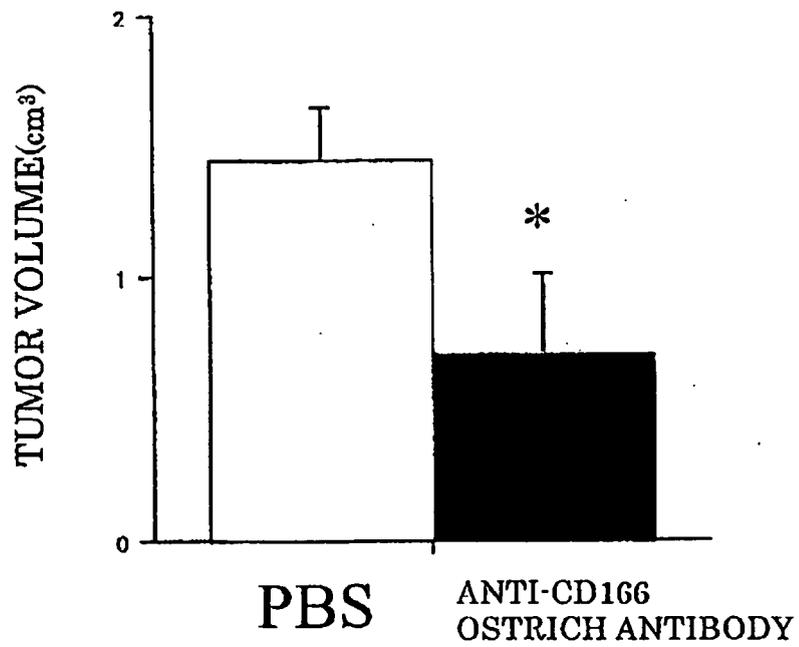
[Fig.2]



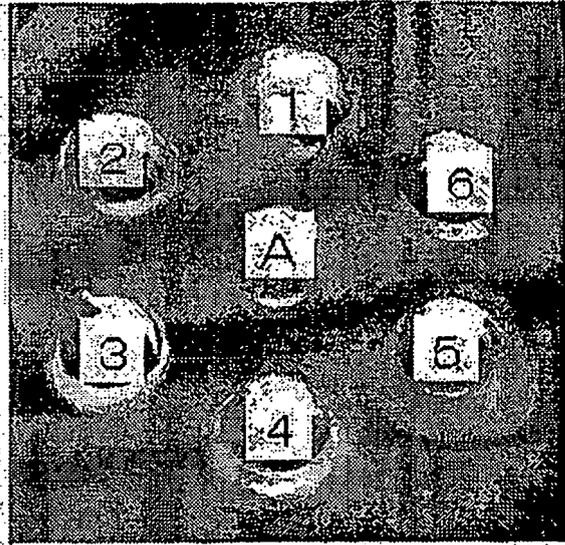
[Fig.3]



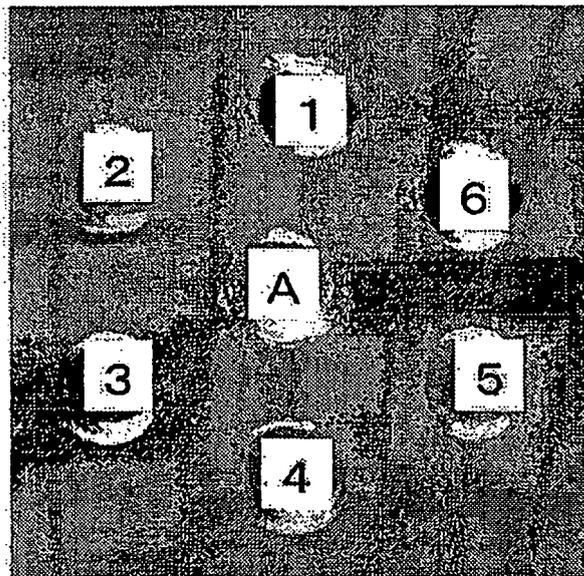
[Fig.4]



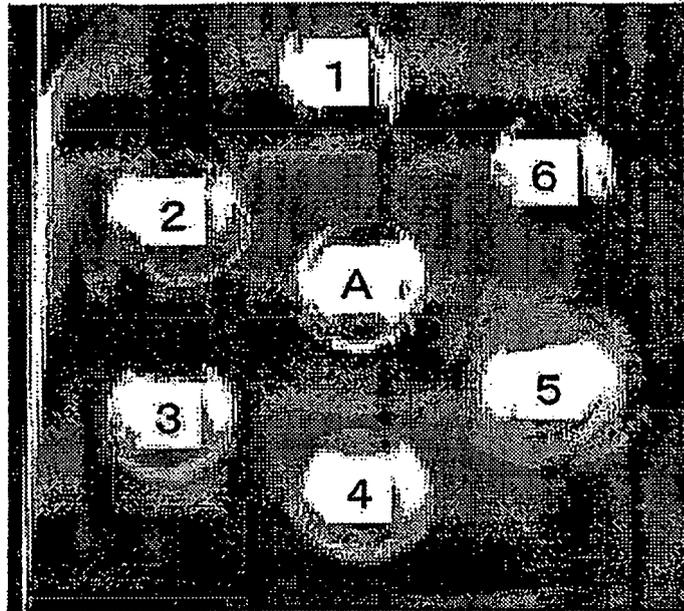
[Fig.5]



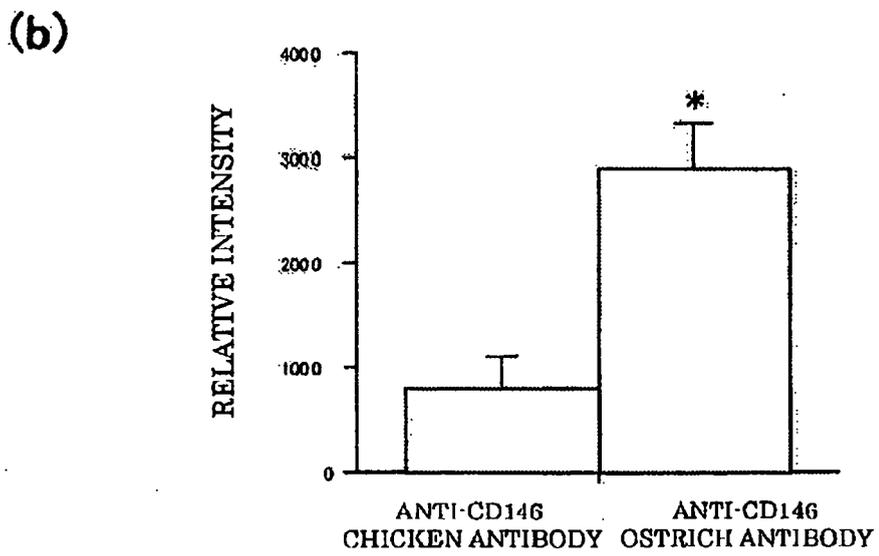
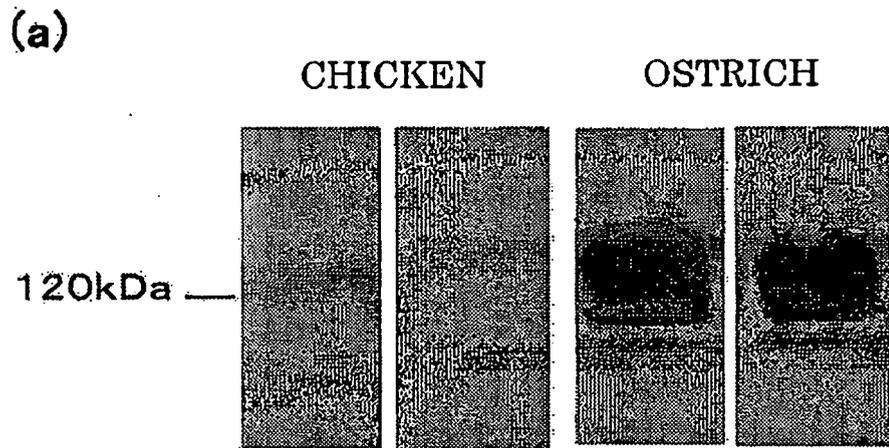
[Fig.6]



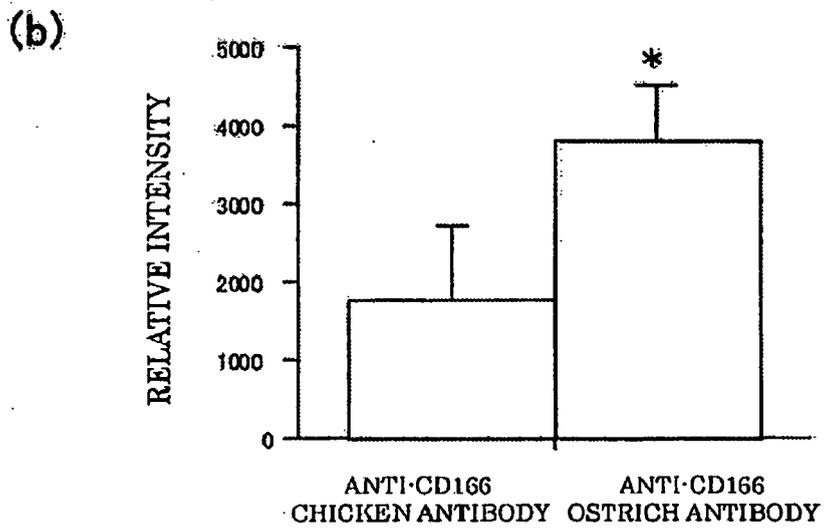
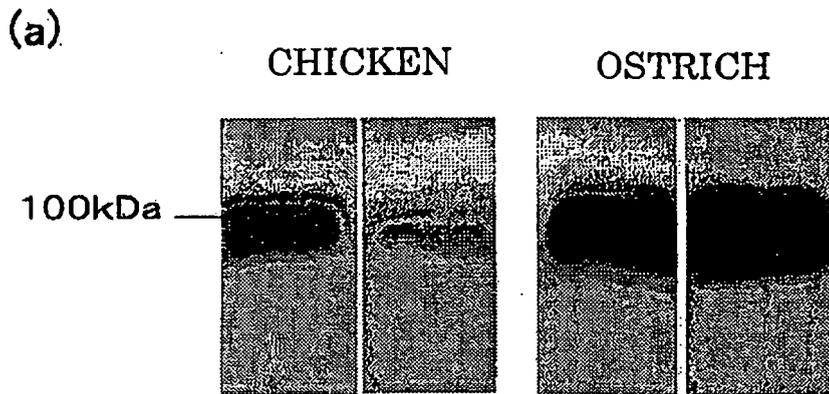
[Fig.7]



[Fig.8]



[Fig.9]



**REFERENCES CITED IN THE DESCRIPTION**

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