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(54) **TRANSPOSASE AND METHOD OF GENE MODIFICATION**

(57) A transposase encoded by the Tol2 element; a polynucleotide encoding the same; a method of modifying the gene structure of a cell (preferably a vertebrate cell) by using the above protein; a method of modifying the function of a cell by modifying the gene structure

thereof; and a cell having been modified in function by these methods. Also the structure of a cis element necessary in gene transfer is clarified and presented.

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**Description**Technical Field

5 **[0001]** The present invention relates to a new protein having a transposase-like activity, a transposase composed of the above protein, a method of modifying the gene structure of a cellular gene by using these protein and transposase, a method of modifying the function of a cell by this method, a method of introducing a gene by this method, a plasmid used for this method, and a cell having been modified in function by this method.

10 Background Art

**[0002]** The medaka fish (*Oryzias latipes*) is a teleost inhabiting in East Asia and has been used for studying vertebrate genetics. The mutations at the *i* locus of the medaka fish cause amelanotic skin and red-colored eyes. This *i* locus is known to encode a gene for tyrosinase. From one of the *i* alleles, *i*<sup>4</sup>, DNA of about 4.7-kb was cloned and found to have a transposon-like sequence; that is to say, it included open reading frames homologous to transposases of transposons of the hAT family including hobo of *Drosophila*, Ac of maize and Tam3 of snapdragon, and short terminal inverted repeats. This medaka element was named Tol2. The laboratory strains of the medaka fish contain about 10 copies of this element per haploid genome.

20 **[0003]** In the *i*<sup>4</sup> mutant fish, the Tol2 element found in the tyrosinase gene locus has been shown by PCR to be excised from the target locus during embryonic development (Koga et al., 1996).

25 **[0004]** Zebrafish (*Danio rerio*), as well as the medaka fish (*Oryzias latipes*), is a small teleost and has been developed as a model animal to study vertebrate genetics and development (Takeuchi, 1966; Yamamoto, 1967; Streisinger et al., 1981). In zebrafish, large-scale chemical mutagenesis screens have been performed (Driever et al., 1996; Haffter et al., 1996), and, to facilitate cloning of the mutated genes, an insertional mutagenesis method using a pseudotyped retrovirus has been developed and performed (Lin et al., 1994; Gaiano et al., 1996; Amsterdam et al., 1997). Also, in an attempt to develop transposon technologies that would allow enhancer trap and gene trap screens to be performed, transposition of transposons of the Tc1/mariner family in fish has been tested and demonstrated (Ivics et al., 1997; Raz et al., 1997; Fadool et al., 1998). Although these results are encouraging, neither highly efficient transgenesis nor insertional mutagenesis methods using a transposon have not yet been developed.

30 **[0005]** The present inventors have been interested in developing novel transposon technologies using the Tol2 element. As a first step towards this goal, the present inventors developed a transient embryonic excision assay using zebrafish embryos, in which zebrafish fertilized eggs were injected with a plasmid DNA harboring the Tol2 element, showed that the Tol2 element was excisable from the injected plasmid DNA, and indicated that the Tol2 element is an autonomous member and is active in zebrafish (Kawakami et al., (1998) *Gene* **225**, 17-22). Although the DNA sequence of the Tol2 element is similar to those of transposases of transposons of the hAT family, neither an active enzyme, which can function in trans, nor cis-elements essential for the excision reaction have been identified. In order to develop the Tol2 element as a useful tool for transgenesis and insertional mutagenesis, it is necessary to dissect and characterize cis and trans requirements. The functional transposase encoded by the Tol2 element had not yet been identified prior to the present invention.

40 Disclosure of the Invention

**[0006]** The present invention first aims to identify mRNA transcribed from the Tol2 element injected in zebrafish embryos. Secondly, in order to determine whether the transcript encodes an active enzyme or not, the present invention develops a novel assay method, in which zebrafish fertilized eggs are co-injected with RNA synthesized in vitro using the Tol2 cDNA as a template and a plasmid DNA harboring a nonautonomous Tol2 element, which has a deletion in the transposase coding region.

**[0007]** The present invention also identifies the active trans-factor and essential cis-elements, that function in excision of the Tol2 element in zebrafish.

50 **[0008]** Consequently, the present invention results in a new protein encoded by the Tol2 element and a polynucleotide encoding the same. Also the present invention, by using the above protein, results in a method of modifying the gene structure of a cell, preferably the gene structure of a vertebrate, in a method of modifying the function of a cell by modifying the gene structure thereof, and in a cell having been modified in function by these methods. Furthermore, the present invention discloses the cis-element structures essential for transposition, and presents the same.

55 **[0009]** The present invention relates to a protein having the transposase-like activity, which has an amino acid sequence shown on sequence number 2 of SEQUENCE LISTING, an optionally substituted amino acid sequence with any replacements or deletions in part of the original amino acid sequence, or an optionally substituted amino acid sequence with addition of other amino acids to the original amino acid sequence. Also the present invention relates to

a transposase comprising the said protein.

**[0010]** Further, the present invention relates to the nucleic acid encoding the said protein, wherein the nucleic acid is preferably DNA having a nucleotide sequence shown on sequence number 1 of SEQUENCE LISTING or DNA which can hybridize to the said DNA, or is the corresponding RNA.

**[0011]** The present invention reveals that the said protein has a transposase-like activity which catalyzes transposition of the above transposon, and relates to a method of modifying the gene structure comprises the excision in part of a gene in a cell, preferably a vertebrate cell, or the insertion of the excised part into any other locus in the presence of the said protein or the nucleic acid which can produce the said protein. It is preferable that the said excised gene has nucleotide sequences containing at least one inverted repeat (the Angel elements ) in forepart of its nucleotide sequence.

**[0012]** Further, the present invention relates to a method of inserting a foreign gene into a gene of a cell, and a method of modifying a function of a cell based on gene expression, and furthermore relates to a cell having been modified in function by the said method.

**[0013]** Also, the present invention relates to a plasmid used in these methods and, more in detail, a plasmid which contains DNA having a nucleotide sequence that includes at least one inverted repeat sequence in the forepart of its nucleotide sequence.

**[0014]** Furthermore, in a method of inserting any DNA into the genomic DNA of a vertebrate, the present invention relates to a method of inserting any DNA into the genomic DNA of a vertebrate which is characterized by operating the said insertion of DNA autonomously using the transposase activity, wherein a preferable DNA is the Tol2 element and the vertebrate is fish.

#### Brief Description of the Drawings

##### **[0015]**

Figure 1 shows the structure of the Tol2 plasmid and the transcript, and the structure of cDNAs described in the present invention. The dotted lines indicate introns. The inverted repeat (the Angel elements) in the first intron and positions of primers used in the present invention are shown by arrows.

Figure 2 shows a comparison of amino acid sequences of transposases of the Tol2 element described in the present invention and the Ac element.

Figure 3 shows a scheme for the transient embryonic excision assay by co-injection described in the present invention. Primers (tyr-ex4f and tyr-ex5r) used to detect the excision products are shown by arrows.

Figure 4 shows photos, substitutes for drawings, which show the results of the PCR analysis of the excision reaction in zebrafish embryos described in the present invention.

Figure 5 shows the structures of the (Tol2-tyr) $\Delta$ RV plasmid used for transposition of the Tol2 element into the genome, the Tol2-tyr plasmid and Tol2 cDNA. The black line in the upper part of Figure 5 shows the probe used for Southern blot analysis.

Figure 6 shows photos, substitutes for drawings, which show the results of Southern blot analysis of F1 progeny fish from each parental fish (ff-1 and ff-7), wherein the presence of Tol2 is identified (Figure 6, A) and show the results of PCR (Figure 6, B).

Figure 7 shows the nucleotide sequences surrounding the Tol2 element inserted in the genome of F1 progeny A, B and C from ff-7.

#### Best Mode for Carrying Out the Invention

**[0016]** Previously, the present inventors injected the Tol2-tyr plasmid, a plasmid harboring the Tol2 element cloned from the tyrosinase gene locus, into zebrafish fertilized eggs and showed that the Tol2 element is excisable from the injected plasmid DNA (Kawakami et al., 1998). In order to identify a transcript encoding a putative transposase activity, total RNA from embryos injected with the Tol2-tyr plasmid were prepared. The present inventors first performed 3' RACE using four pairs of nested primers that annealed different parts of the Tol2 sequence.

**[0017]** Nested forward primers used to perform 3' RACE are:

Tol2f2; 5'-TTGGTCAGACATGTTTCATTG-3' and

Tol2f3; 5'-ATGTTTCATTGGTCCTTTGGA-3',

Tol2f4; 5'-ATAGCTGAAGCTGCTCTGATC -3' and

Tol2f5; 5'-CTGCTCTGATC ATGAAACAG-3',

Tol2f8; 5'-GCTTAATAAAGAAATATCGGCC -3' and

Tol2f9; 5'-AATATCGGCCTTCAAAAGTTCG-3', and

Tol2f12; 5'-CTGTAATCAGAGAGTGTATGTGTA -3' and

Tol2f13; 5'-ATTGTTACATTTATTGCATACAAT -3'.

**[0018]** cDNAs with polyadenylation were successfully amplified by 3' RACE using Tol2f8 and Tol2f9, and Tol2f4 and Tol2f5, but not by 3' RACE using Tol2f2 and Tol2f3, and Tol2f12 and Tol2f13.

**[0019]** Then, using nested reverse primers designed to perform 5' RACE,

Tol2r4; 5'-CTCAATATGCTTCCTTAGG -3' and

Tol2r5; 5'-CTTCCTTAGGTTTGATGGCG-3',

5' RACE was performed and the full-length Tol2 transcript of 2156 nucleotides was identified (Figure 1).

**[0020]** The cDNA sequence obtained is shown in sequence number 1 of SEQUENCE LISTING.

**[0021]** Figure 1 shows the structures of the Tol2 plasmids and its transcript. The top line of Figure 1 shows the full-length Tol2 (Tol2-tyr). Dotted lines in the figure show introns. The inverted repeat (the Angel elements) in the first intron and positions of the said primers are shown by arrows. The lower three lines of Figure 1 show the results of 3' RACE and 5' RACE. In each case, introns are shown as dotted lines.

**[0022]** The fifth line shows the structure of the full-length mRNA. The translated region corresponds to the nucleotide sequence between the 85th (ATG) and the 2032nd nucleotide (TAG) of cDNA of sequence number 1.

**[0023]** The two lines in the bottom show the structures of deletion mutants, (Tol2-tyr)ARV, and (Tol2-tyr)  $\Delta$ in1 $\Delta$ RV.

**[0024]** In the 5' RACE analysis, aberrant transcripts that started from the plasmid sequence and jumped into cryptic splice acceptor sites in the first exon of the Tol2 element were also found (data not shown). These transcripts were not studied further.

**[0025]** DNA sequencing of the cDNA revealed the exon-intron structure of the Tol2 element (i.e., four exons and three introns) (as shown in the upper part of Figure 1). The cDNA encodes a protein of 649 amino acids. The amino acid sequence of this protein is shown in sequence number 2 of SEQUENCE LISTING.

**[0026]** Although the Tol2 element had been known to have a transposon-like sequence, the present invention for the first time identified that the Tol2 element encodes a protein and the expression of the protein described here generates the function. That is to say, the present invention results in a new protein encoded by the Tol2 element and also a polynucleotide which encodes the protein described here.

**[0027]** Figure 2 shows a comparison of amino acid sequences of the protein described in the present invention and a known transposase of a transposon of the hAT family. This comparison shows these proteins are similar, especially in the middle part (Figure 2). But the amino acid sequences of NH<sub>2</sub>- and COOH-terminus rather varies.

**[0028]** In order to determine whether the protein (the Tol2 transcript) identified in the present invention encodes a functional enzyme, a new transient embryonic excision assay by co-injection was developed and, by using this method, identification of the enzymatic activity was performed.

**[0029]** Zebrafish fertilized eggs were co-injected with mRNA synthesized in vitro using the cDNA shown in the sequence number 1 as a template and the (Tol2-tyr) $\Delta$ RV plasmid containing (Tol2-tyr) $\Delta$ RV (see Figure 1), which has a deletion of the nucleotides between the EcoRV sites of the Tol2 element. About 8 hours after the co-injection, DNA was prepared from each embryo and analyzed by PCR using primers, tyr-ex4 f and tyr-ex5r,

tyr-ex4f: 5'- GCTACTACATGGTGCCATTCCT-3'

tyr-ex5r : 5'-CACTGCCAGATCTGCTGGGCTT-3'

which were prepared based on the sequence adjacent to the Tol2 element. Figure 3 shows a scheme of this method and these primers are shown in Figure 4A.

**[0030]** PCR products of about 250bp, indicative of excision of the Tol2 element from (Tol2-tyr) $\Delta$ RV plasmid, was amplified in all embryos analyzed (56 out of 56, see Figure 4B lanes 1-10). This PCR product was never detected from embryos injected only with the (Tol2-tyr) $\Delta$ RV plasmid DNA (0 out of more than 50, see Figure 4B lanes 11-20).

**[0031]** The PCR products from six different embryos were cloned and sequenced. Three of them had the wild-type medaka fish tyrosinase gene sequence (Figure 4C, excision product a), indicating that precise excision had occurred, and the other three had nearly wild type sequences with addition of a few nucleotides (Figure 4C, excision products b and c), characteristic to excision of transposons of the hAT family (Pohlman et al., 1984; Sutton et al., 1984; Koga et al., 1996; Kawakami et al., 1998), suggesting that the excision event in this experiment is catalyzed by a transposase-like activity.

**[0032]** These results, i.e., when co-injected with the mRNA which has the nucleotide sequence shown in sequence number 1 of the present invention, the PCR product characteristic to excision of the transposon was obtained and, when co-injected without the mRNA, such PCR products were not obtained, indicate that the protein (the Tol2 transcript) described in the present invention encodes a functional transposase, which can catalyze the excision. Further, these results show that the (Tol2-tyr) $\Delta$ RV plasmid contains sequences of cis-elements essential for the excision.

**[0033]** Figure 4 shows the results of this experiment and arrows in Figure 4A show positions and directions of the primers used in the analyses. The upper panel of Figure 4B is a photo, a substitute for a drawing, which shows the PCR products using primers tyr-ex4f and tyr-ex5r, and the lower panel shows the PCR products using primers Tol2f1 and Tol2r3. In lanes 1-10, zebrafish embryos were injected with both the (Tol2-tyr) $\Delta$ RV plasmid and the Tol2 mRNA, in lanes 11-20 the (Tol2-tyr) $\Delta$ RV plasmid alone was injected, and in lane G and P PCR products were amplified from 50ng of the zebrafish genomic DNA and from 10pg of the (Tol2-tyr) $\Delta$ RV plasmid DNA. Figure 4C shows the DNA sequences of the excision products obtained in the above experiments. The Tol2 sequence is shown in bold and 8bp direct repeat sequences flanking to the Tol2 element are underlined.

**[0034]** It was noted that, although the excision products could be detected after a single-round PCR amplification in the experiments in the present invention, two rounds of PCR were required in the previous analysis, in which fertilized eggs were injected with a sole plasmid DNA containing the full-length Tol2 element without mRNA prepared in vitro. The higher efficiencies of the excision reaction observed here can be explained as more transposases were supplied by RNA injection than those supplied by DNA injection.

**[0035]** The first intron of the Tol2 element contains about 300bp of large inverted repeats, and the repeat was recently identified as the Angel element (Izsvak et al., 1999) (see Figure 1). To test whether the sequence in the intron is essential for excision, we constructed the (Tol2-tyr)  $\Delta$ in1 $\Delta$ RV plasmid, containing (Tol2-tyr)  $\Delta$ in1 $\Delta$ RV (see the bottom of Figure 1), which completely lacked the sequences of the first intron, and its activity was analyzed by co-injection with the Tol2 mRNA as the said method. This result is shown in Figure 4D.

**[0036]** The upper panel of Figure 4D is a photo, a substitute for a drawing, which shows PCR products amplified using primers tyr-ex4f and tyr-ex5r and, the lower panel shows PCR products amplified using primers Tol2f1 and Tol2r3. In lanes 1-8, zebrafish embryos were injected with both the (Tol2-tyr)  $\Delta$ in1 $\Delta$ RV plasmid and the Tol2 mRNA, in lanes 9-12 both the (Tol2-tyr) $\Delta$ RV plasmid and the Tol2 mRNA were injected, and, in lanes 13-16, the (Tol2-tyr)  $\Delta$ in1 $\Delta$ RV plasmid alone was injected. Lane P shows the PCR product amplified from 10pg of the (Tol2-tyr)  $\Delta$ in1 $\Delta$ RV plasmid DNA.

**[0037]** In lanes 9-12, the said experiment was conducted as controls and the PCR products indicating the excision could be detected, but the excision product could not be detected in lanes 1-8 when the plasmid lacking the intron part (0 out of 16, see Figure 4D lanes 1-8) was used, suggesting that the first intron contains cis-elements essential for excision.

**[0038]** Further, the (Tol2-tyr)  $\Delta$ in1 plasmid, containing (Tol2-tyr)  $\Delta$ in1 which restored the $\Delta$ RV deletion and was about the same size as the (Tol2-tyr)  $\Delta$ RV plasmid, i.e., which has a deletion between the 644th and 2163rd nucleotides of the Tol2 element, was also examined by the co-injection assay, but PCR product indicating the excision could not be obtained (0 out of 16, data not shown).

**[0039]** Although, further analyses using smaller deletions and point mutations in the first intron sequence will be needed to define the essential cis-sequences for excision precisely, since these results show that the intron part is essential for the excision and the intron contains the Angel elements as inverted repeats, it can be thought that the inverted repeats are essential sequences for the excision described in the present invention.

**[0040]** Thus, we successfully identified for the first time the transcript (the protein described in the present invention)

encoded by the Tol2 element and also identified a transposase activity of this protein and cis-sequences essential for transposition. These discoveries will lead to a biochemical characterization of the Tol2 transposase.

**[0041]** On the other hand, transposition of transposons belonging to the Tc1/mariner family into the zebrafish genome has been reported (Raz et al., 1997; Fadool et al., 1998). In the experiments described in their reports, zebrafish one-cell-stage embryos were co-injected with transposase RNA transcribed in vitro and transposon vectors containing essential cis-sequences.

**[0042]** While a transposon belonging to a different family may have different specificities and efficiencies for insertion into the genome, by a method of present invention which is a novel transposon technology in fish using the Tol2 element, since its transposon excision procedure has been carried out in the way of Raz et al., it might be possible to transpose DNA such as the Tol2 element into the genome in the way conducted by using transposons of the Tc1/mariner family.

**[0043]** Therefore, we tested whether the Tol2 element can be inserted into the zebrafish genome by transposition. It is known that the zebrafish genome does not contain the Tol2 element.

**[0044]** To test whether the Tol2 element encodes a transposase that can catalyze transposition, zebrafish fertilized eggs were co-injected with RNA transcribed in vitro using the Tol2 cDNA as a template, which encoded a putative transposase, and a plasmid DNA harboring the (Tol2-tyr) $\Delta$ RV element, which has a deletion in part of the region presumed to code the transposase.

**[0045]** The structures of (Tol2-tyr) $\Delta$ RV plasmid and Tol2 cDNA are shown in Figure 5. 3' and 5' indicate the direction of transcription.

**[0046]** The injected eggs were raised to adulthood and mated to non-injected fish. And the progeny fish were analyzed for the presence of the Tol2 sequence.

**[0047]** Two out of eight injected fish could transmit the Tol2 sequence to their progeny. These two fishes were named ff-1 (founder fish-1) and ff-7 (founder fish-7).

**[0048]** Two fish out of 68 F1 fish from the ff-1 fish had the Tol2 sequence. These two fish had the sequence of the plasmid portion as well as the Tol2 sequence. On the other hand, 25 fish out of 50 F1 fish from the ff-7 fish had the Tol2 sequence. These 25 fish did not have the plasmid sequence and were classified into three groups, A, B and C, from the result of Southern blot shown in Figure 6A. 7 fish were grouped as A, 3 fish as B, and 15 fish as C.

**[0049]** Figure 6A is a photo, a substitute for a drawing, which shows the result of Southern blot analysis using a probe shown in Figure 5, in which DNA samples prepared from caudal fins of F1 fish from ff-1 and ff-7 were digested with EcoRV. Two samples from ff-1 showed the same pattern but samples from ff-7 showed three patterns, A, B and C.

**[0050]** Then, PCR analyses of F1 fish from ff-1 and ff-7 were performed. Primers used were shown in Figure 5 as PCR1, PCR2 and PCR3. As controls, zebrafish genomic DNA (G) and genomic DNA plus (Tol2-tyr)ARV plasmid DNA (G+P) were used. In F1 fish from ff-7, PCR products using PCR2 and PCR3 could not be amplified. This indicated that progeny fish from ff-7, unlike progeny fish from ff-1, did not have the plasmid sequence flanking to the Tol2 element.

**[0051]** From the ff-7 progeny fish, DNA fragments containing the Tol2 sequence and the flanking region were cloned by inverse PCR and sequenced. In each three case, A, B and C, the Tol2 sequence was surrounded by zebrafish genomic sequences and 8bp duplications were created adjacent to the insertion. 8bp duplications at both ends of the Tol2 element are characteristic to integration of transposons of the hAT family, indicating that the integration described here was catalyzed by a transposase.

**[0052]** Figure 7 shows the determined nucleotide sequences of three types, A, B and C. Tol2 in Figure 7 shows the Tol2 sequence. In A repeats of  $\lceil$  CTCAACTG  $\rfloor$ , in B repeats of  $\lceil$  TATAGAGA  $\rfloor$ , and in C repeats of  $\lceil$  GTTTTCAG  $\rfloor$  were created at both ends of and adjacent to the Tol2 sequence.

**[0053]** In the vertebrate cultured cells and the germ line, transposition activities of Sleeping Beauty which was reconstituted and activated artificially (Ivics, Z., et al., Cell, 91, 501-510 (1977)), Tc3 of *C. elegans* (Raz, E., et al., Current Biology, 8, 82-88 (1977)) and mariner of *Drosophila* (Fadool, J.M., et al., Proc. Natl. Acad. Sci. USA, 95, 5182-5186 (1988)), all belonging to the Tc1/mariner family, have been reported. No autonomous transposon activity residing endogenously in any vertebrate genome, however, has been reported.

**[0054]** The present invention is the first report that identified an autonomous element from a vertebrate genome and also for the first time reported a functional transposase activity in vertebrate.

**[0055]** Therefore, the present invention relates not only to a method to excise a gene autonomously in vertebrate but also to a method to insert the excised gene into any locus or any gene on the genome.

**[0056]** The protein in the present invention has the amino acid sequence shown in sequence number 2 of SEQUENCE LISTING, but all of the amino acids shown there are not necessarily required, and the protein in the present invention can include a protein having the transposase activity described in the present invention or similar activities described above (both of these are called transposase-like activities) and also can include a protein having replacement or deletion in part of amino acids of the above protein, or having addition of any other amino acids to the above protein. And preferably it has the amino acid sequence derived from the Tol2 element. Further, the protein in the present invention includes a protein which is produced from mRNA having the nucleotide sequence corresponding to sequence number 1 of SEQUENCE LISTING.

**[0057]** The nucleic acid in the present invention encodes the amino acid sequence which is related to the said protein, and preferably which has the polynucleotide having the sequence shown in sequence number 1 of SEQUENCE LISTING. The nucleic acid in the present invention includes not only the said nucleotide sequence but also a nucleotide sequence which can hybridize to the said nucleic acid, preferably under stringent conditions.

5 **[0058]** As for a method of modifying the gene structure of a gene in a cell in the presence of the protein in the present invention or the nucleic acid which can produce the said protein, by introducing the protein or the nucleic acid, for instance the mRNA which can produce the protein described here, and, at the same time, by introducing genes including a gene to be transposed, for instance a plasmid, the gene structure in a cell can be modified by the enzymatic activity of the protein in the present invention. The modification in the present invention is involved preferably in autonomous transposition. The cell is preferably an animal cell, more preferably a vertebrate cell, and much more preferably fish cell including a zebrafish cell.

10 **[0059]** Genes containing the said gene to be transposed can be substances which do not exist in a natural cell, such as a plasmid carrying a foreign gene to be transposed, and also can be a genomic gene existing in a natural cell. In this case, cis-elements required for transposition could be added to the gene if necessary. The gene to be transposed is preferably a transposon, in certain circumstances, it may be a gene that insert a normal gene into the cell which has a disease caused by abnormalities of genes of various kinds.

15 **[0060]** Further, a method of modification in the present invention may only include the excision of part of a gene in a cell such as an inserted plasmid, however, may also include the insertion of all or part of the gene excised by this method, into any gene.

20 **[0061]** A gene excised in a method of modification in the present invention preferably has nucleotide sequences containing at least one inverted repeat in the forepart of its nucleotide sequence. The inverted repeat is thought as a cis-element or part of cis-elements for transposition of the gene.

25 **[0062]** Further, the present invention, by using the said methods of modification, relates to a method of introducing a foreign gene into a gene of a cell and to a method of modifying a function of a cell based on expression of the gene. By performing the said methods, for example, it is possible for a foreign gene on a plasmid to be transposed into the genome in a cell, and for a new gene, which the cell concerned does not contain originally, to be inserted into a cell. Further, by expression of the newly inserted gene, it is possible to modify a function of a cell. Furthermore, the present invention can result in a cell, whose function has been modified by this method. The said cell is preferable as a cell described in this method.

30 **[0063]** As a plasmid in the present invention which contains the nucleotide sequence having at least one inverted repeat in the forepart of its nucleotide sequence, an optional substitute is to mediate transposition of a gene therein, contains a region containing at least one inverted repeat and a gene to be transposed near the repeat, and is easy to be inserted into a cell.

### 35 Examples

**[0064]** The present invention will be described by Examples below more precisely, but these Examples do not limit the present invention.

40 **[0065]** In the experiments in the present invention, eggs for injection were obtained from zebrafish strains, Tuebingen, TL and brass and were used for the following experiments.

#### Example 1 (cloning of cDNA)

45 **[0066]** Zebrafish fertilized eggs were injected with the (Tol2-tyr) plasmid and, 9 hours after the injection, total RNA was extracted from 50 of zebrafish embryos with Tri Zol Reagent (Life Technologies, Inc.) and about 3 $\mu$ g of the total RNA obtained was used for 3' RACE and 5' RACE, respectively.

**[0067]** Nested forward primers used to perform 3' RACE are:

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Tol2f2; 5' -TTGGTCAGACATGTTTCATTG-3' and  
 Tol2f3; 5' -ATGTTTCATTGGTCCTTTGGA-3' ,  
 Tol2f4; 5' -ATAGCTGAAGCTGCTCTGATC -3' and  
 Tol2f5; 5' -CTGCTCTGATC ATGAAACAG-3',  
 Tol2f8; 5'-GCTTAATAAAGAAATATCGGCC -3' and  
 Tol2f9; 5'-AATATCGGCCTTCAAAAGTTCG-3', and  
 Tol2f12; 5'-CTGTAATCAGAGAGTGTATGTGTA -3' and  
 Tol2f13; 5'-ATTGTTACATTTATTGCATACAAT -3'.

Nested reverse primers used for 5' RACE are:

Tol2r4; 5'-CTCAATATGCTTCCTTAGG -3' and

Tol2r5; 5'-CTTCCTTAGGTTTGATGGCG-3'.

**[0068]** The 3' RACE and 5' RACE products were gel-extracted, cloned with TOPO TA Cloning Kit (Invitrogen, Inc.) and sequenced using the ABI PRISM 310 Genetic Analyzer.

**[0069]** The sequence determined is shown in sequence number 1 of SEQUENCE LISTING and the amino acid sequence of its translated region is shown in sequence number 2 of SEQUENCE LISTING.

**[0070]** Also, the summary is shown in Figure 1. The numbers in the parentheses are bp from the 5' end of the Tol2 element. DDBJ/EMBL/Genbank accession number for the cDNA sequence is AB032244.

Example 2 (Construction of the (Tol2-tyr)  $\Delta$ in1 $\Delta$ RV plasmid)

**[0071]** The (Tol2-tyr) $\Delta$ in1 $\Delta$ RV plasmid was first constructed by replacing the NruI-NspV of the (Tol2-tyr) plasmid with the NruI-NspV fragment of the cDNA and the resulting plasmid was digested with EcoRV and self-ligated.

Example 3 (mRNA synthesis, injection to embryos and PCR analysis)

**[0072]** The cDNA encoding the entire coding region of the transposase was cloned in pBluescript SK+ (Stratagene), linearized, digested with proteinase K and phenol/chloroform extracted. mRNA was generated by in vitro transcription by using T7 RNA polymerase and the mCAP mRNA Capping kit (Stratagene). The concentration and the size of the transcript were examined on agarose gel electrophoresis.

**[0073]** Zebrafish fertilized eggs were injected with 1-2nl of a DNA solution (~25ng/ $\mu$ l of a plasmid DNA) with or without the mRNA (~5ng/ $\mu$ l of the Tol2 mRNA) and incubated at 28°C for ~8 hours. Each embryo was soaked in 50 $\mu$ l of 10mM EDTA, 10mM Tris-HCl (pH8.0), 200 $\mu$ g/ml proteinase K and incubated at 50°C for 3 hours.

**[0074]** Then 1 $\mu$ l of the lysed embryo was used for PCR (35 cycles of 94°C 30sec, 55°C 30sec and 72°C 30sec) using tyr-ex4f and tyr-ex5r primers (Kawakami et al., 1998). The PCR products were analyzed on 2% agarose gel electrophoresis. The result is shown in Figure 4.

**[0075]** For the DNA sequencing analysis, the PCR products were gel-extracted, cloned with TOPO TA Cloning (Invitrogen) and sequenced. The presence of the injected plasmid DNA in each sample was verified by PCR (25 cycles of 94°C 30sec, 55°C 30sec and 72°C 30sec) using

Tol2f1 (5'-TCCACCCATGCTTCCAGCAGTA-3') and

Tol2r3 (5'-CGTTGTGGTTGCAATCCATTCAAC-3') primers.

Industrial Applicability

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**[0076]** The present invention results in a new protein having a transposase-like activity of a gene and the nucleic acid encoding the same.

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**[0077]** Further, the present invention discloses that a transposase of a different family is able to generate an enzymatic activity which can catalyze transposition of a gene in a vertebrate cell, and greatly contributes to the development of technologies concerning the transposition of a gene in vertebrate and the analyses of mutants generated by the said transposition. On the other hand however, since recent gene technologies are extending from modification of a cell to modification of an organism, a method of transposition of a gene in a cell in the present invention is expected not to be limited only to the modification of a cell but also applicable to modification of the structures and functions of genes of mammals in the medical and agricultural fields as one of the methods for modifying the organism of the traits.

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It can be expected to be a powerful method especially for the gene therapy and the improvement of fish breeding.

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## SEQUENCE LISTING

5 <110> Japan Science and Technology Corporation

10 <120> TRANSPOSASE AND METHOD OF GENE MODIFICATION

15 <130> JA905073

<150> JP 11-345508

20 <151> 1999-12-03

<150> JP 2000-109033

25 <151> 2000-04-11

30 <160> 2

<210> 1

35 <211> 2156

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40 <213> *Oryzias latipes*

<400> 1

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50 ataacagtca atcagtgga gaaaatggag gaagtaigtg attcaicagc agctgcgagc 120

55 agcacagicc aaaatcagcc acaggatcaa ggcacccgt ggccgtatc| tcgccaattc 180

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1 ttttcttttaa gtggigtaaa taaagaitca itcaagaiga aatgtgtcct ctgtctcccg 240  
 5  
 2 ctttaataaag aaataicggc ctcaaaaagt tcgccaatcaa acctaaggaa gcatattgag 300  
 10  
 3 agaatgcacc caaattacct caaaaactac tciaaaatga cagcacagaa gagaaagatc 360  
 15  
 4 gggacctcca cccatgcttc cagcagtaag caactgaaag ttgactcagt ttcccagtc 420  
 20  
 5 aaacatgigt ctccagtcac tgigaacaaa gctatattaa ggtacatcat tcaaggactt 480  
 25  
 6 catccitca gcacigtiga tcgccaatca lltaaagagc tgattaglac actgcagcct 540  
 30  
 7 ggcatctctg tcattacaag gcttacttta cgtccaaga tagctgaagc tgcctgatac 600  
 35  
 8 atgaaacaga aagtgcctgc tgccatgagt gaagtigaat ggatigcaac cacaacggat 660  
 40  
 9 tgttggactg cacgtagaaa gtcattcatt ggtgtaactg ctactggat caacctgga 720  
 45  
 10 agtcttgaaa gacattccgc tgcactigcc tgcaaaagat taatgggctc tcaactttt 780  
 50  
 11 gaggtactgg ccagtgccat gaatgatata cactcagagt atgaaatagc tgacaaggtt 840  
 55  
 12 gtttgcacaa ccacagacag tggttccaac tttatgaagg ctctcagagt ttttgggtg 900  
 13 gaaaacaatg atatcgagac tgaggcaaga aggttggaaa gtagatgacac tgattctgaa 960  
 14 ggctgtggtg agggaagiga tgggtggaa ttccaagatg cctcagagt cctggaccaa 1020

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5  
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10  
agatctgtct ttggcaaaig ccaagcttla tggaataaaa gcagccgatc ggctctagca 1200  
15  
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25  
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30  
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35  
gcggaacga atacacagct ggggtggctg ctgcctagtg tccatcagtt aagcttgaaa 1500  
40  
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55  
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70  
atttcagcc tctctatcaa gaclaalaca cctcttcccg catcggctgc ctgtgagagg 1920

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5 cttttcagca ctgcaggatt gcttttcagc cccaaaagag ctaggcttga cactaacaat 1980  
 10 ttlgagaatc agcttctact gaagttaaata ctgaggtttt acaactttga gtagcgtgta 2040  
 15 ctggcattag attgctcgtc ttatagtttg ataaitaaat acaaacagtt ctaaagcagg 2100  
 20 ataaaacctt gtaigcattt catttaagtt tttttgagat taaaagctta aacaag 2156  
 25 <210> 2  
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 <212> PRT  
 <213> Oryzias latipes  
 <400> 2  
 30 Met Glu Glu Val Cys Asp Ser Ser Ala Ala Ala Ser Ser Thr Val Gln  
 1 5 10 15  
 35 Asn Gln Pro Gln Asp Gln Glu His Pro Trp Pro Tyr Leu Arg Glu Phe  
 20 25 30  
 40 Phe Ser Leu Ser Gly Val Asn Lys Asp Ser Phe Lys Met Lys Cys Val  
 35 40 45  
 45 Leu Cys Leu Pro Leu Asn Lys Glu Ile Ser Ala Phe Lys Ser Ser Pro  
 50 55 60  
 55 Ser Asn Leu Arg Lys His Ile Glu Arg Met His Pro Asn Tyr Leu Lys  
 65 70 75 80  
 60 Asn Tyr Ser Lys Leu Thr Ala Gln Lys Arg Lys Ile Gly Thr Ser Thr  
 85 90 95  
 65 His Ala Ser Ser Ser Lys Gln Leu Lys Val Asp Ser Val Phe Pro Val  
 100 105 110

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Lys His Val Ser Pro Val Thr Val Asn Lys Ala Ile Leu Arg Tyr Ile  
 115 120 125  
 5 Ile Gln Gly Leu His Pro Phe Ser Thr Val Asp Leu Pro Ser Phe Lys  
 130 135 140  
 10 Glu Leu Ile Ser Thr Leu Gln Pro Gly Ile Ser Val Ile Thr Arg Pro  
 145 150 155 160  
 Thr Leu Arg Ser Lys Ile Ala Glu Ala Ala Leu Ile Met Lys Gln Lys  
 15 165 170 175  
 Val Thr Ala Ala Met Ser Glu Val Glu Trp Ile Ala Thr Thr Thr Asp  
 180 185 190  
 20 Cys Trp Thr Ala Arg Arg Lys Ser Phe Ile Gly Val Thr Ala His Trp  
 195 200 205  
 25 Ile Asn Pro Gly Ser Leu Glu Arg His Ser Ala Ala Leu Ala Cys Lys  
 210 215 220  
 Arg Leu Met Gly Ser His Thr Phe Glu Val Leu Ala Ser Ala Met Asn  
 225 230 235 240  
 30 Asp Ile His Ser Glu Tyr Glu Ile Arg Asp Lys Val Val Cys Thr Thr  
 245 250 255  
 35 Thr Asp Ser Gly Ser Asn Phe Met Lys Ala Phe Arg Val Phe Gly Val  
 260 265 270  
 40 Glu Asn Asn Asp Ile Glu Thr Glu Ala Arg Arg Cys Glu Ser Asp Asp  
 275 280 285  
 Thr Asp Ser Glu Gly Cys Gly Glu Gly Ser Asp Gly Val Glu Phe Gln  
 290 295 300  
 45 Asp Ala Ser Arg Val Leu Asp Gln Asp Asp Gly Phe Glu Phe Gln Leu  
 305 310 315 320  
 50 Pro Lys His Gln Lys Cys Ala Cys His Leu Leu Asn Leu Val Ser Ser  
 325 330 335  
 55 Val Asp Ala Gln Lys Ala Leu Ser Asn Glu His Tyr Lys Lys Leu Tyr

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	340		345		350
5	Arg Ser Val Phe Gly Lys Cys Gln Ala Leu Trp Asn Lys Ser Ser Arg				
	355		360		365
10	Ser Ala Leu Ala Ala Glu Ala Val Glu Ser Glu Ser Arg Leu Gln Leu				
	370		375		380
15	Leu Arg Pro Asn Gln Thr Arg Trp Asn Ser Thr Phe Met Ala Val Asp				
	385		390		395
	Arg Ile Leu Gln Ile Cys Lys Glu Ala Gly Glu Gly Ala Leu Arg Asn				
		405		410	
20	Ile Cys Thr Ser Leu Glu Val Pro Met Phe Asn Pro Ala Glu Met Leu				
		420		425	
	Phe Leu Thr Glu Trp Ala Asn Thr Met Arg Pro Val Ala Lys Val Leu				
25		435		440	
	Asp Ile Leu Gln Ala Glu Thr Asn Thr Gln Leu Gly Trp Leu Leu Pro				
		450		455	
30	Ser Val His Gln Leu Ser Leu Lys Leu Gln Arg Leu His His Ser Leu				
		465		470	
	Arg Tyr Cys Asp Pro Leu Val Asp Ala Leu Gln Gln Gly Ile Gln Thr				
		485		490	
40	Arg Phe Lys His Met Phe Glu Asp Pro Glu Ile Ile Ala Ala Ala Ile				
		500		505	
	Leu Leu Pro Lys Phe Arg Thr Ser Trp Thr Asn Asp Glu Thr Ile Ile				
45		515		520	
	Lys Arg Gly Met Asp Tyr Ile Arg Val His Leu Glu Pro Leu Asp His				
		530		535	
50	Lys Lys Glu Leu Ala Asn Ser Ser Ser Asp Asp Glu Asp Phe Phe Ala				
		545		550	
	Ser Leu Lys Pro Thr Thr His Glu Ala Ser Lys Glu Leu Asp Gly Tyr				
55		565		570	
					575



least one inverted repeat in the forepart of its nucleotide sequence.

5 14. The method according to any one of claims 6-13, wherein the gene which can modify the gene structure has a nucleotide sequence containing at least one inverted repeat as part of cis-element.

15. A method of inserting a foreign gene into a gene of a cell by using the method according to any one of claims 6-14.

10 16. A the method of modifying the function of a cell which is based on gene expression by using the method according to any one of claims 6-14.

17. A cell having been modified in function by the method according to claim 16.

15 18. A plasmid, which contains DNA having a nucleotide sequence that includes at least one inverted repeat in the forepart of its nucleotide sequence.

19. A plasmid according to claim 18, wherein part of DNA of the plasmid is excisable in the presence of the protein according to claim 1 or in the presence of the nucleic acid which can produce the said protein.

20 20. In a method of inserting any DNA into genomic DNA of a vertebrate animal, which is **characterized by** operating the said insertion of DNA by using the transposase activity.

21. The method according to claim 20, wherein a DNA is the Tol2 element having the characters according to claims 13-14.

25 22. The method according to claim 20 or 21, wherein the vertebrate is fish.

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FIG. 1

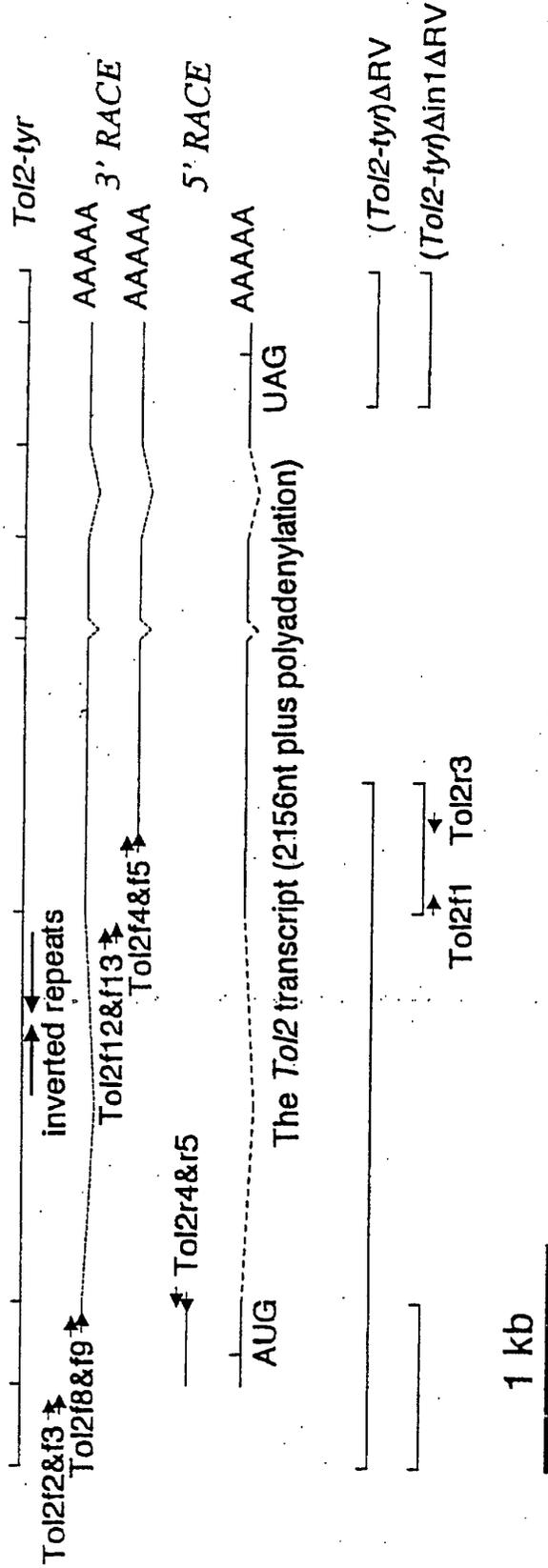


FIG. 2

To12 : 106 VDSVFPVKHVSPTVTKAILRYIIQGLHPFSTVDLPSEKELISTLQPGISVITRPTLRSK 165  
       ++ + P K+ V++ K L II +PF+ V+ F E + +L+P + +R I R  
 Ac : 216 INLIEPYKYDEVVSLKHLHLA-IIIMHEYPFNIVEHEYVEFVKSLRPHFPIKSRVTARKY 273  
  
 To12 : 166 IAEAALIMKQKVTAAAMSEVE-WIATTTDCWTA-RRKSFIGVTAHWIINPG-SLERHSAALA 222  
       I + L K+K+ + +V+ +TT D WT+ + KS++ VT HWI+ L++  
 Ac : 274 IMDLYLEEKEKLYGKLDVQSRFSTTMDMWTSCQNKSYMVYTIHWIDDDWCLOKRIYGVFF 333  
  
 To12 : 223 CKRLMGSHTFEVLA SAMNDIHSEYEIROKYVCTTTDSSNFMKAFRYFGVENNDIETEAR 282  
       + G HT + L+ I ++ I K+ + D+ S N++  
 Ac : 334 ---HVEGRHTGQRLSQFTTAIMVKWNIKKLFALSLDNAS-----ANEVAVHDI 379  
  
 To12 : 283 RCESDDTDSEGGEGSDGYEFQDASRVLDQDDGFEFQLPKHKKCACHLLNLVS 335  
       + DTDS DG F H +CACH+LNLV+  
 Ac : 380 IEDLQDTDSNLV---CDGAFF-----HVRCAHILNLVA 410

FIG. 3

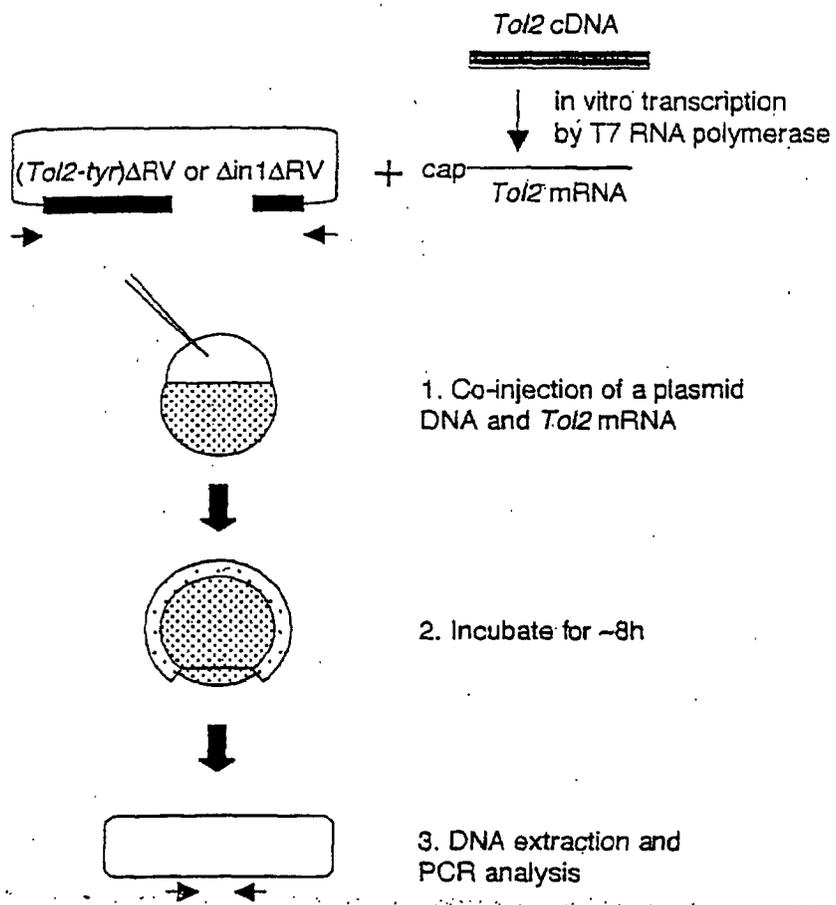


FIG. 4



FIG. 5

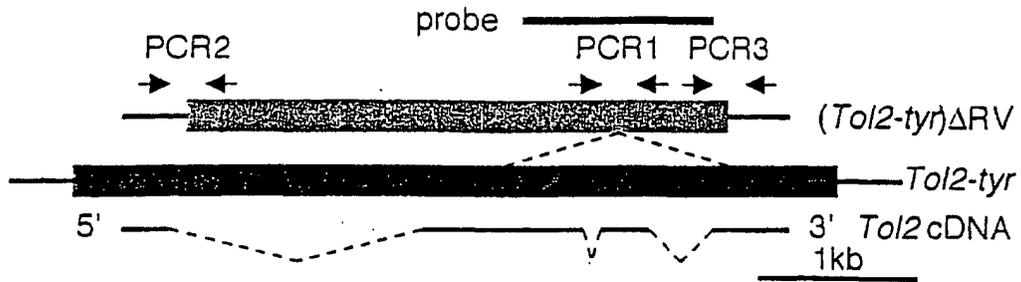


FIG. 6

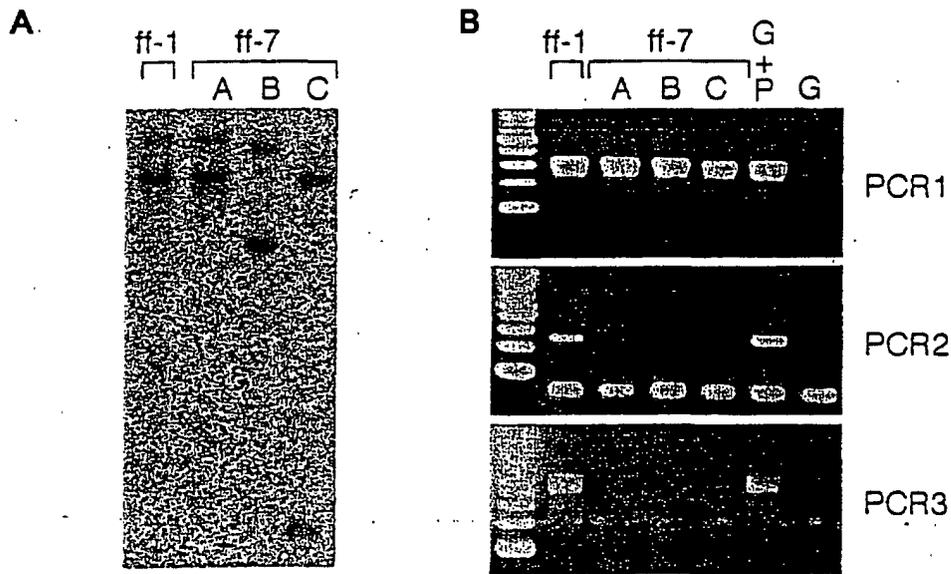


FIG. 7

A GTTTTTTTTTTACATCTCAACTG-*Tol2*-CTCAACTGATAGTCTAATCACAC  
 B CGCTGAGCTCTCTTATATAGAGA-*Tol2*-TATAGAGATGGCTGTTATACGAG  
 C AAGTGACGTC AATGTGTTTCAG-*Tol2*-GTTTTCAGCTCATCTGTTTCATTA

## INTERNATIONAL SEARCH REPORT

International application No.

PCT/JP00/08014

A. CLASSIFICATION OF SUBJECT MATTER Int.Cl. <sup>7</sup> C12N15/54, C12N9/12, C12N15/90, C12N5/10		
According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED		
Minimum documentation searched (classification system followed by classification symbols) Int.Cl. <sup>7</sup> C12N15/54, C12N9/12, C12N15/90, C12N5/10		
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched		
Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) JICST FILE (JOIS), WPI (DIALOG), BIOSIS (DIALOG), MEDLINE (STN), EMBL/DDBJ/Genbank/PIR/Swissprot/Geneseq		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	Gene, Vol.240 [1] (November, 1999), K. Kawakami et al., "Identification of the Tol2 transposase of the medaka fish <i>Oryzias latipes</i> that catalyzes excision of nonautonomous Tol2 element in zebrafish <i>Danio rerio</i> ", pp.239-244	1-9,11,13, 15-19
X Y	Gene, Vol.225 [1/2] (1998), K. Kawasaki et al., "Excision of the Tol2 transposable element of the medaka fish, <i>Oryzias latipes</i> , in zebrafish <i>Danio rerio</i> ", pp.17-22	1-4,18-19 5-17
PX	Genbank, (10 December, 1999), A. Koga et al., Acc. No.D84375: "Oryzias latipes DNA"	1-4
X Y	FEBS Lett., Vol.461 [3] (November, 1999), A Koga et al., "Amino acid sequence of a putative transposase protein of the medaka fish transposable element Tol2 deduced from mRNA nucleotide sequences", pp.295-298	1-4 6-17,19,21-22
X Y	Nature, Vol.383 [6595] (September, 1996), A Koga et al., "Transposase element in fish" p.30	1-4 6-7,19,21-22
X Y	Proc. Natl. Acad. Sci. USA, Vol.95 [3] (1998), L. Guangbin et al., "Chromosomal transposition of a TC1/marine-like	18-20 6-17,21-22
<input checked="" type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/> See patent family annex.		
* Special categories of cited documents:	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention	
"A" document defining the general state of the art which is not considered to be of particular relevance	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone	
"E" earlier document but published on or after the international filing date	"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art	
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"&" document member of the same patent family	
"O" document referring to an oral disclosure, use, exhibition or other means		
"P" document published prior to the international filing date but later than the priority date claimed		
Date of the actual completion of the international search 19 March, 2001 (19.03.01)	Date of mailing of the international search report 03 March, 2001 (03.04.01)	
Name and mailing address of the ISA/ Japanese Patent Office	Authorized officer	
Facsimile No.	Telephone No.	

Form PCT/ISA/210 (second sheet) (July 1992)

## INTERNATIONAL SEARCH REPORT

International application No.

PCT/JP00/08014

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
	element in mouse embryonic stem cells", pp.10769-10773	
X	Nucleic Acids Res., Vol.26[16] (1998), L. Zhang et al.,	18,20
Y	"The <i>Hemerl mariner</i> transposase cloned in a recombinant adenovirus vector is functional in mammalian cells", pp.10769-10773	6-17,19,21-22
X	Nat. Biothechnol., Vol.16[11] (1998), A Sherman et al.,	18-20
Y	"Transposition of the <i>Drosophila</i> element <i>mariner</i> into the chicken germ line", pp.10769-10773	6-17,21-22
X	Somatic Cell Mol. Genet., Vol.24[6] (1998), Z-H. Li et al.,	18-20
Y	"Inversion and transposition of Tc1 Transposon of <i>C. elegans</i> in Mammalian Cells", pp.363-369	6-17,21-22

Form PCT/TSA/210 (continuation of second sheet) (July 1992)

## INTERNATIONAL SEARCH REPORT

International application No.

PCT/JP00/08014

**Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)**

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1.  Claims Nos.:  
because they relate to subject matter not required to be searched by this Authority, namely:
  
2.  Claims Nos.:  
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
  
3.  Claims Nos.:

**Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)**

This International Searching Authority found multiple inventions in this international application, as follows:

The inventions as set forth in claims 1 to 17 relate to transposases having a specific sequence. The inventions as set forth in claims 18 and 19 relate to plasmids having a specific structure. The inventions as set forth in claims 20 to 22 relate to a method of inserting a gene into genome with the use of the activity of a transposase. (Although a prefix "said" is attached to the transposase, it is not clearly stated therein that the transposase has a specific sequence and no other claims are cited therein.)

As clearly stated in the description, transposase has been well known prior to the application of the present case. Accordingly, there is no special technical matter, as specified in Rule 13.2 of the Regulations under the PCT, common to all of the claims. Such being the case, it is recognized that the inventions as set forth in claims 1 to 22 involve three groups of inventions, i.e., the group of the inventions as set forth in claims 1 to 17, the group of the inventions as set forth in claims 18 and 19, and the group of the inventions as set forth in claims 20 to 22.

1.  As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
  
2.  As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
  
3.  As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
  
4.  No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

**Remark on Protest**  The additional search fees were accompanied by the applicant's protest.  
 No protest accompanied the payment of additional search fees.